

## **Supplemental Information**

### **Bispecific BAFF-R/BCMA CAR T cells control growth of heterogeneous plasma cells in multiple myeloma**

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## **Supplemental Material and methods**

### **Construction of bispecific bicistronic chimeric antigen receptors**

Three bicistronic vectors encoding an anti-BAFF-R CAR and an anti-BCMA CAR linked by a P2A element were constructed using codon-optimized, synthesized DNA fragments. The first construct, "anti-BAFF-R-CD8 $\alpha$ -28 $\zeta$  1XX-P2A-anti-BCMA-IgG1mut-28 $\zeta$  1XX" (Bi-8a-1XXz-CAR), encodes both CARs with the -1XX version of the CD3 $\zeta$  activation domain. The anti-BAFF-R CAR features a CD8 $\alpha$  hinge, spacer, and TM domain. The second construct, "anti-BAFF-R-CD8 $\alpha$ -BB $\zeta$  WT-P2A-anti-BCMA-IgG1mut-28 $\zeta$  1XX" (Bi-8a-1Bz-CAR), was derived from the first construct by replacing the intracellular signaling domains of the anti-BAFF-R CAR with the 4-1BB costimulatory domain and the unmodified CD3 $\zeta$  activation domain. The third construct, "anti-BAFF-R-CD28-BB $\zeta$  WT-P2A-anti-BCMA-IgG1mut-28 $\zeta$  1XX" (Bi-28-1BBz-CAR), was generated by substituting the CD8 $\alpha$  spacer and TM domains with CD28 spacer and TM domains in the anti-BAFF-R CAR. Gene synthesis all performed by GeneArt (Thermo Fisher).

**Table S1. Frequency distribution of BAFF-R expression in MM bone marrow specimen**

BAFF-R expression in CD138<sup>+</sup> cells was evaluated on BM biopsies and extramedullary manifestations from 58 MM cases using immunohistochemistry. Specimens are classified as newly diagnosed MM (NDMM), post-treatment and extramedullary. Number of BAFF-R<sup>+</sup> and BAFF-R<sup>-</sup> specimens are reported.

<b>Clinical cases</b>	<b>n=58</b>	<b>BAFF-R<sup>-</sup></b>	<b>BAFF-R<sup>+</sup></b>
<b>NDMM</b>	<b>n=20</b>	<b>9</b>	<b>11</b>
<b>Post-treatment</b>	<b>n=30</b>	<b>17</b>	<b>13</b>
<b>Extra-medullary</b>	<b>n=8</b>	<b>8</b>	<b>0</b>

**Table S2. Antibody staining panel for the detection of plasma cells and B cells**

<b>Antibody</b>	<b>Fluorochrome</b>	<b>Clone</b>	<b>Isotype</b>
BCMA (CD269)	BV421	19F2	mouse IgG2a, k
CD19	BV510	HIB19	mouse IgG1, k
CD38	FITC	HIT2	mouse IgG1, k
BAFF-R (CD268)	PE	11C1	mouse IgG1, k
CD56	PE-Cy7	5.1H11	mouse IgG1, k
CD138	APC	DL-101	mouse IgG1, k
CD3	APC-Cy7	HIT3a	mouse IgG2a, k

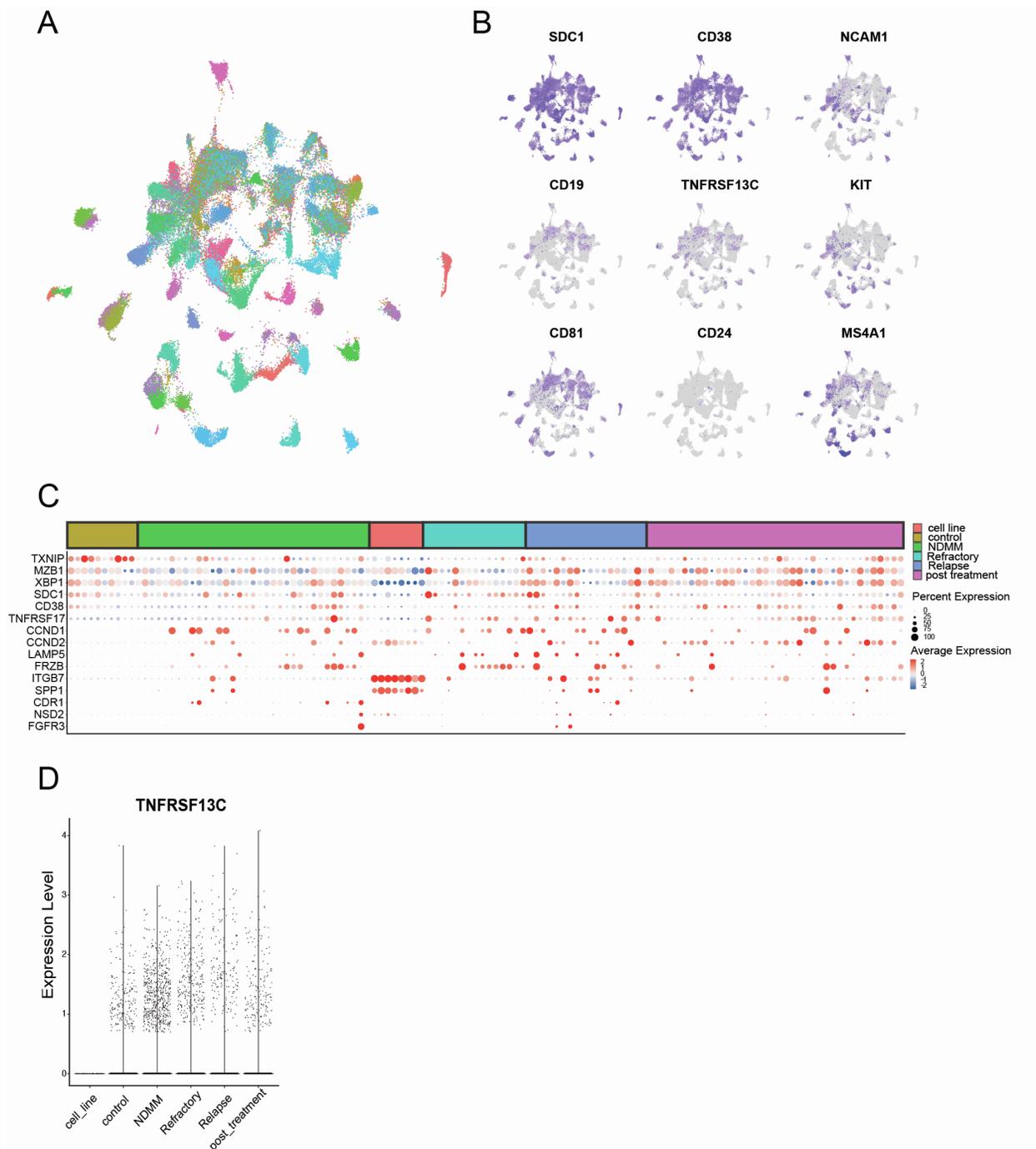
**Table S3. Antibody panel for the detection and discrimination of CAR T cells and target cells**

