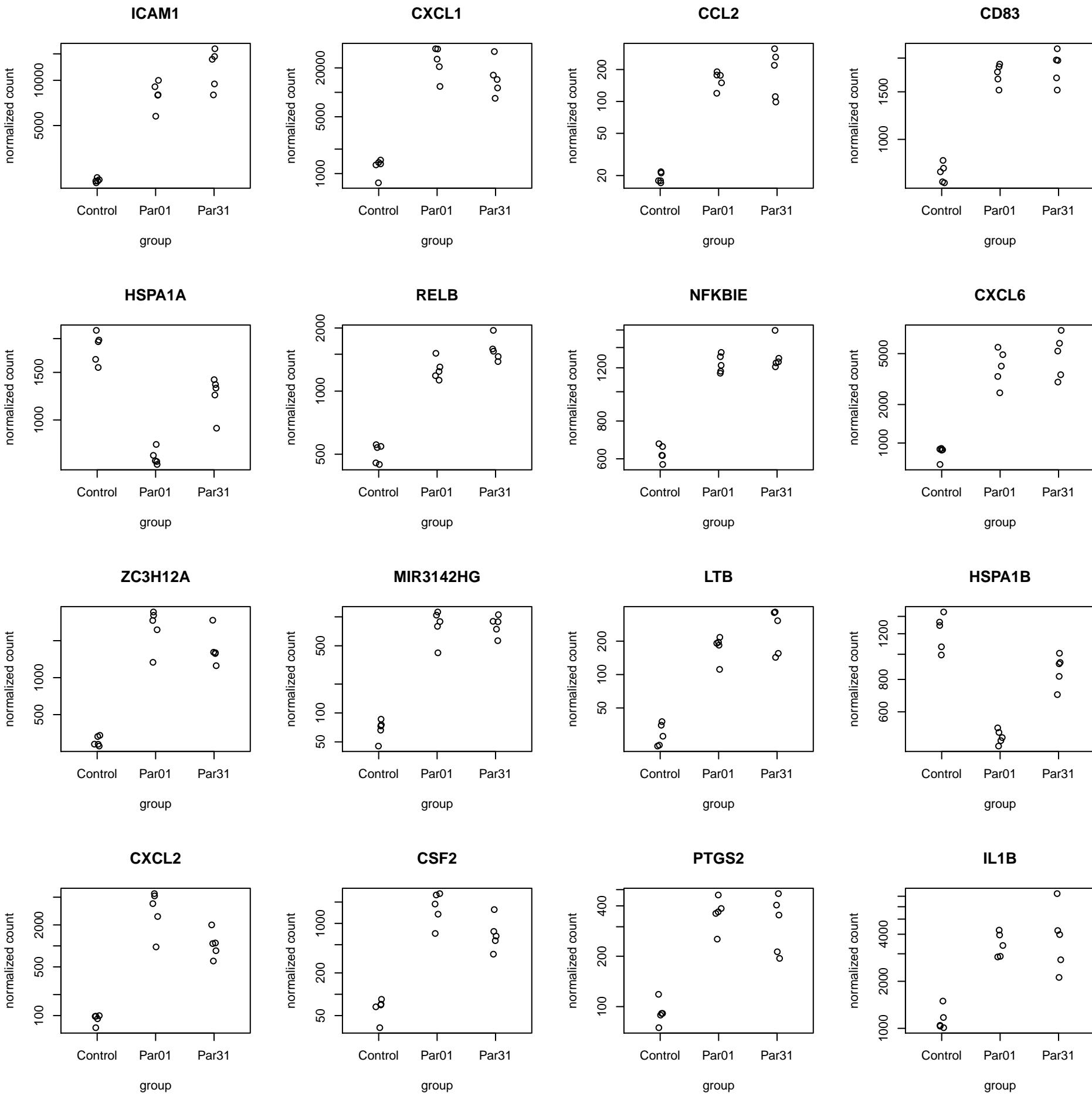
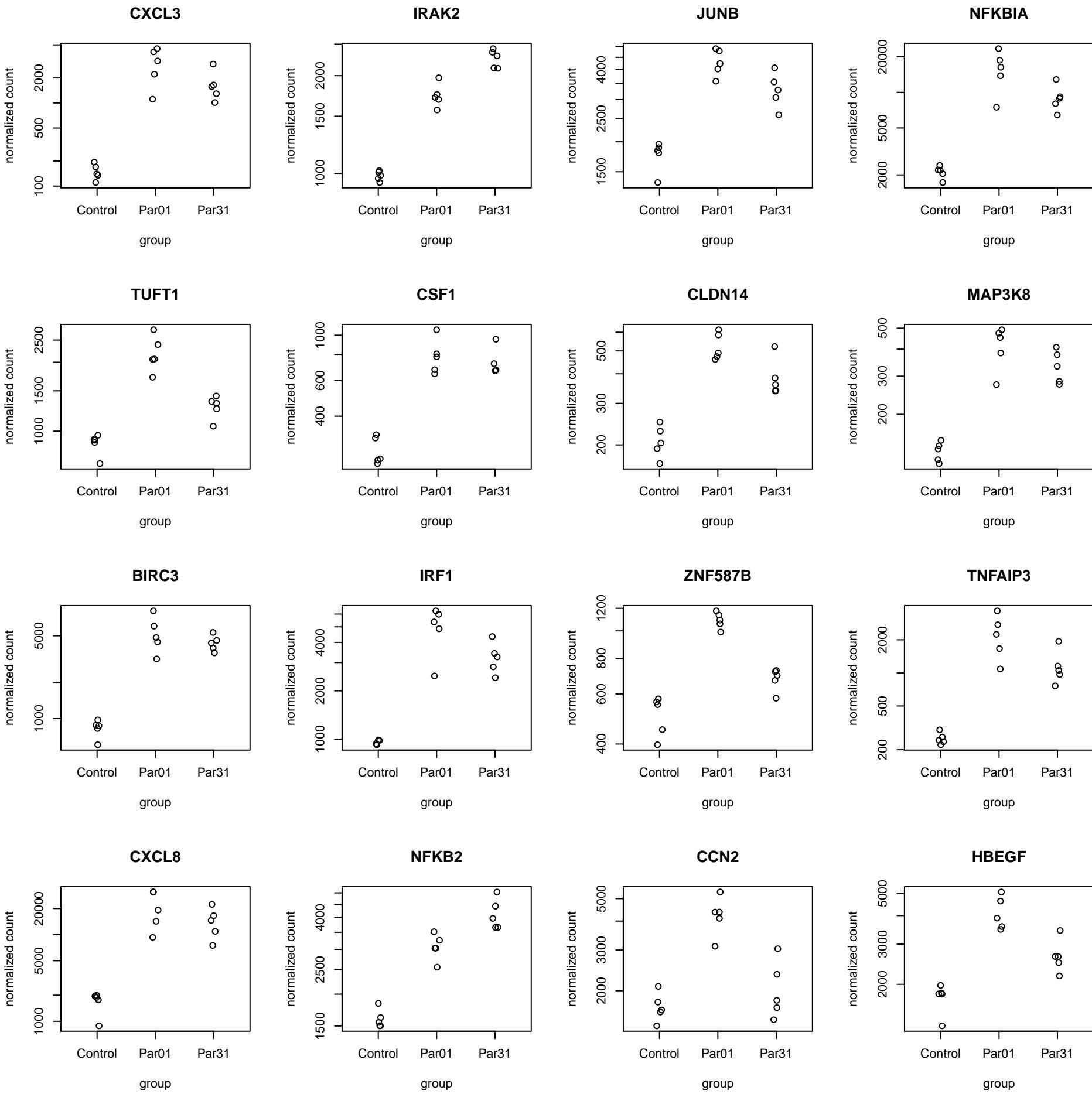
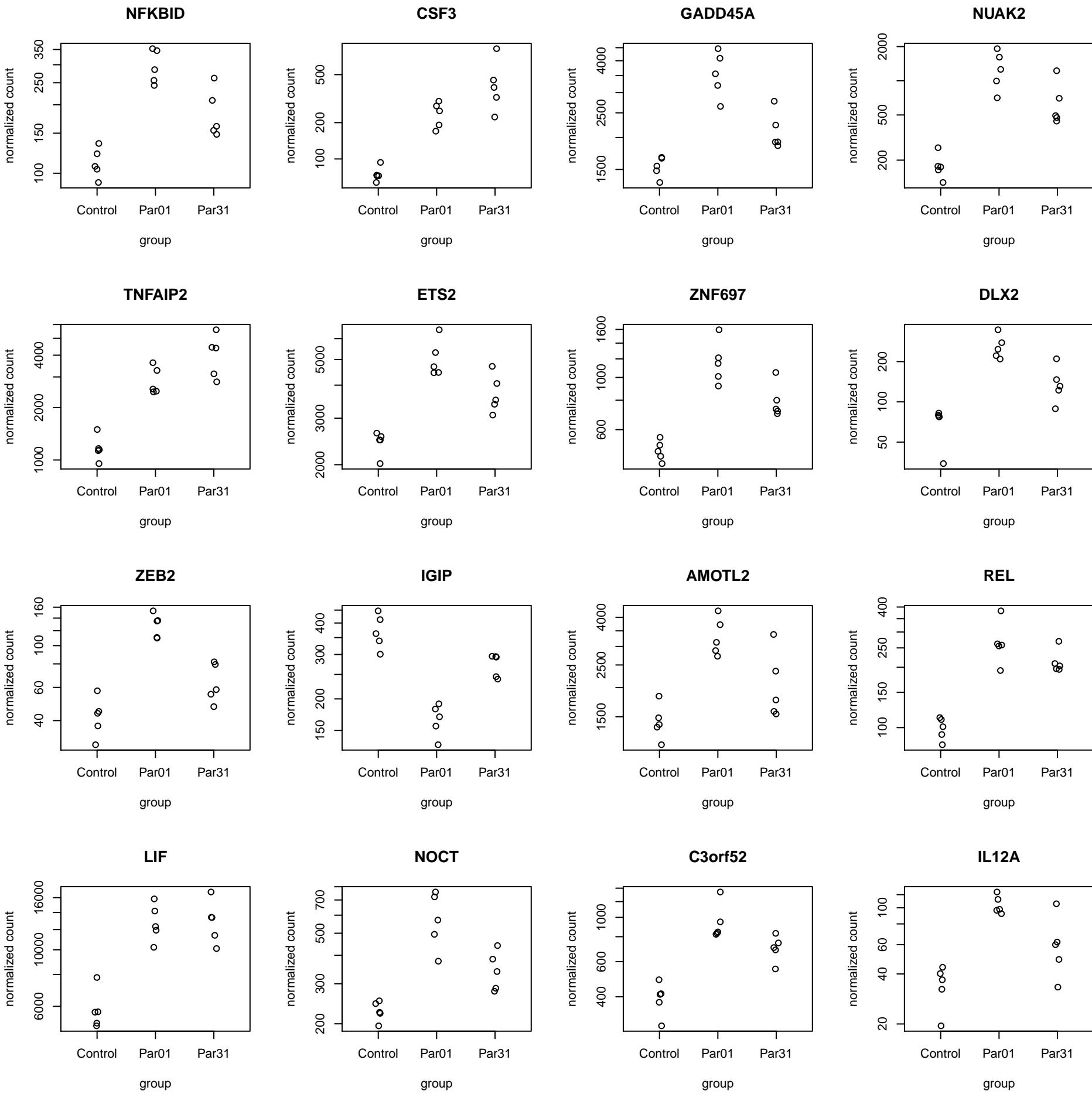
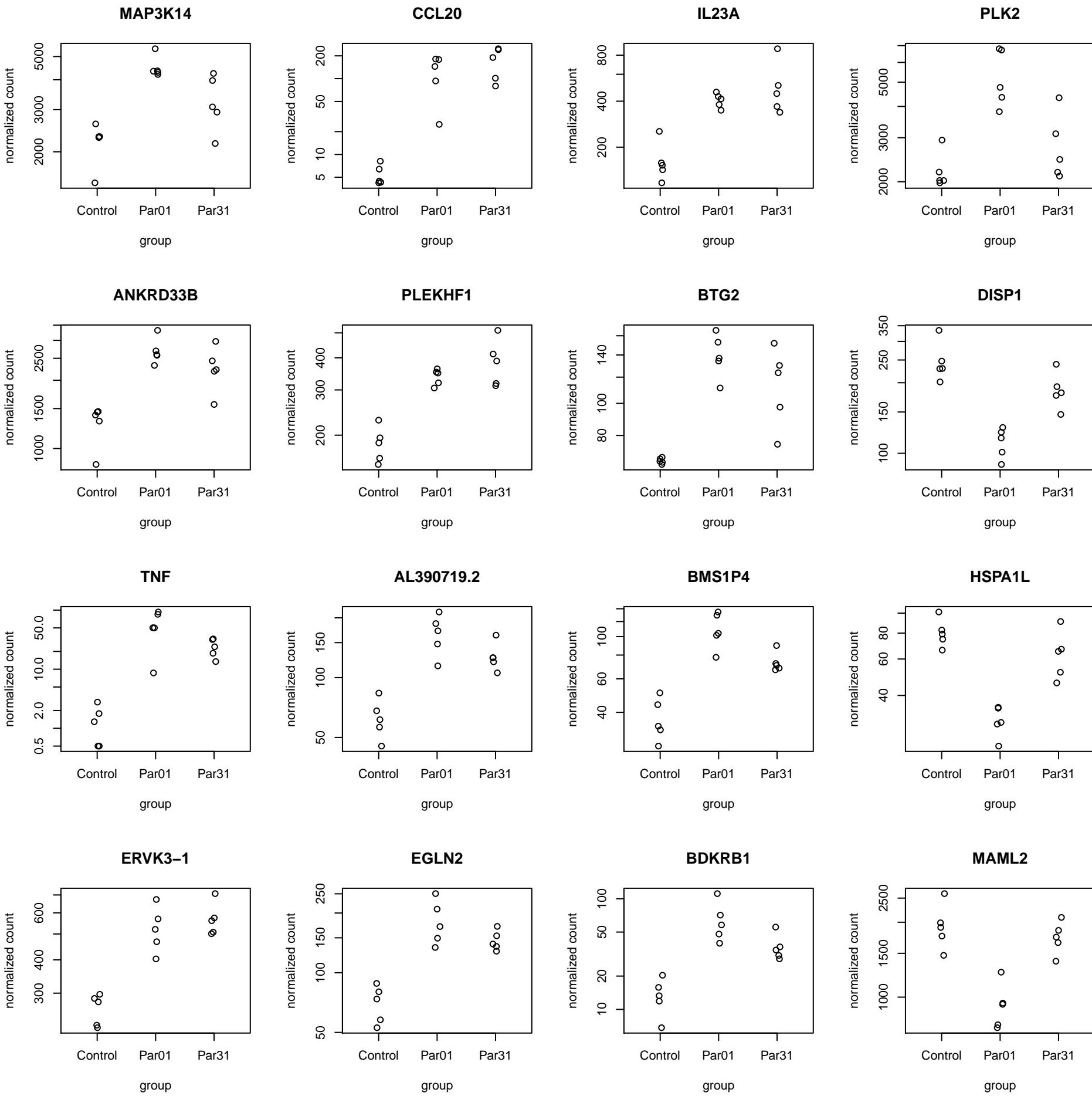


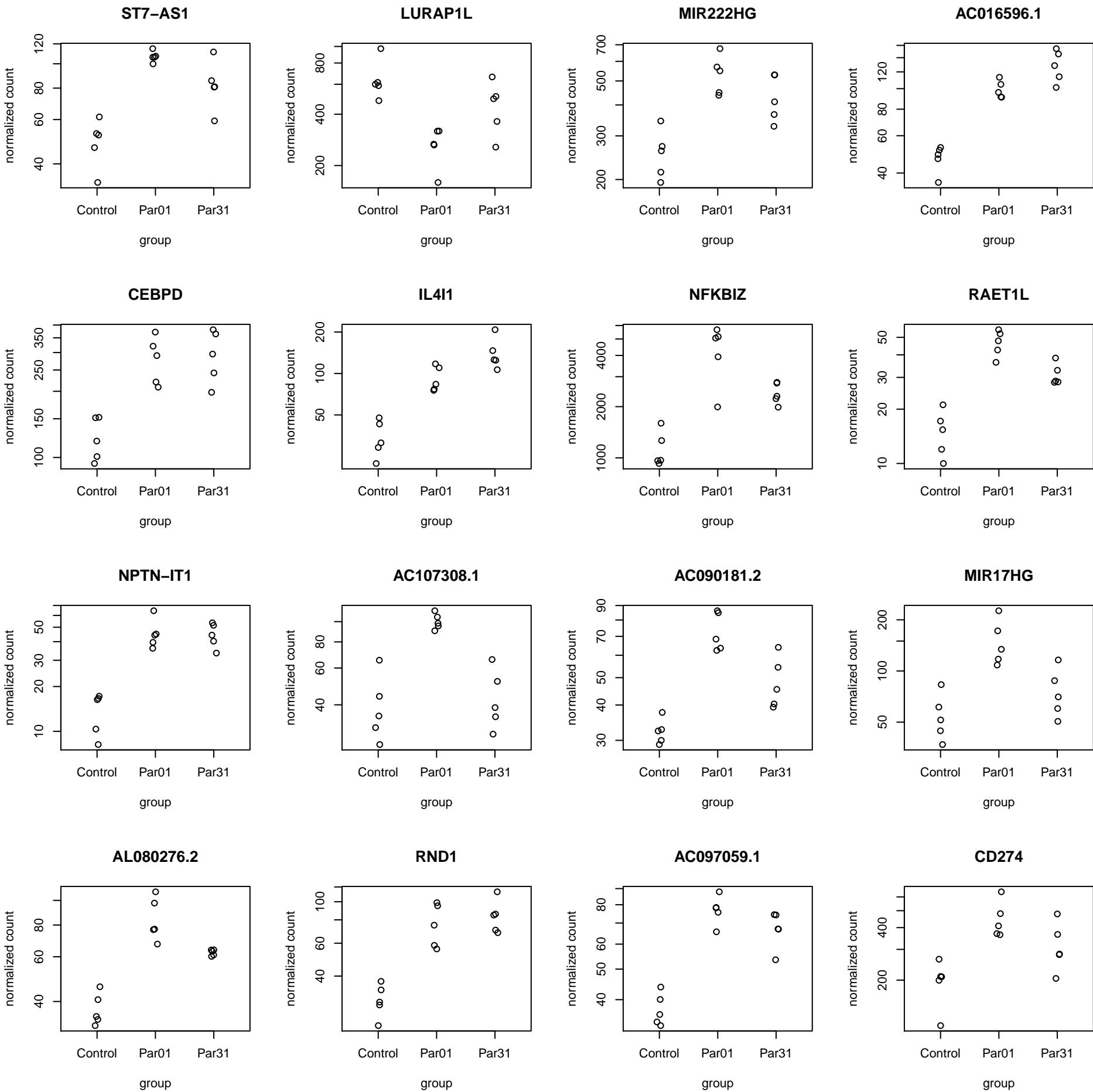
Figure S1: Impact of altered gravity on intracellular organization of F-actin: Confocal laser scanning microscopy of TRITC-conjugated phalloidin stained (F-actin) samples. A-C: PC-3 cells cultivated under altered gravity conditions. A: Static 1g ground control. B: After the first parabola (1P). C: After the 31st parabola (31P). The white arrows show psudopodia and lamellipodia and yellow arrows indicate stress fibers. Scale bars represent 30 μ m.

