- 1 Supplementary Information for "Lipid-facilitated opening of the ADAM10 sheddase
- 2 revealed by enhanced sampling simulations"

- 4 Adrien Schahl^{1,*}, Nandan Haloi¹, Marta Carroni², Shengpan Zhang^{3,4}, Quentin James
- 5 Sattentau^{4,5}, Erdinc Sezgin^{6,*}, Lucie Delemotte^{1,*}, Rebecca J Howard^{2,*}

6

7

- 8 Science for Life Laboratory, Department of Applied Physics, KTH Royal Institute of Technology, 12121
- 9 Solna, Stockholm, Stockholm County 11428, Sweden
- 10 ² Science for Life Laboratory, Department of Biochemistry and Biophysics, Stockholm University, Solna,
- 11 SE-171 65, Sweden
- ³ The Kennedy Institute of Rheumatology, University of Oxford, Roosevelt Drive, Oxford OX3 7FY, UK.
- ⁴ Sir William Dunn School of Pathology, University of Oxford, Oxford, OX1 3RE, UK.
- 14 ⁵ The Max Delbrück Centre for Molecular Medicine, Campus Berlin-Buch, 13125 Berlin, Germany.
- 15 ⁶ Science for Life Laboratory, Department of Women's and Children's Health, Karolinska Institutet,
- 16 Tomtebodavägen 23, 17165 Solna, Sweden

17

18

- * Correspondence
- 19 adrien.schahl@scilifelab.se
- 20 erdinc.sezgin@ki.se
- 21 lucie.delemotte@scilifelab.se
- 22 <u>rebecca.howard@scilifelab.se</u>