<b>Antibody</b>	<b>Fluorochrome</b>	<b>Clone</b>	<b>Isotype</b>
CD19	BV510	HIB19	mouse IgG1, k
CD38	FITC	HIT2	mouse IgG1, k
CD56	PE-Cy7	5.1H11	mouse IgG1, k
CD138	APC	DL-101	mouse IgG1, k
CD3	APC-Cy7	HIT3a	mouse IgG2a, k
	7-AAD		
	eFluor670		

**Table S4. Antibody panel for the analysis of T cells and MM.1s tumor cells**

*in vivo*

<b>T cell panel</b>			
<b>Antibody</b>	<b>Fluorochrome</b>	<b>Clone</b>	<b>Isotype</b>
PD-1	PE	EH12.2H7	mouse IgG1, k
LAG-3	AF647	11C3C65	mouse IgG1, k
TIM-3	BV421	F38-2E2	mouse IgG1, k
CD8	PE-Cy7	HIT8a	mouse IgG1, k
CD3	APC-Cy7	HIT3a	mouse IgG2a, k
<b>CAR T cell panel</b>			
<b>Antibody</b>	<b>Fluorochrome</b>	<b>Clone</b>	<b>Isotype</b>
CD8	PE-Cy7	HIT8a	mouse IgG1, k
CD3	APC-Cy7	HIT3a	mouse IgG2a, k
Goat anti human IgG	PE	Polyclonal	FMO
<b>Tumor cell panel</b>			
<b>Antibody</b>	<b>Fluorochrome</b>	<b>Clone</b>	<b>Isotype</b>
CD138	BV421/APC	DL-101	mouse IgG1, k
BCMA (first antibody, unlabeled)		J22.9-FSY, human recombinant	
BAFF-R	PE/APC	11C1	mouse IgG1, k
Goat anti-human IgG (secondary antibody)	PE	Polyclonal	



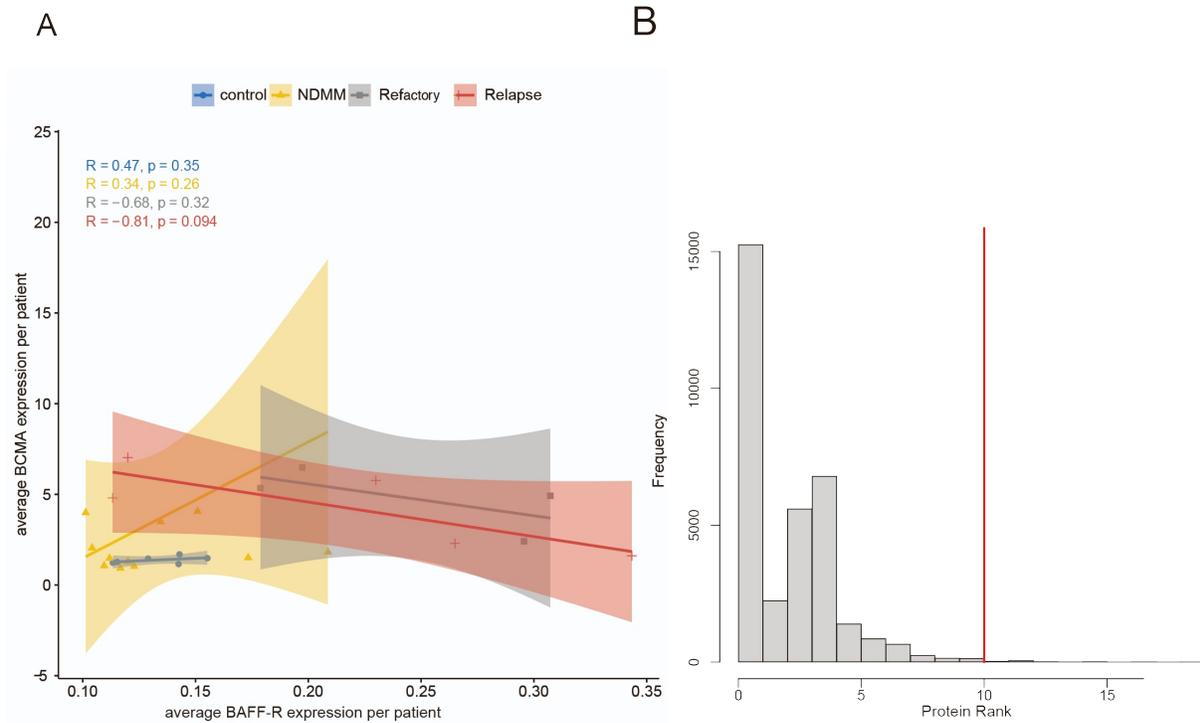
**Figure S1. scRNAseq reveals the heterogeneity of Multiple Myeloma (MM) and the proportion of *TNFRSF13C*-positive malignant plasma cells in disease stages**

**A)** UMAP plot of primary MM patient samples, showing a high heterogeneity among patients, while patients before and after treatment cluster together.