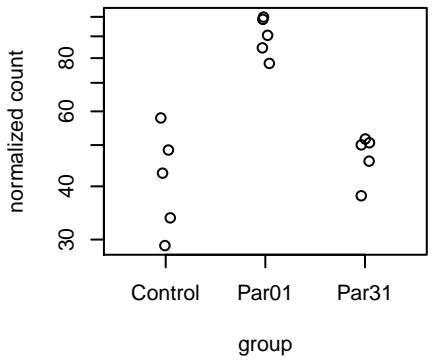
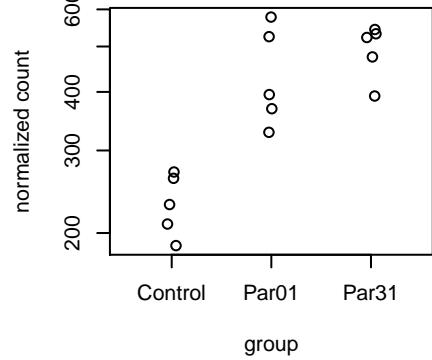
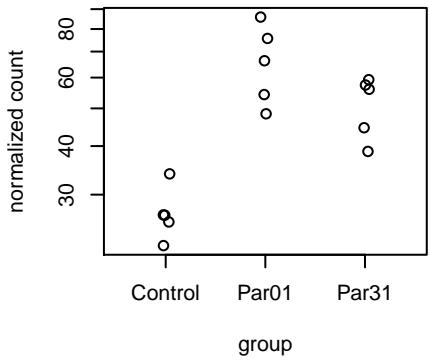
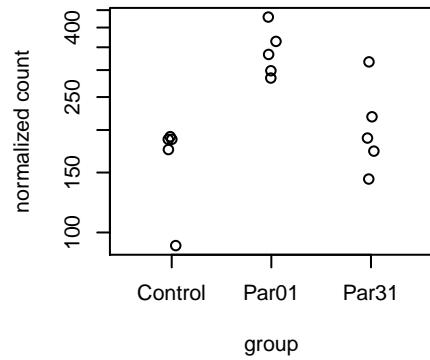
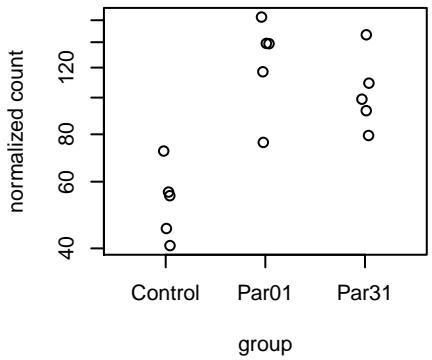
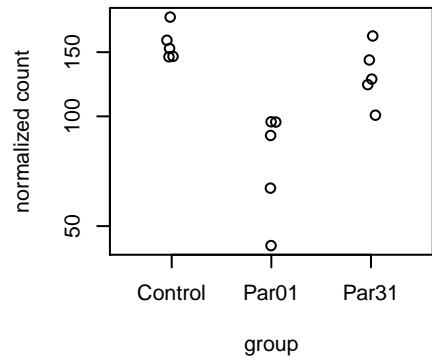
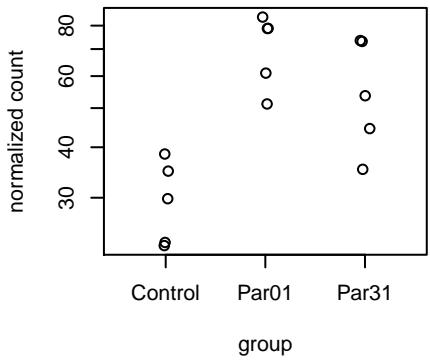
