

**Supplemental information**

**Efficient CRISPR-Cas9-mediated  
mutagenesis in primary human B cells  
for identifying plasma cell regulators**

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**Table S1: Amount of each plasmid in plasmid mixtures**

Ratio (Main/Gag- Pol/Envelope)	Mol (picomole) <sup>1</sup>			DNA mass (μg) <sup>2</sup>			
	Main	Gag-Pol	Envelope	Main	Gag-Pol	Envelope	Total
3:1:1	0.16742	0.05581	0.05581	0.76	0.42	0.32	1.50
6:1:1	0.22262	0.03710	0.03710	1.00	0.28	0.22	1.50
12:1:1	0.26657	0.02221	0.02221	1.20	0.17	0.13	1.50
24:1:1	0.29577	0.01232	0.01232	1.33	0.10	0.07	1.50
48:1:1	0.31291	0.00652	0.00652	1.41	0.05	0.04	1.50
96:1:1	0.32224	0.00336	0.00336	1.45	0.03	0.02	1.50
192:1:1	0.32712	0.00170	0.00170	1.477	0.013	0.010	1.50
Only sgRNA (Main plasmid)	0.33215	0	0	1.50	0	0	1.50

Plasmid sizes (base pairs): Main (7,308), Gag-Pol (12,227), Envelope (9,345)

<sup>1</sup> Picomole calculation of each plasmid in total of 1.5 μg DNA:

$$P \text{ (picomole of } M_xG_yE_z) = \frac{1.5 (\mu\text{g}) * 10^6}{(x*a + y*b + z*c) * 617.96 + 36.04}$$

Picomole of Main plasmid = P\*x

Picomole of Gag-Pol plasmid = P\*y

Picomole of Envelope plasmid = P\*z

$M_xG_yE_z$ : Formula of plasmid mixture

M: Main plasmid

G: Gag-Pol plasmid

E: Envelope plasmid

x, y, z: Plasmid ratios (Main/Gag-Pol/Envelope)

a, b, c: Plasmid sizes (Main/Gag-Pol/Envelope) (base pairs)

<sup>2</sup> DNA mass calculation of each plasmid:

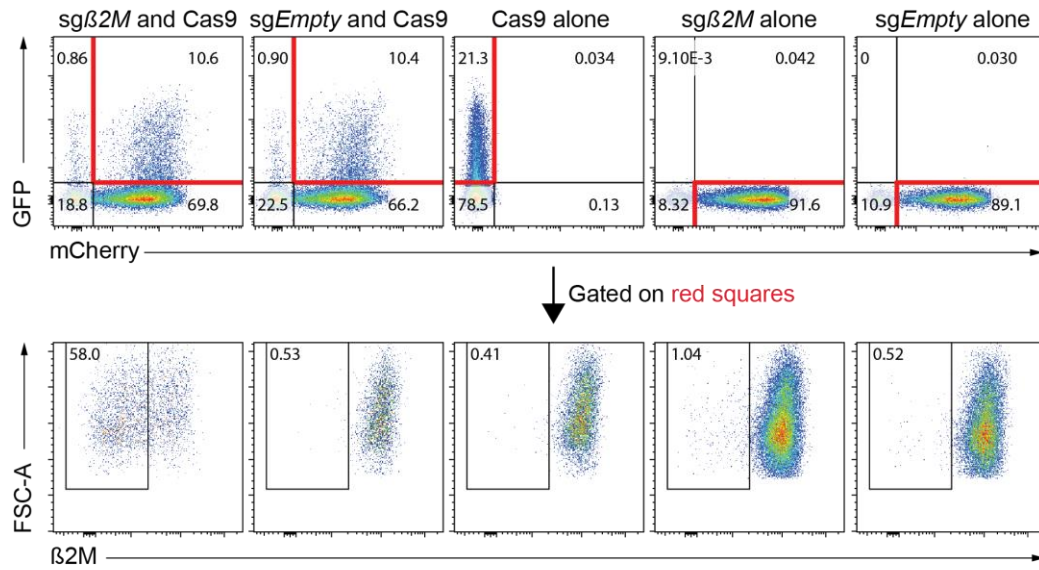
$$\text{DNA mass } (\mu\text{g}) = \frac{\text{Picomole of plasmid} * (\text{Plasmid size} * 617.96 + 36.04)}{10^6}$$

The calculations in Table S1 were adapted following formula at:

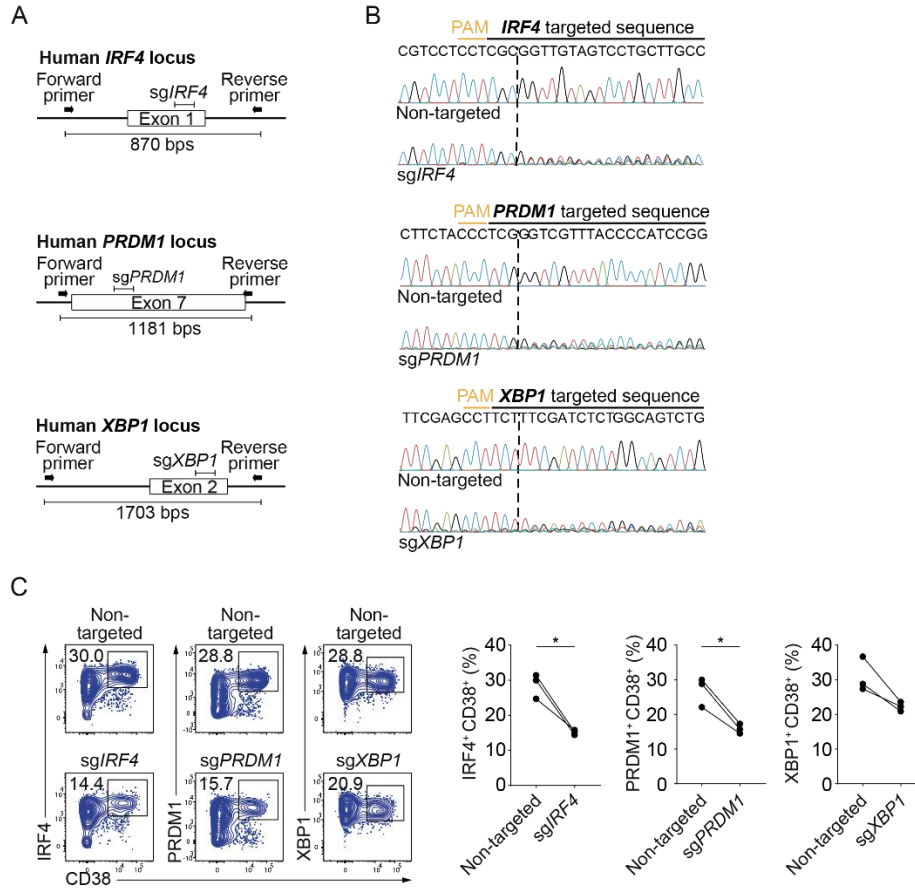
<https://nebiocalculator.neb.com/#!/dsdnaamt>