**B)** UMAP visualization of expression distribution of MM-associated genes in all primary MM patients. *TNFRSF13C* gene expression compared to hematopoietic stem cell markers and other MM-associated genes.

**C)** Enrichment of MM-signature genes in disease subgroups. MM cell lines exhibit a distinctive expression profile. Dot plot representing the size-coded percentage of MM cases expressing selected MM signature genes. Expression changes are depicted according to the color scale.

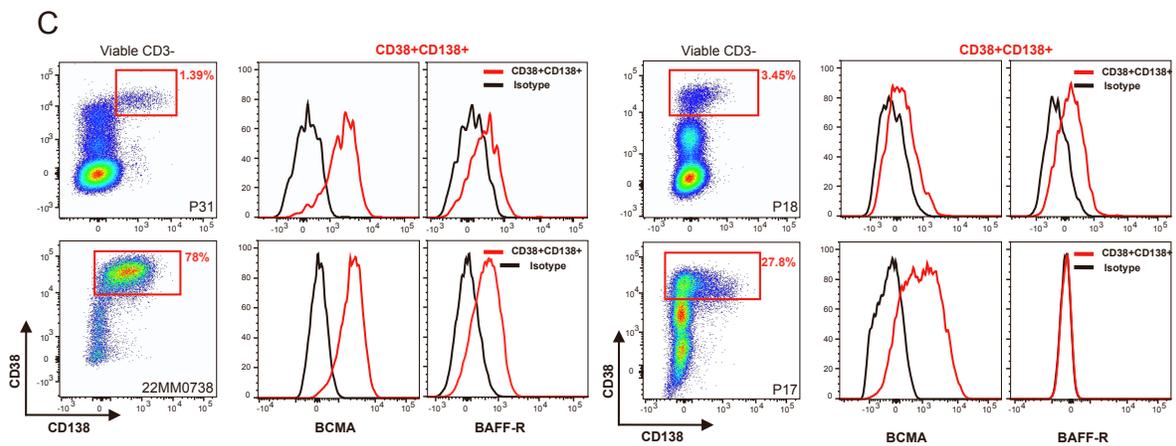
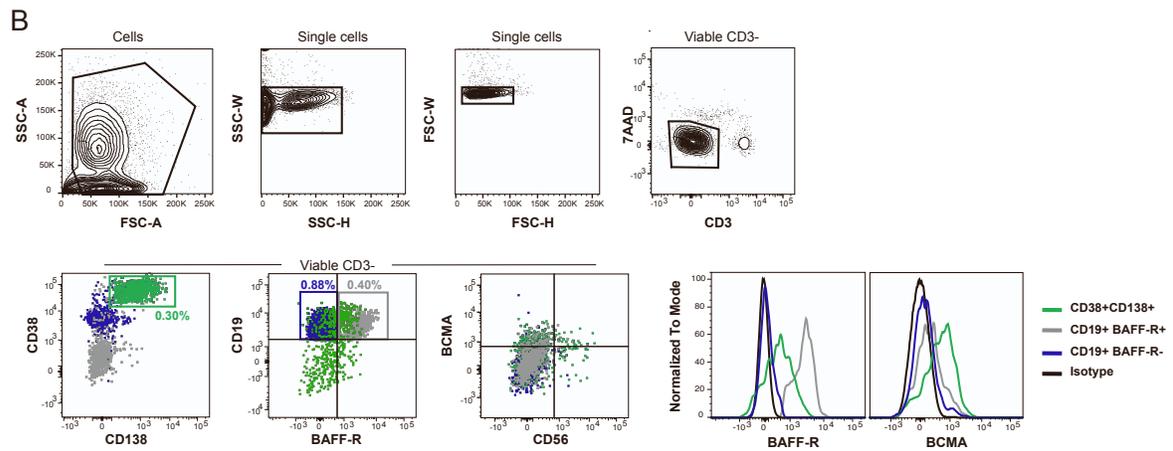
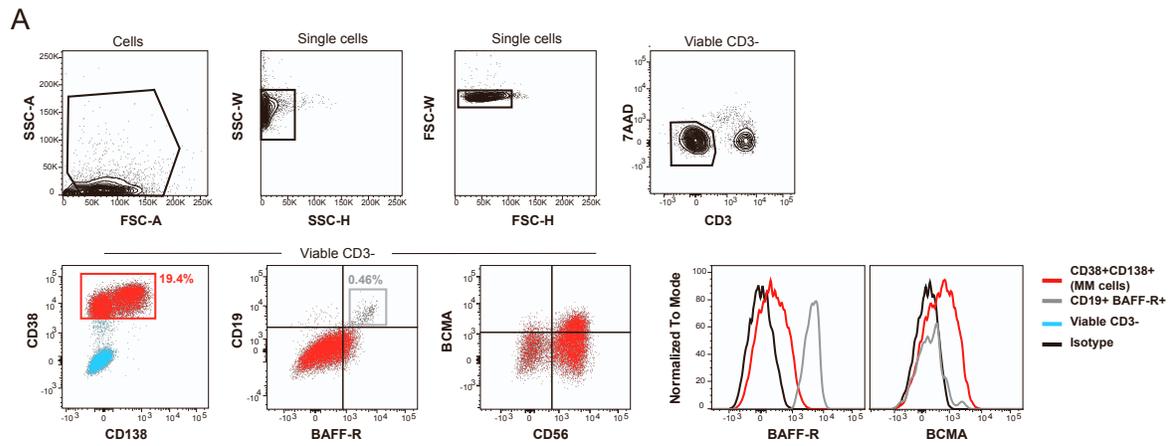
**D)** *TNFRSF13C* (BAFF-R) is not expressed in MM cell lines, but in primary tumor cells from all disease stages.