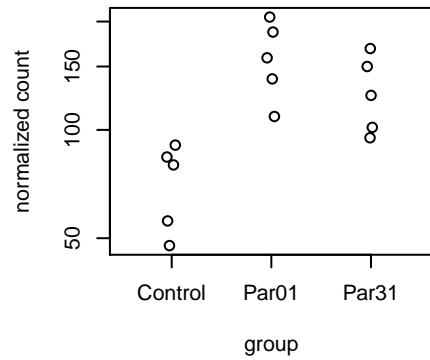
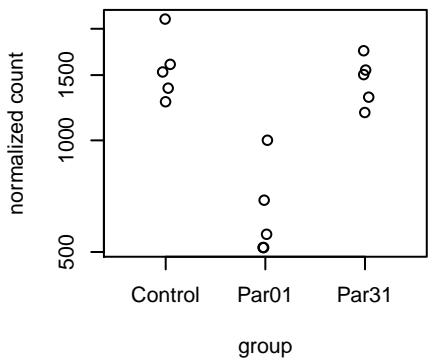
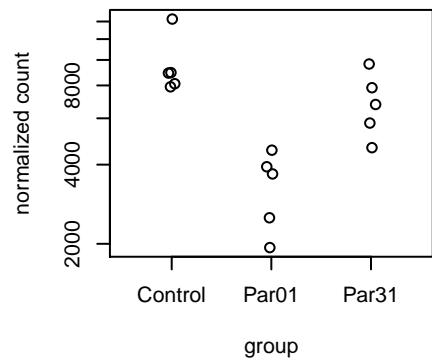
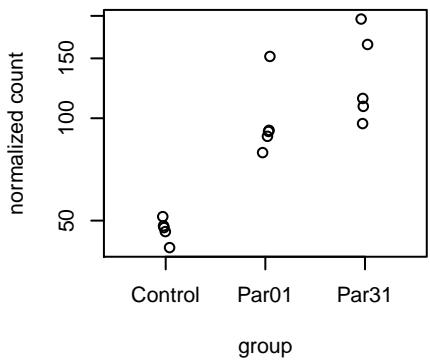
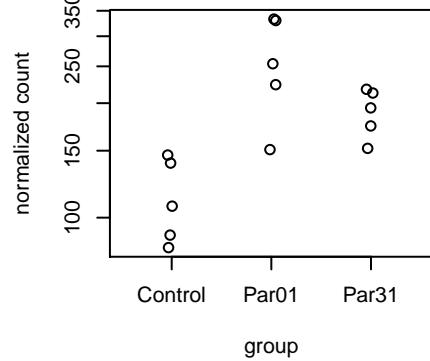
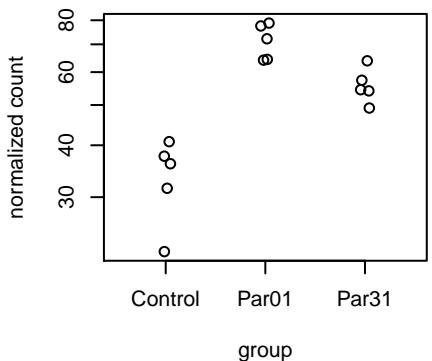
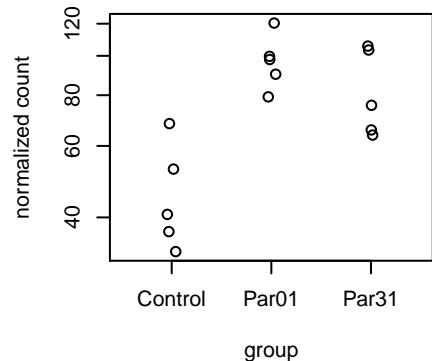
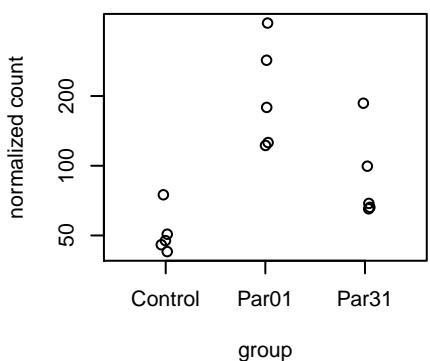