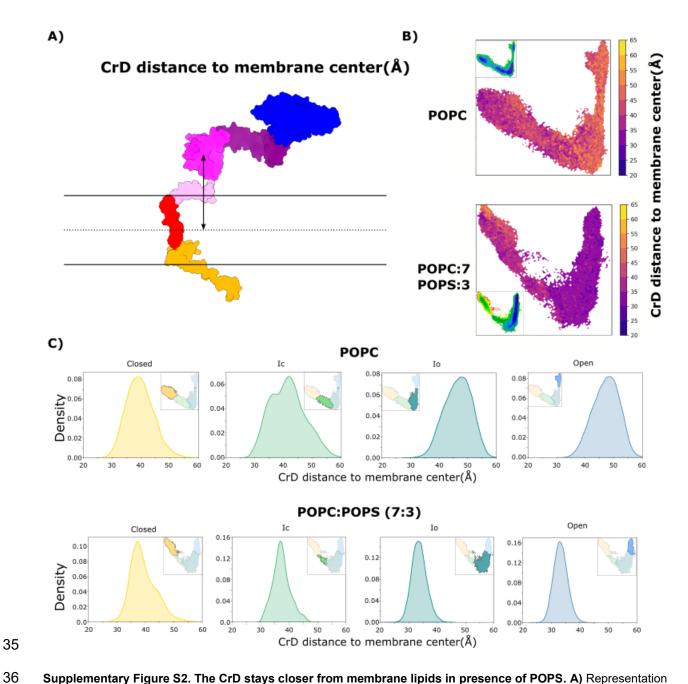
23

FAST features "Tspan" features

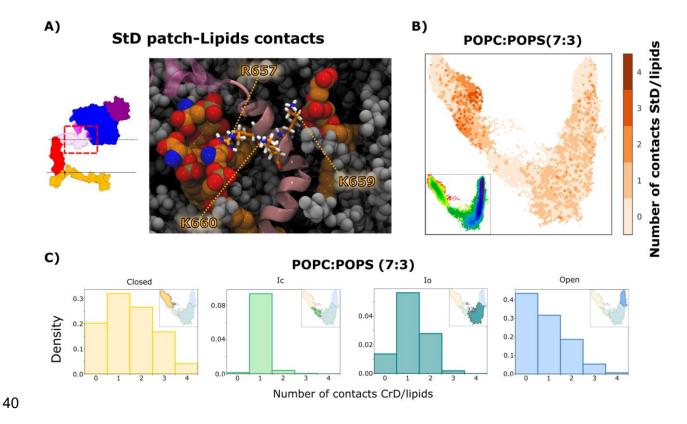
Supplementary Figure S1. Features used for adaptive sampling and MSM building. Left: Residues of the MpD (Cyan), CrD/StD (Red), used to define interdomain distances to select seeds for successive generations of FAST sampling. Right: Residues of the MpD (Cyan), CrD/StD (Red), added as features for the construction of MSM. The numerotation of residues is displayed in Supplementary Table ST1.

Supplementary Table ST1. Features numerotation used for adaptive sampling and MSM building Numerotation of residues involved in the definition of pairwise distances, colored according to Supplementary Figure S1

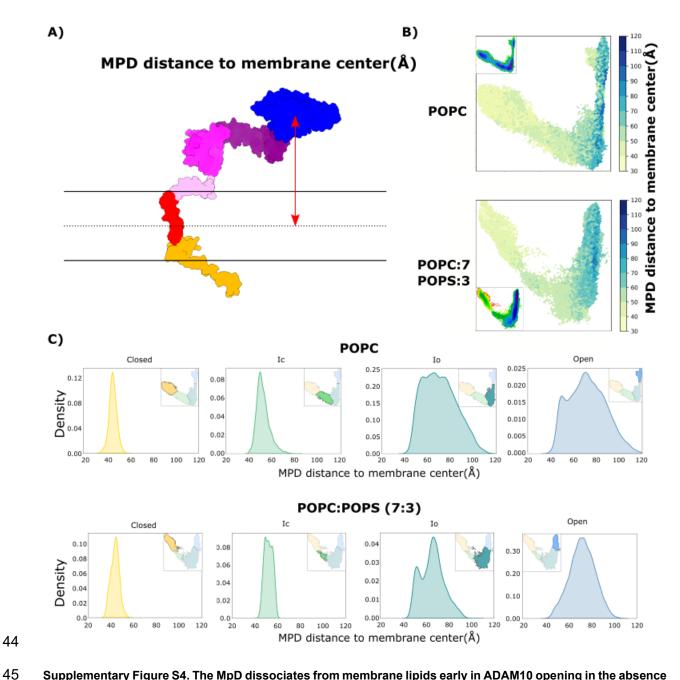
Features	CrD/StD residues number	MPD residues number
FAST sampling	564-570	374-376; 424-426
	601-604	407-412
	630-636	401-405; 413-414
	646-651	419-421
Tspan chelation	556-557;	248-249; 251;
	627-631; 635;	375; 378;
	638-642; 646;	407; 410-411;
	648-652	425; 430



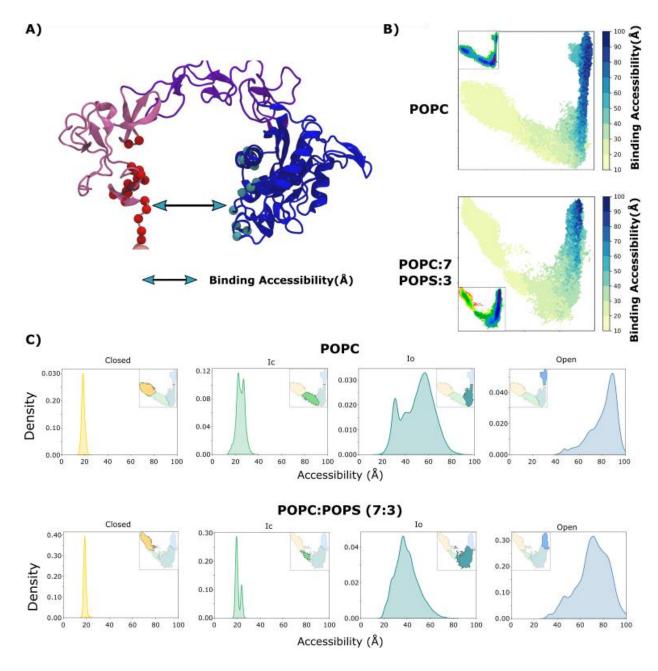
Supplementary Figure S2. The CrD stays closer from membrane lipids in presence of POPS. A) Representation of the distance between the CrD and the center of the membrane **B)** Projection of this distance over free energy maps in POPC and POPC-POPS simulations. **C)** Distribution of this distance in each macro state obtained in both conditions.