**Figure S2. Correlation analysis of BCMA and BAFF-R expression in Multiple Myeloma patients**

**A)** Correlation analysis of scRNAseq data, as in **Figure 1**. In cases of increased BAFF-R (*TNFRSF13C*) expression, control and NDMM patients show a positive correlation between BAFF-R and BCMA (*TNFRSF17*). In R/R patients, BCMA negatively correlates with an increasing BAFF-R expression.

**B)** Analysis of surfaceome data from Reference 43. In a ranking of potential Multiple Myeloma target antigens, BAFF-R is found in the top 1% by a combined evaluation based on gene expression and protein data. The red line indicates the rank of BAFF-R.

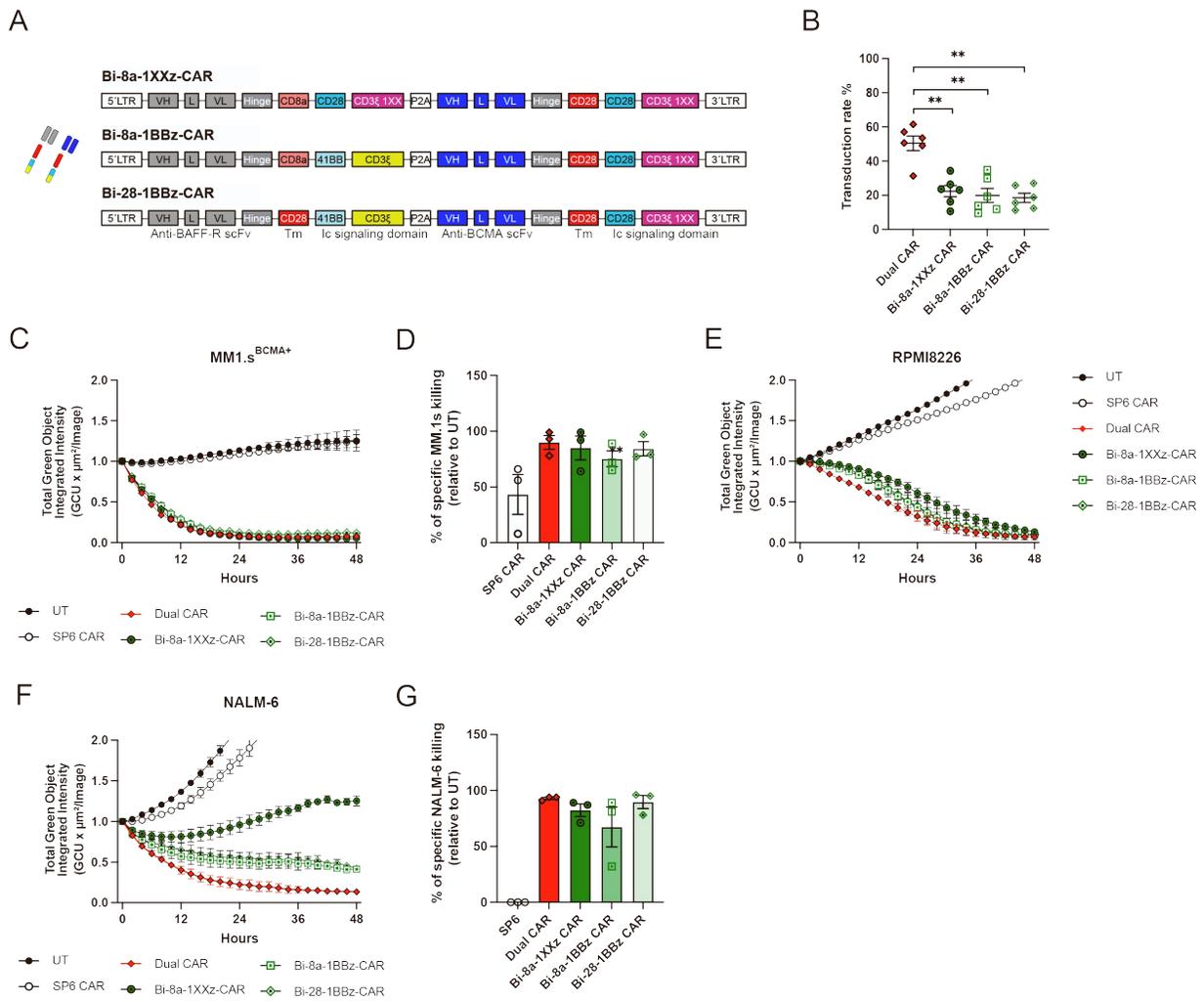


**Figure S3. Flow cytometry detection of BAFF-R<sup>+</sup> expression can differentiate between PCs and B cells**

**A)** Representative gating strategy for the characterization of malignant PCs in BM-derived MM samples. The cell population was first gated for singlets and then viable CD3<sup>-</sup> cells (light blue) are selected. Frequency of the gated populations within viable CD3<sup>-</sup> cells is reported. Histograms represent BAFF-R and BCMA in color-coded defined subpopulations. Malignant PCs are identified as CD38<sup>+</sup>CD138<sup>+</sup> cells (red) and BAFF-R<sup>+</sup> B cells as CD19<sup>+</sup>BAFF-R<sup>+</sup> (grey), respectively. Isotype control, black open histogram.