**Table S2: Oligonucleotide and primer sequences**

Name	Sequence (5'-3')	Purpose
hU6-forward	acgatacaaggctgtagagag	For sequencing of inserted gene specific targeting sequence
Furin-T2A-GFP-forward	aatgaaaagcttaggcggaagcgggggtcagg agagggcagaggaagtcttctaacaatgcgg	Amplify T2A-GFP sequence from plasmid pMSCV-Cas9-2A-GFP-sgRNA (Addgene)
Furin-T2A-GFP-reverse	taacaaaagcttttactgtacagctcgtccatgcc gaga	Amplify T2A-GFP sequence from plasmid pMSCV-Cas9-2A-GFP-sgRNA (Addgene)
sg $\beta 2M$ -forward	caccggagtagcgcgagcacagcta	Human $\beta 2M$ targeting sequence
sg $\beta 2M$ -reverse	aaactagctgtgctcgcgctactcc	Human $\beta 2M$ targeting sequence
$\beta 2M$ -forward	gtctagaatgagcgccccg	Amplify human $\beta 2M$ locus
$\beta 2M$ -reverse	tgctctggagaatctcacgc	Amplify human $\beta 2M$ locus
sgIRF4-forward	caccgcaagcaggactacaaccgcg	Human IRF4 targeting sequence
sgIRF4-reverse	aaaccgcggttgtagtctctgcttgc	Human IRF4 targeting sequence
IRF4-forward	actgacagagtcgcggggaag	Amplify human IRF4 locus
IRF4-reverse	agagccgaggcctccttctctc	Amplify human IRF4 locus
sgPRDM1-forward	caccgggatggggtaaacgaccgga	Human PRDM1 targeting sequence
sgPRDM1-reverse	aaactcgggtcgtttaccccatccc	Human PRDM1 targeting sequence
PRDM1-forward	tcagttctcttagccctctgtgaatcgc	Amplify human PRDM1 locus
PRDM1-reverse	gactgctctctcaaggcctaccttcag	Amplify human PRDM1 locus
sgXBP1-forward	caccggactgccagagatcgaaaga	Human XBP1 targeting sequence
sgXBP1-reverse	aaactcttctgatctctggcagtcc	Human XBP1 targeting sequence
XBP1-forward	aattggactgggggacggag	Amplify human XBP1 locus
XBP1-reverse	ataggggctgaaacaacttggg	Amplify human XBP1 locus



**Figure S1: Efficient CRISPR/Cas9-mediated knockout of  $\beta 2M$  housekeeping gene in primary human B cells.** Representative FACS plots show the frequency of  $\beta 2M^{-}$  B cells in *sg $\beta 2M$ <sup>+</sup>Cas9<sup>+</sup>*, *sgEmpty<sup>+</sup>Cas9<sup>+</sup>*, *Cas9<sup>+</sup>*, *sg $\beta 2M$ <sup>+</sup>* and *sgEmpty<sup>+</sup>* transduced B cells at day 4 after transduction. *sgEmpty* is sgRNA without the targeted sequence.



**Figure S2: *IRF4*, *PRDM1*, and *XBP1* are crucial for B cell survival and PC differentiation *in vitro*.** (A) The schemes show the targeted site of indicated genes and primer sites for amplifying of flanking regions of *IRF4*, *PRDM1*, and *XBP1*-targeted sites by PCR. (B) Sanger sequencing signal traces show noise peaks at *IRF4*, *PRDM1*, and *XBP1*-targeted site compared to non-targeted samples, respectively. The dashed lines indicate Cas9 cleavage sites. (C) Representative FACS plots (left) and graphs (right) show the percentage of IRF4<sup>+</sup>CD38<sup>+</sup>, PRDM1<sup>+</sup>CD38<sup>+</sup> and XBP1<sup>+</sup>CD38<sup>+</sup> cells at day 8 post transduction from non-targeted compared with corresponding sg*IRF4*, sg*PRDM1*, and sg*XBP1*-targeted samples. IRF4<sup>+</sup>CD38<sup>+</sup>, PRDM1<sup>+</sup>CD38<sup>+</sup>, XBP1<sup>+</sup>CD38<sup>+</sup> cells were determined based on their isotype control stainings. Data were pooled from 3 donors. Statistical significance was calculated using paired *t*-test, only significant *p* value is shown: \**p* < 0.05. Each gene was independently targeted by three different sgRNAs. Data for one sgRNA are shown.