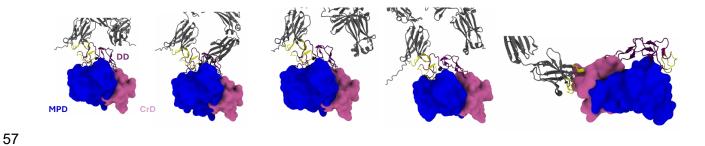
Supplementary Figure S3. POPS interactions with basic residues in the ADAM10 StD. A) Representation of the contacts between POPS lipids and basic residues of the StD **B)** Projection of the number of contacts over free energy maps in POPC-POPS simulations. **C)** Distribution of the number of contacts in each macro state.



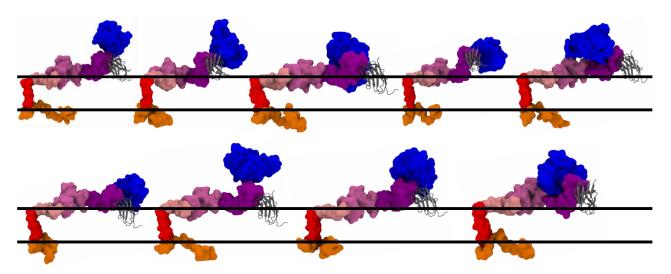
Supplementary Figure S4. The MpD dissociates from membrane lipids early in ADAM10 opening in the absence or presence of POPS. A) Representation of the distance between the MpD and the center of the membrane B) Projection of this distance over free energy maps in POPC and POPC-POPS simulations. C) Distribution of this distance in each macro state obtained in both conditions.



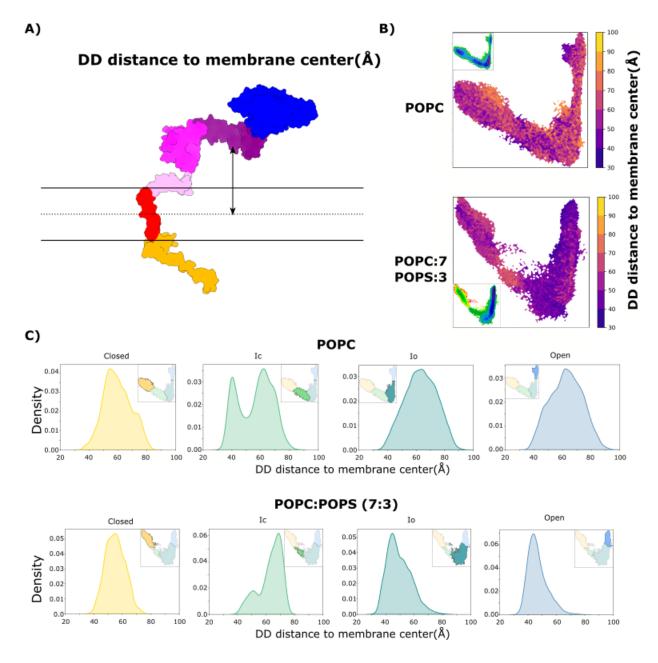
Supplementary Figure S5. Structural accessibility of ADAM10 to Tspan binding. A) Representation of the distance defining the accessibility to Tspan **B)** Projection of this distance over free energy maps in POPC and POPC-POPS simulations. **C)** Distribution of this distance in each macro state obtained in both conditions.