**B)** Representative gating strategy for the characterization of healthy BM specimen. The cell population was first gated for singlets and then viable CD3<sup>-</sup> cells were selected. Among those the following cell populations were defined: CD38<sup>+</sup>CD138<sup>+</sup> (green), CD19<sup>+</sup>BAFF-R<sup>-</sup> (blue), and CD19<sup>+</sup>BAFF-R<sup>+</sup> (grey). Histograms represent BCMA and BAFF-R expression in the aforementioned populations. Isotype control, black open histogram.

**C)** Representative gating strategy for the detection of malignant PCs within four individual BM-derived MM samples (#P31, #22MM0738, #P18 and #P17). Malignant PCs are identified as CD38<sup>+</sup>CD138<sup>+</sup> cells (red boxed gate) and their frequency among the CD3<sup>-</sup> viable population is reported. Histograms represent BCMA and BAFF-R expression in the gated population. Isotype control, black open histogram.



**Figure S4. Bicistronic CAR designs are impaired in retroviral transduction rates, but endow T cells with efficient cytolytic capacity**

**A)** Schematic representation of the bicistronic BAFF-R/BCMA CAR constructs Bi-8a-1XXz-CAR, Bi-8a-1BBz-CAR and Bi-28-1BBz-CAR. VH variable heavy chain, L Whitlow linker, VL variable light chain, Tm transmembrane region, Ic intracellular signalling domain, LTR long terminal repeat, Hinge human IgG1 Ch2CH3 constant region, P2A element.

**B)** Transduction rates in percent of T cells, data are mean  $\pm$  SEM, n= 6 independent T cell donors transduced with retroviruses encoding: dual CAR, Bi-8a-1XXz-CAR, Bi-8a-1BBz-CAR and Bi-28-1BBz-CAR.

**C)** IncuCyte assay displaying killing kinetic of dual and bicistronic CAR T cells in coculture with GFP<sup>+</sup> MM1.s<sup>BCMA<sup>+</sup></sup>. Data are mean values ± SEM. UT n=3, SP6 CAR n=3; Dual CAR of n=2, Bi-8a-1XXz-CAR n=1, Bi-8a-1BBz-CAR n=2 and Bi-28-1BBz-CAR n=2 CAR T cell donors.

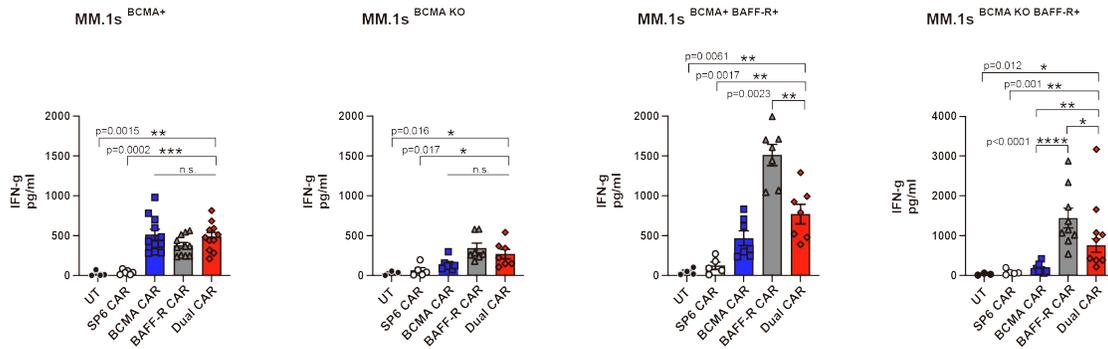
**D)** Flow cytometry-based cytotoxicity assay of CAR T cells and MM.1s<sup>BCMA<sup>+</sup></sup> in a 1:1 E:T coculture. Specific MM.1s<sup>BCMA<sup>+</sup></sup> cell killing is reported in percentage and it is calculated relative to coculture with untransduced (UT) T cells; n=3 CAR T cell donors. Mean ± SEM.

**E)** IncuCyte assay over 48 hours displaying killing kinetic of dual and bicistronic CAR T cells in coculture with GFP<sup>+</sup> RPMI-8226 MM cell line (BCMA<sup>+</sup> BAFF-R<sup>-</sup>). Mean ± SEM. UT n=2, SP6 CAR n=2; Dual CAR of n=2, Bi-8a-1XXz-CAR n=2, Bi-8a-1BBz-CAR n=2 and Bi-28-1BBz-CAR n=2 CAR T cell donors.

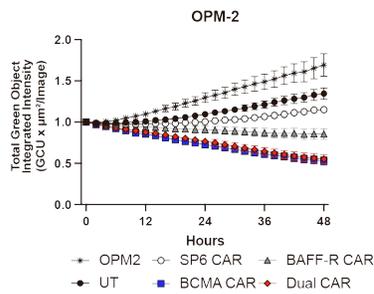
**F)** Killing kinetic of dual and bicistronic CAR T cells in coculture with GFP<sup>+</sup> NALM-6 cells (BAFF-R<sup>+</sup> BCMA<sup>-</sup>). Data are Mean ± SEM. UT n=2, SP6 CAR n=2; Dual CAR of n=2, Bi-8a-1XXz-CAR n=2, Bi-8a-1BBz-CAR n=2 and Bi-28-1BBz-CAR n=2 CAR T cell donors.

**G)** Flow cytometry-based cytotoxicity assay of CAR T cells and NALM-6 in 1:1 coculture. Specific NALM-6 cell killing is reported in percentage and it is calculated relative to coculture with untransduced (UT) T cells. Data are mean ± SEM. In **B**, **D**) and **G**), statistical significance determined by Mann-Whitney test, data points represent single CAR T cell donors. \*\*, p<0.01.