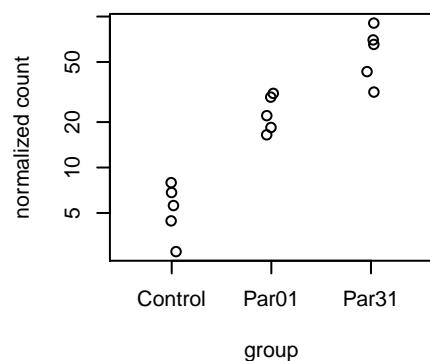


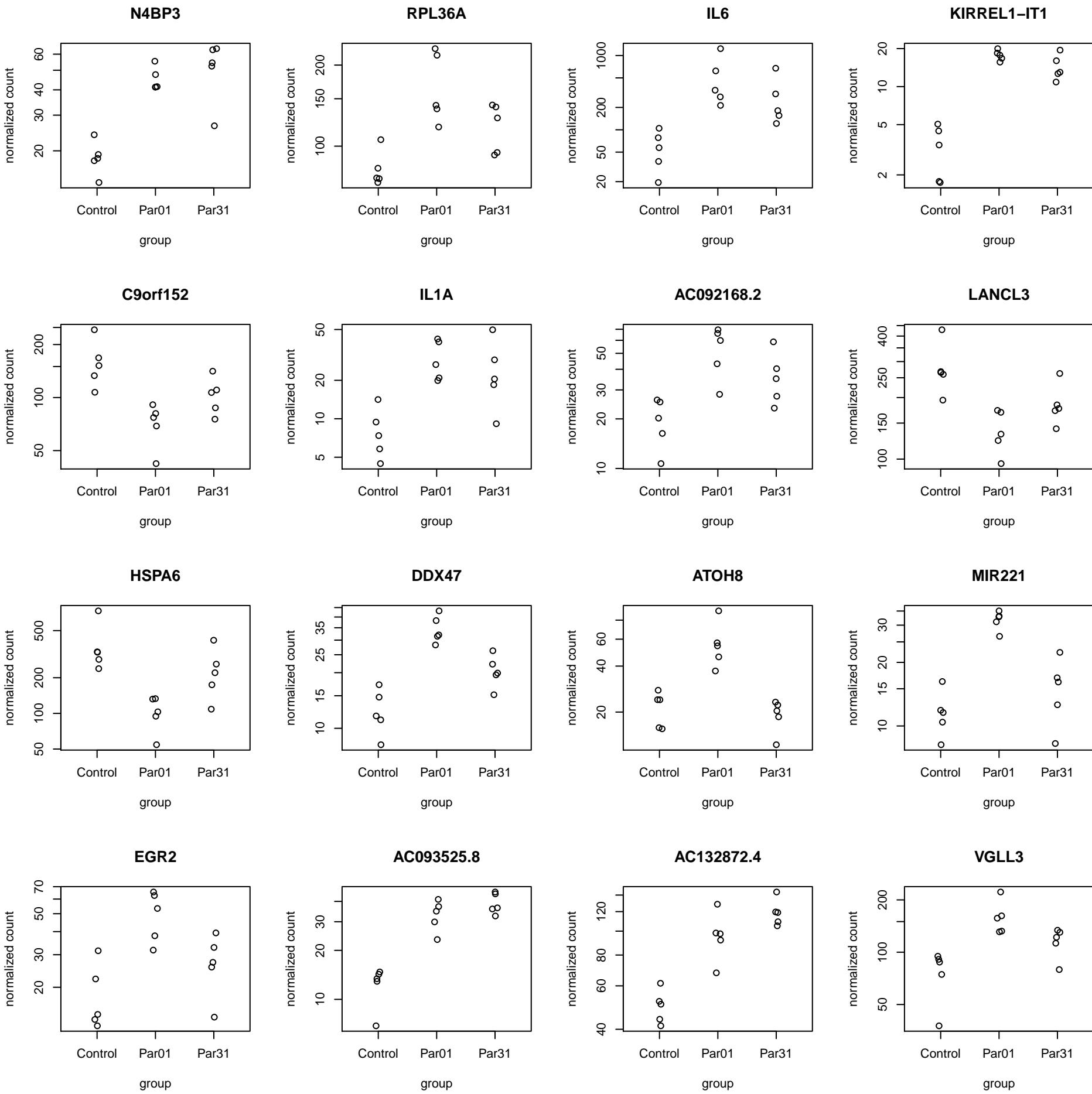


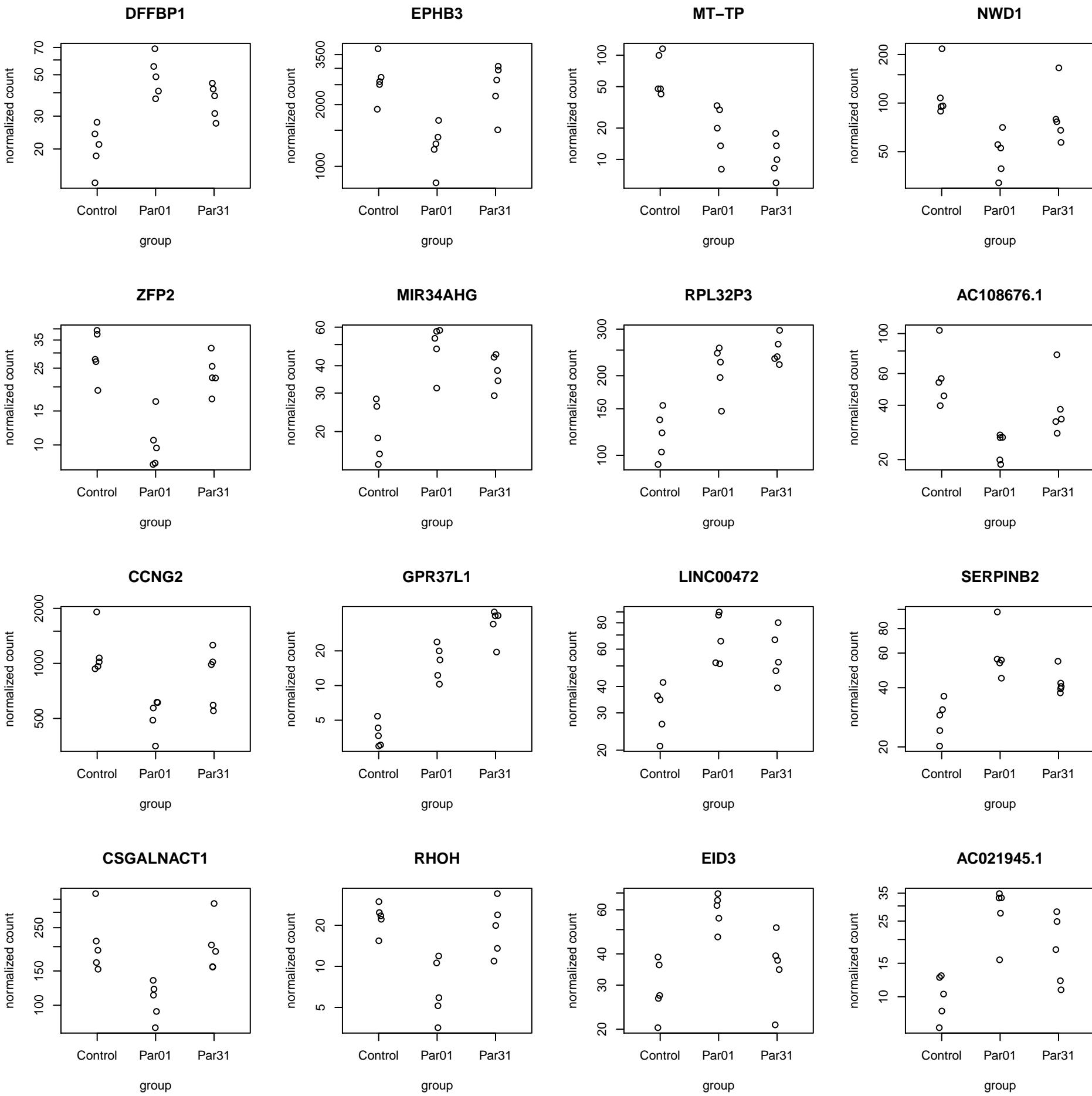


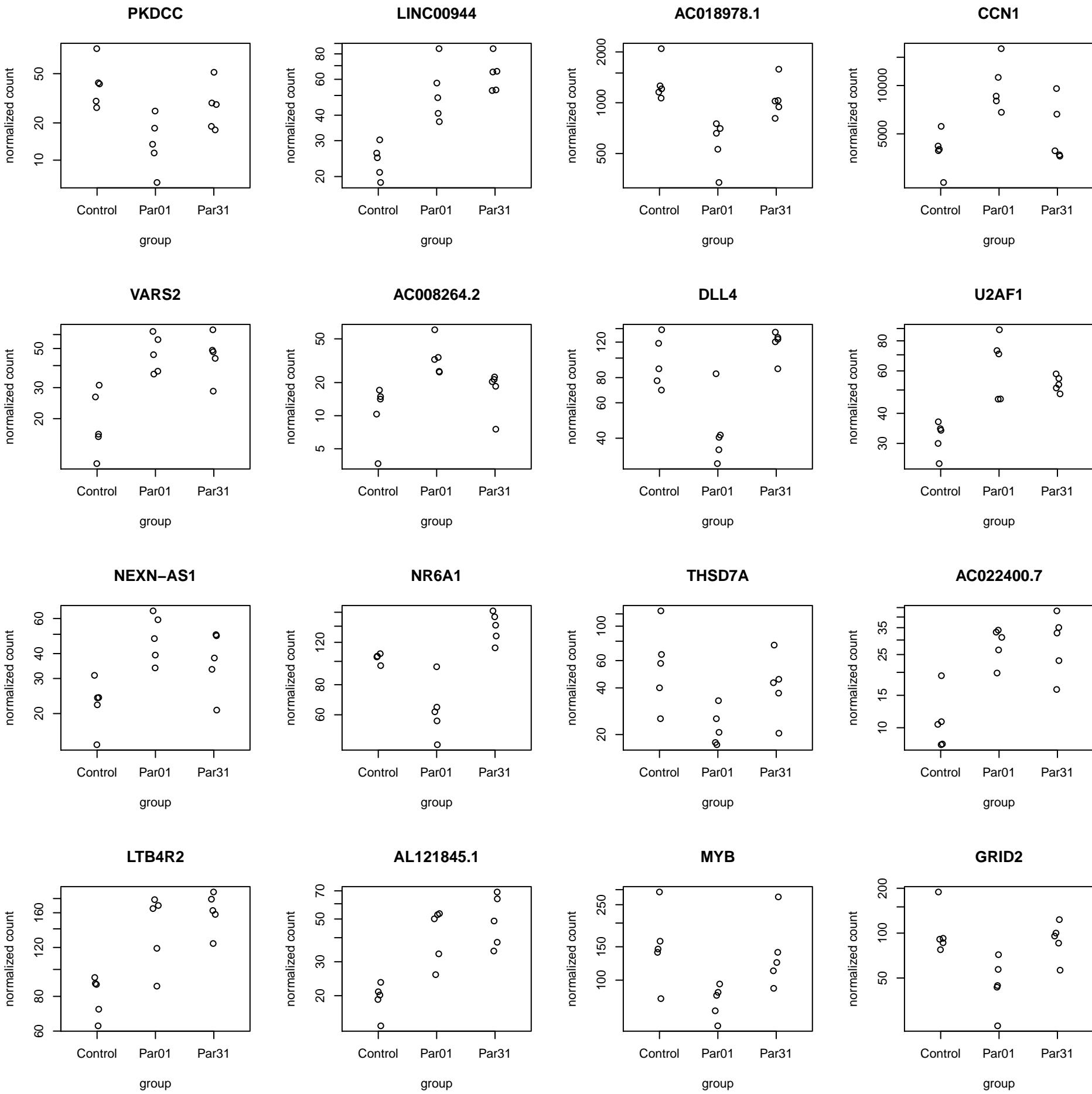


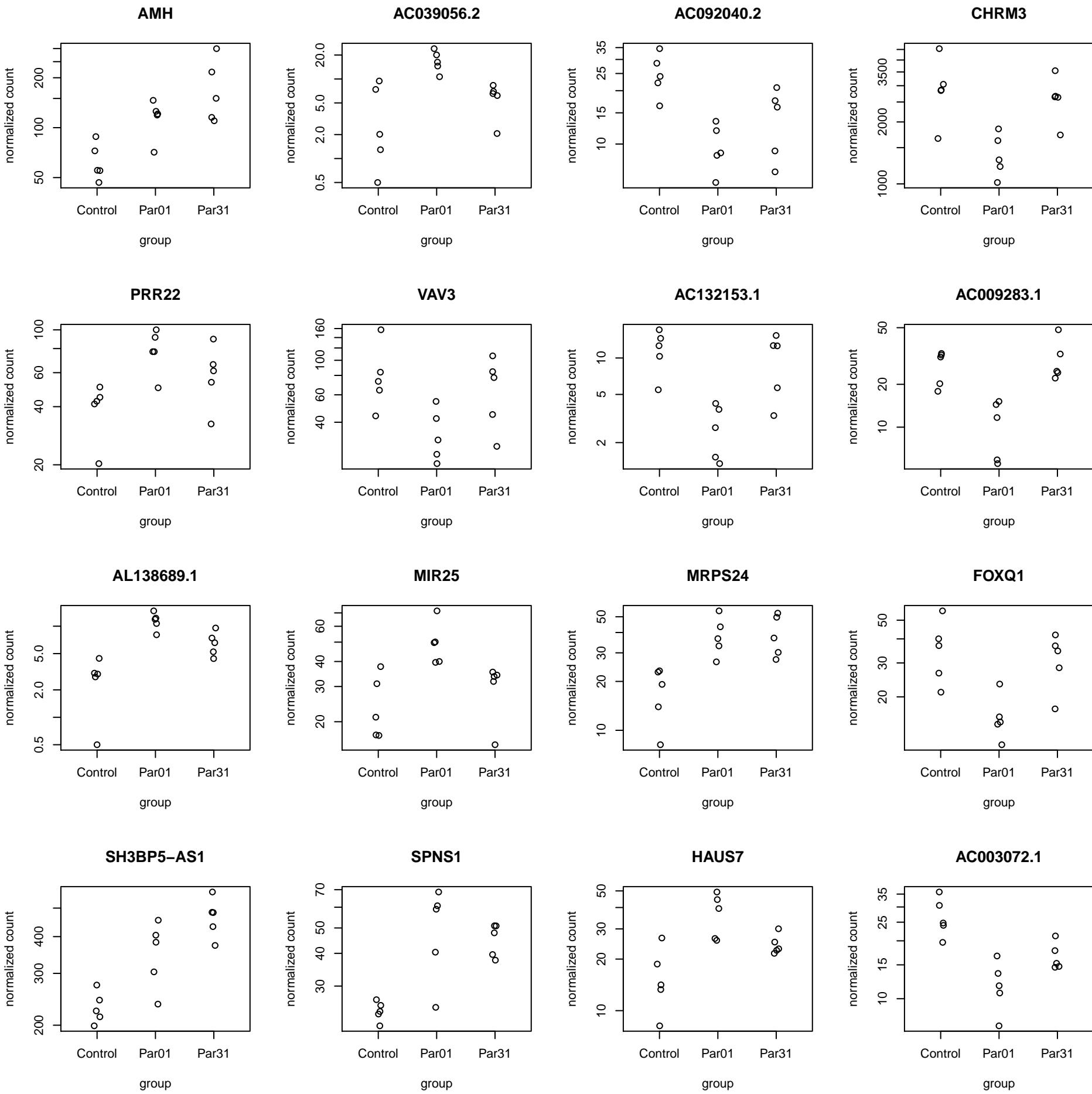


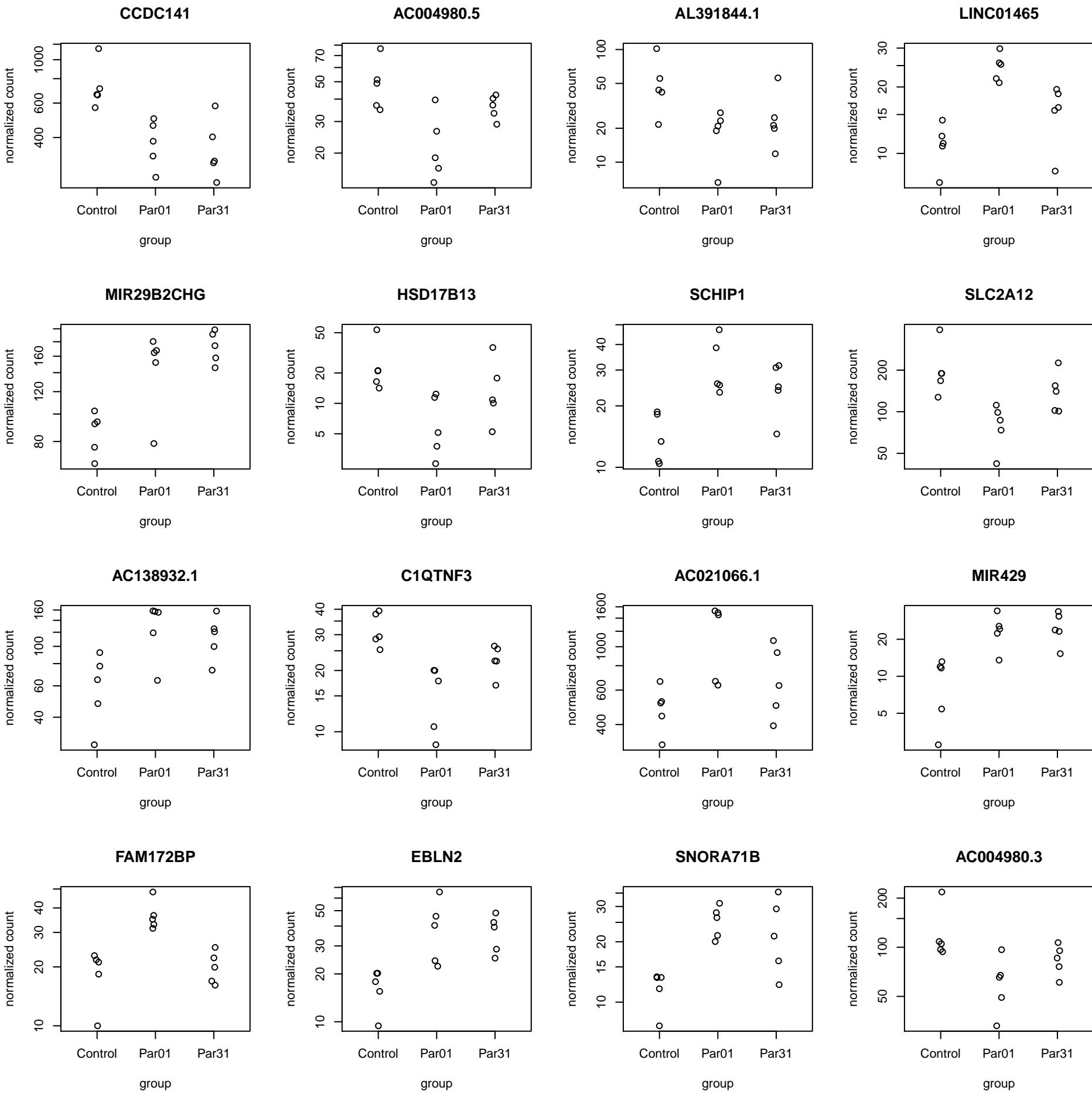