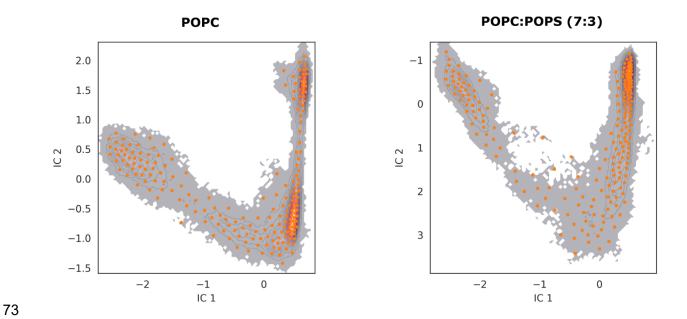
Supplementary Figure S6: Outputs produced by AF3 when predicting antibody binding to ADAM10. In 4 of 5 structures generated with AF3, residues loops 48-54, 71-77 and 118-123 from the Fab (in Yellow) interact with the residues 470-498 (DD, in yellow) of ADAM10. In the last predicted model, we observed an interaction between loops 48-54, 71-77 and 118- 123 from the Fab and residues 576-617 (CrD) from ADAM10.



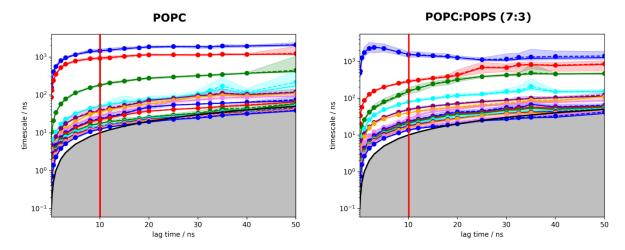
Supplementary Figure S7: Projection of antibody binding to ADAM10 in most represented state of POPC-POPS condition. The structure were extracted from the most populated state of POPC:POPS(7:3) simulation using the "sample_by_distributions" routine of pyEmma. The Fab_{cryo} structure was then superimposed with its corresponding binding site on ADAM10 in each of these structures. For clarity, lipids are represented with black lines.



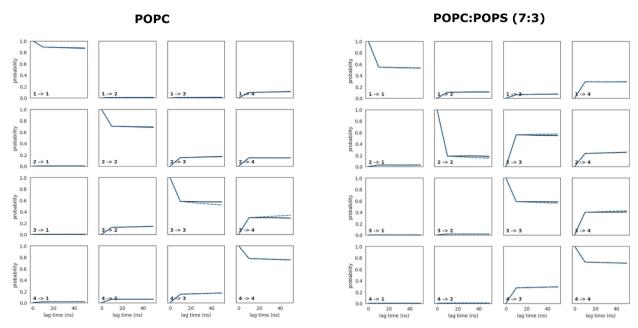
Supplementary Figure S8. The DD stays closer from membrane lipids in presence of POPS. A) Representation of the distance between the DD and the center of the membrane **B)** Projection of this distance over free energy maps in POPC and POPC-POPS simulations. **C)** Distribution of this distance in each macro state obtained in both conditions.



Supplementary Figure S9. Distribution of cluster center obtained after k-means clustering. 200 microstates were assigned in both POPC and POPC:POPS(7:3) conditions.



Supplementary Figure S10. Assessment of convergence of the MSMs with implied timescales. Implied timescales with MSMs estimated with different lag times for POPC and POPC:POPS(7:3) systems. Red lines correspond to the chosen lag times for analyses.



Supplementary Figure S11. Assessment of convergence of the POPC and POPC:POPS(7:3) MSM with Chapman Kolmogorov tests. Chapman-Kolmogorov tests with four states for both systems.