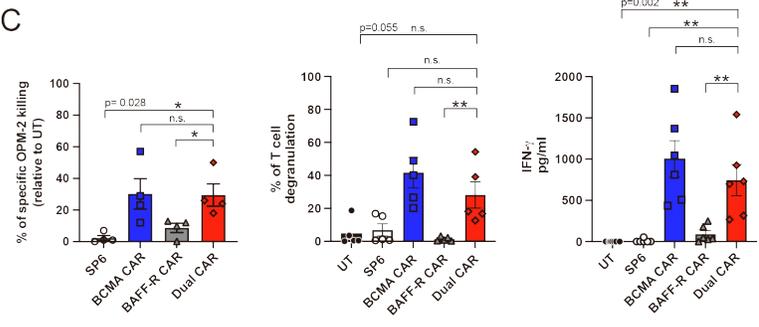
A



B



C

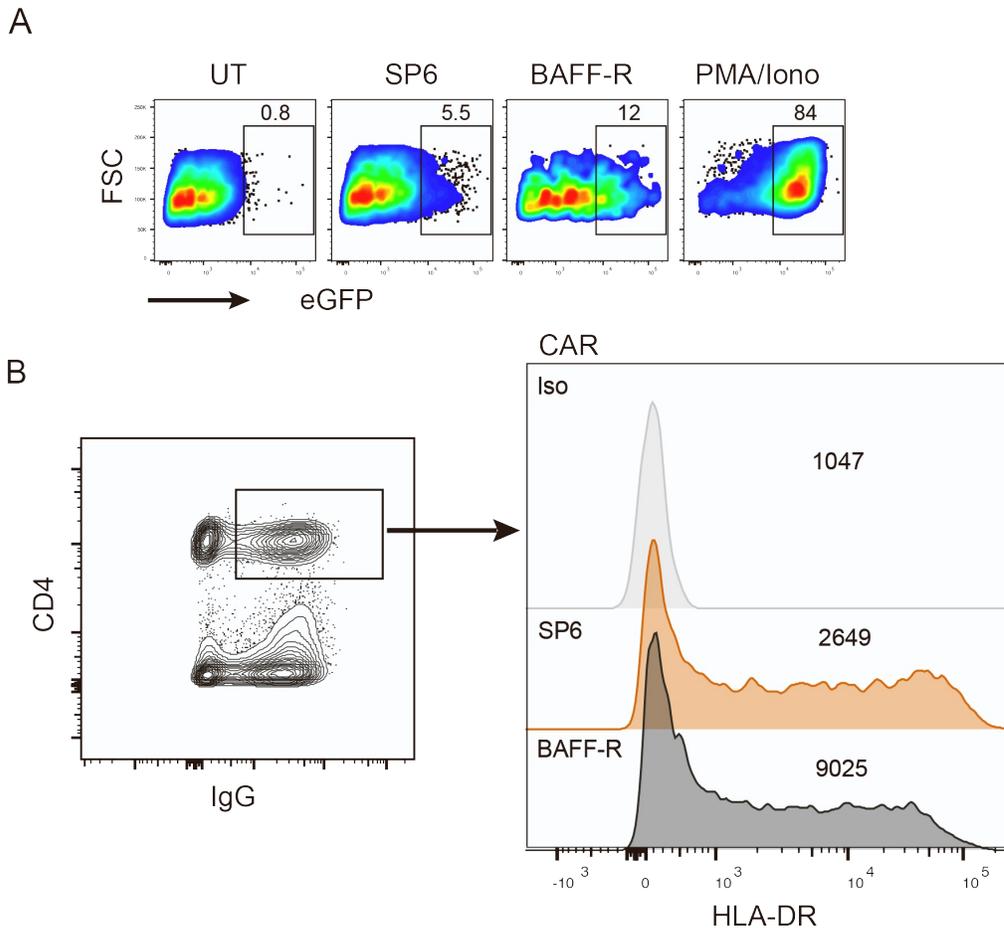


**Figure S5. Dual CAR T cells exhibit similar cytolytic potency against OPM-2 MM cells as monotargeted BCMA CAR T cells**

**A)** IFN- $\gamma$  quantification in supernatants of CAR T and MM.1s cocultures after 18 hours. Mean  $\pm$  SEM. Significance determined by Mann-Whitney test, data points represent single CAR T cell donors.

**B)** IncuCyte assay displaying killing kinetic of CAR T cells in coculture with GFP<sup>+</sup> OPM-2 MM cell line (BCMA<sup>+</sup> BAFF-R<sup>-</sup>). Mean  $\pm$  SEM. UT n=19, SP6 CAR n=19; BCMA CAR n=17, BAFF-R CAR n=17, Dual CAR of n=19 T cell donors; n=14 independent assays performed.

**C)** Left panel: flow cytometry-based cytotoxicity assay of CAR T cells and OPM-2 in 1:1 coculture. Specific OPM-2 cell killing is reported in percentage and it is calculated relative to coculture with untransduced (UT) T cells. Data are Mean  $\pm$  SEM. Middle: percentage of CAR T cell degranulation after 4 hours of coculture with OPM-2. Mean  $\pm$  SEM. Right: IFN- $\gamma$  quantification in supernatants of CAR T and OPM-2 coculture after 18 hours. Mean  $\pm$  SEM, a Mann-Whitney test was applied, data points represent single CAR T cell donors. n.s., non significant.