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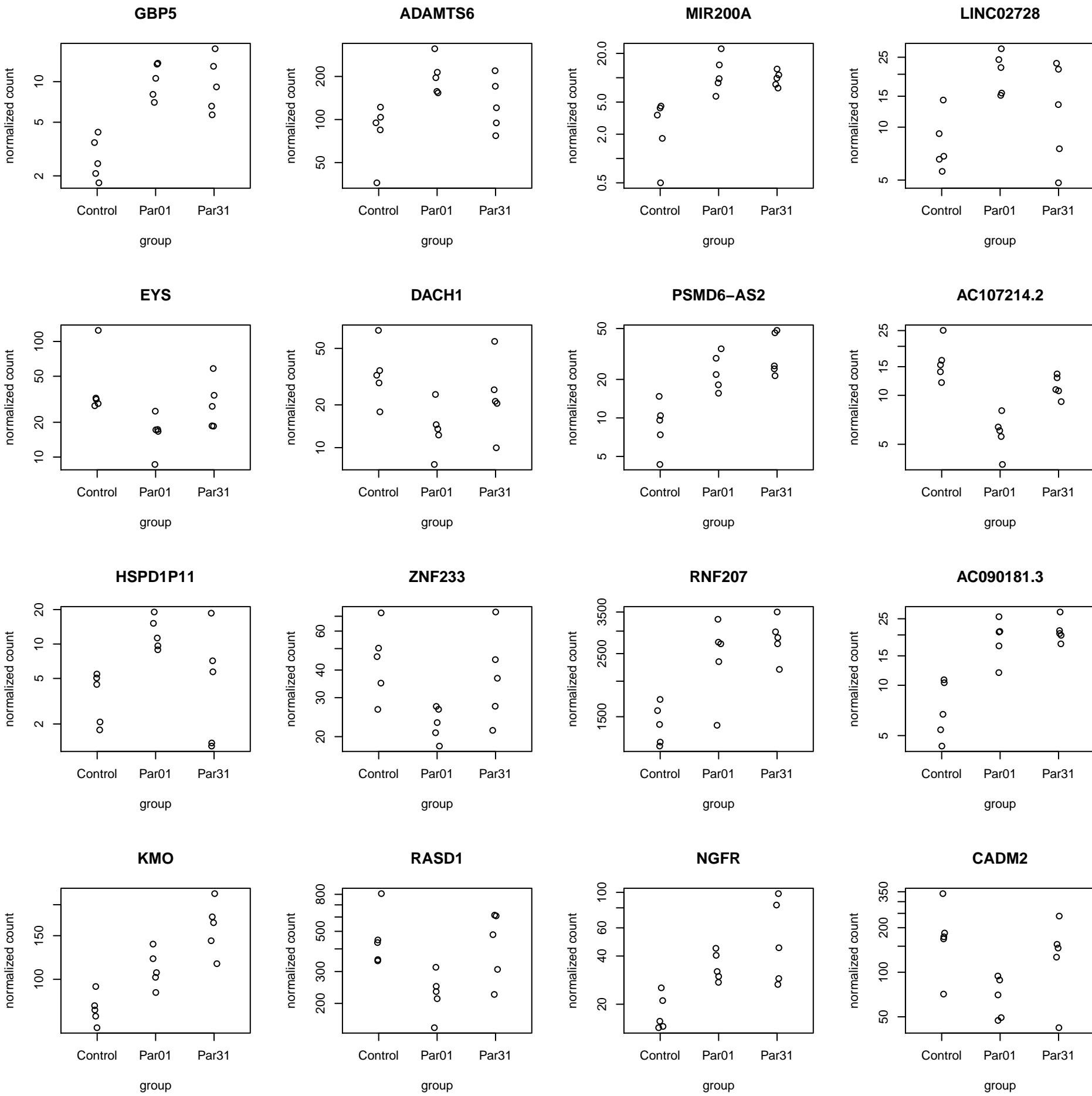


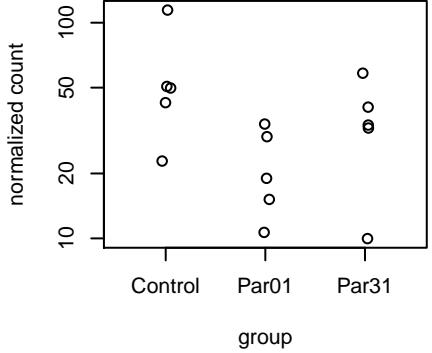
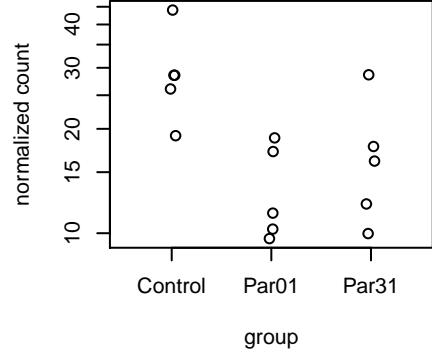
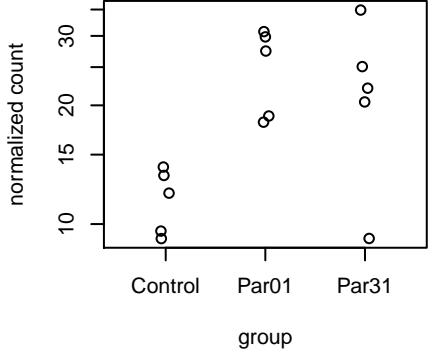
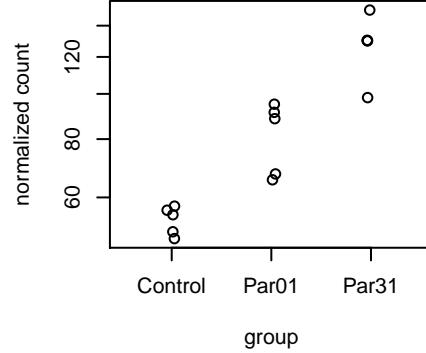
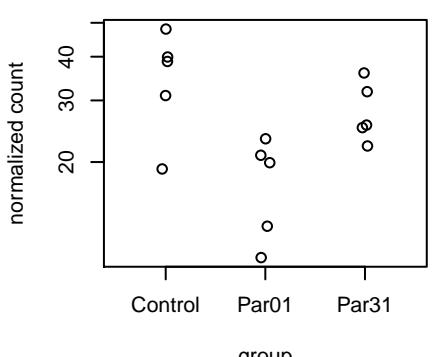
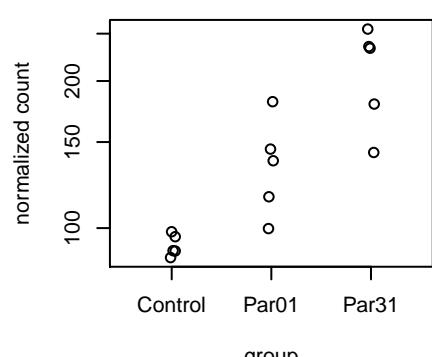