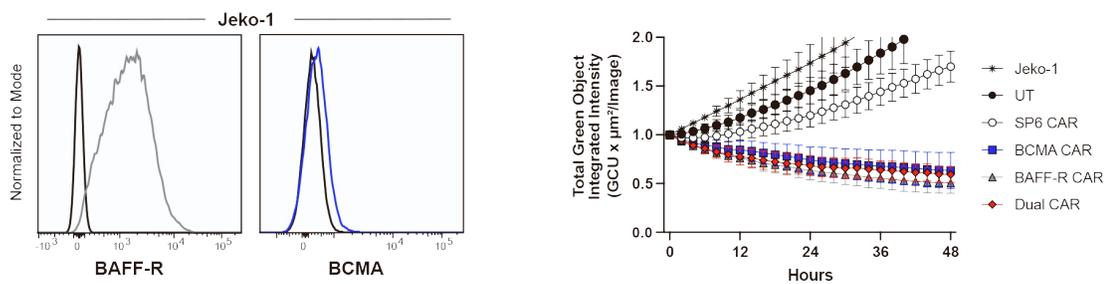


**Figure S6. CAR-induced activation of an NF-KB reporter gene in Jurkat cells**

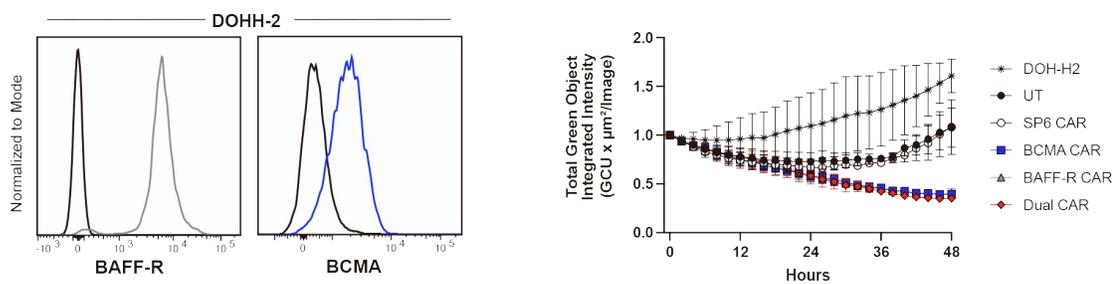
**A)** NF-KB -eGFP Jurkat reporter cells were transduced with the CAR constructs as indicated; UT, untransduced. Ten days after retroviral transduction, CAR<sup>+</sup> Jurkat cells were analyzed for GFP expression by FACS; PMA/ionomycin used as a positive control for activation. Numbers on the gates indicate the frequency in percent of GFP-positive reporter cells. n=2 independent experiments.

**B)** Primary human T cells were transduced with the CAR constructs as indicated. At day ten, CAR<sup>+</sup> CD4<sup>+</sup> T cells were analyzed for HLA-DR (LN3 antibody) expression; gMFI values are given in the histograms. Iso, isotype control. n=2 independent experiments.

A



B

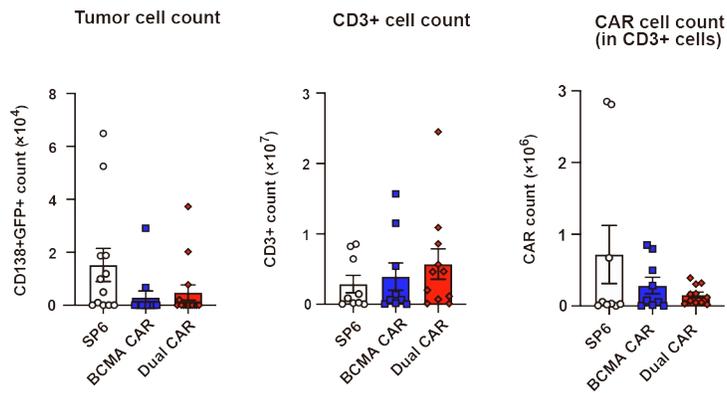


**Figure S7. BCMA<sup>low</sup> expressing B-NHL cell lines are efficiently targeted by dual CAR T cells**

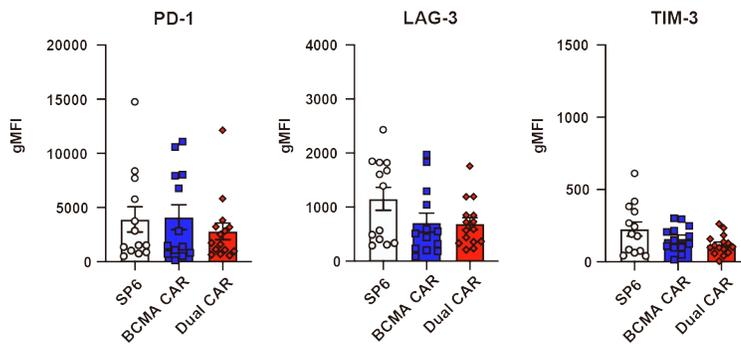
**A)** Flow cytometry analysis of JeKo-1 mantle cell lymphoma cell line stained with anti-BAFF-R and anti-BCMA antibodies. Isotype control, black line; blue line, BCMA expression. On the right, IncuCyte assay for CAR T cells in coculture with GFP<sup>+</sup> JeKo-1 B-NHL cell line (BCMA<sup>+</sup> BAFF-R<sup>+</sup>). Mean  $\pm$  SEM. UT n=7, SP6 CAR n=7; BCMA CAR n=7, BAFF-R CAR n=7, Dual CAR of n=7 T cell donors; n=2 independent experiments performed.

**B)** On the left, FACS histograms of antibody stained DOHH-2 cell line (immunoblastic lymphoma, DLBCL). Right: IncuCyte assay for CAR T cells in coculture with GFP<sup>+</sup> DOHH-2 cell line (BCMA<sup>+</sup> BAFF-R<sup>+</sup>). Mean  $\pm$  SEM. UT n=4, SP6 CAR n=4; BCMA CAR n=4 BAFF-R CAR n=4, Dual CAR of n=4 T cell donors; DOHH-2 n=2.

A



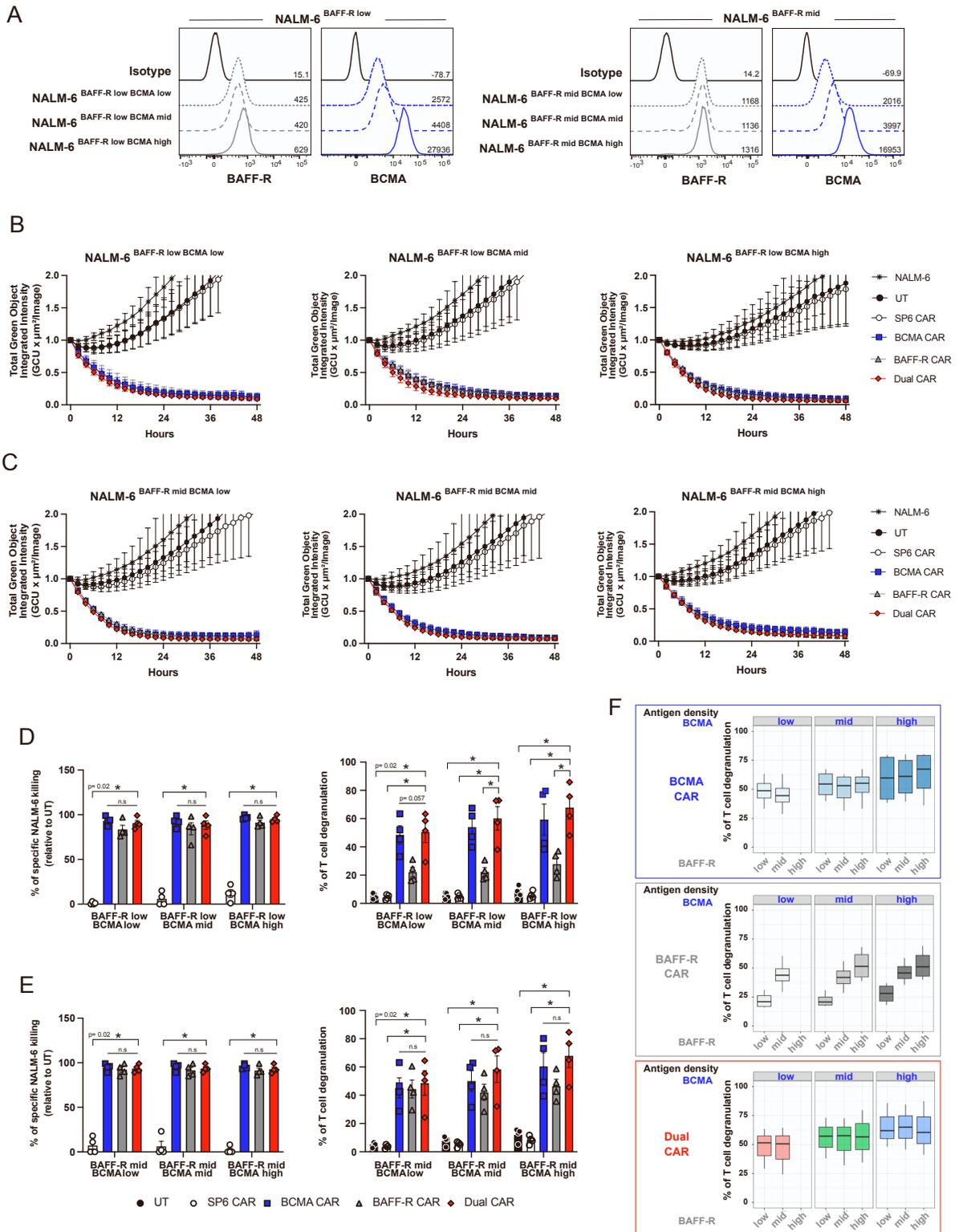
B



**Figure S8. Efficient and comparable depletion of MM.1s tumor cell count in spleen of BCMA CAR and dual CAR T cell treated NSG mice**

**A)** Counts of tumor cells (CD138<sup>+</sup>GFP<sup>+</sup> cells), CD3<sup>+</sup> T cells and CAR T cells in spleen.

**B)** gMFI of PD-1, LAG-3 and TIM-3 exhaustion markers in CD3 cells in spleen. In **A)** and **B)**, mean values ± SEM, n= 3 independent experiments, data points represent individual animals.



**Figure S9. Antigen specificity of dual CAR T cells**

**A)** BAFF-R (grey) and BCMA (blue) antigen expression on NALM-6 cells with titrated amounts of BAFF-R and BCMA. NALM-6 cells were first silenced for BAFF-R expression, and then sequentially transduced for BAFF-R and BCMA. Cells with distinct expression levels were sorted by flow cytometry. For BCMA detection, antibody clone 19F2 was used. gMFI values for the tested antigen are reported. Isotype controls: black open histogram. Representative plots.

**B) and C)** IncuCyte assays of CAR T cells in coculture with GFP<sup>+</sup> NALM-6 cells with various expression levels low-mid-high for BAFF-R and BCMA, as indicated. Mean  $\pm$  SEM. UT, SP6 CAR, BCMA CAR, BAFF-R CAR, Dual CAR n=5 T cell donors; n=7 independent experiments performed.

**D) and E)** Left panels: FACS-based cytotoxicity assay of CAR T cells and NALM-6 with various BAFF-R and BCMA expression levels in a 1:1 coculture. NALM-6 cell killing is reported in percentage, calculated relative to UT T cells.

Middle: percentage of CAR T cell degranulation after 4 hours of coculture with NALM-6.

Mean  $\pm$  SEM with p-value in **D) and E)** by Mann-Whitney test, data points represent single CAR T cell donors. \*, p<0.05; n.s., non significant.

**F)** Quantification of antigen-dependent degranulation capacity of either BCMA CAR (left), BAFF-R CAR (middle), and dual CAR (right panel). On the top of each box and whisker plot, BCMA expression levels are given. On the bottom, corresponding antigen densities for BAFF-R are depicted. Regardless of BCMA antigen density, reactivity of the BAFF-R CAR correlates with increasing BAFF-R antigen expression.