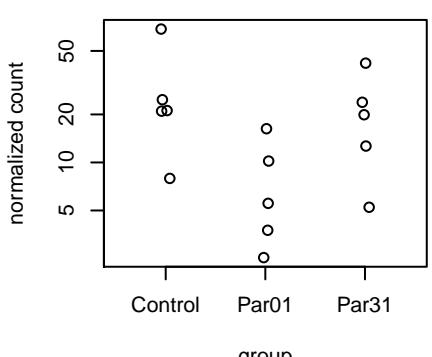
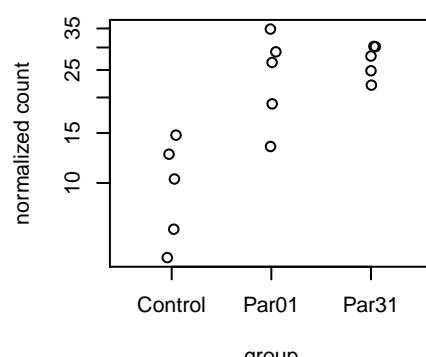
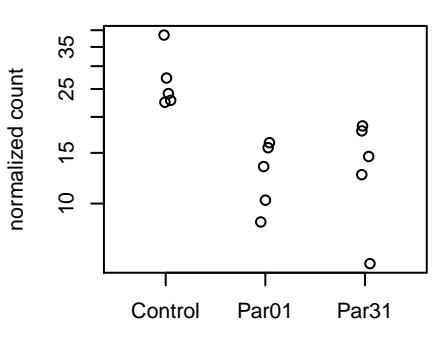
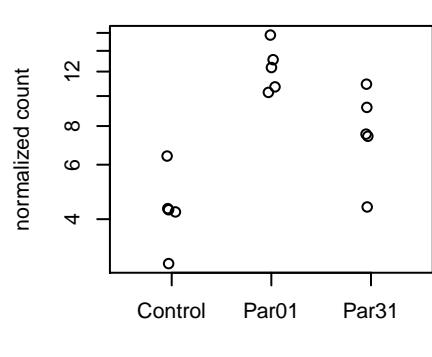
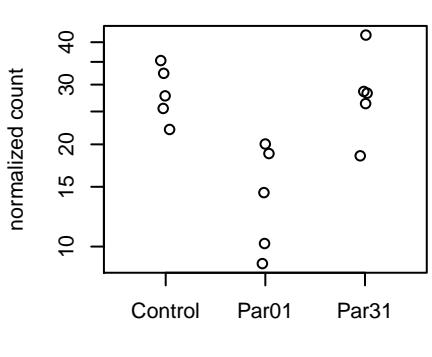
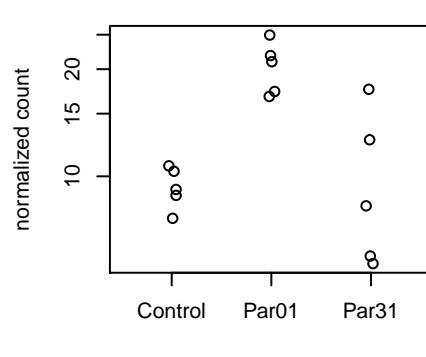
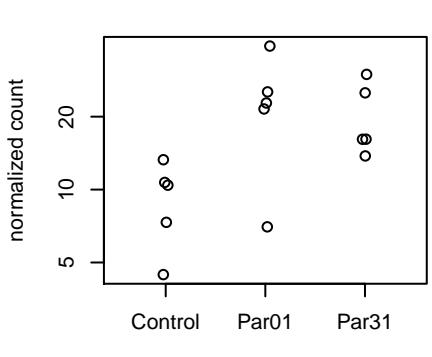
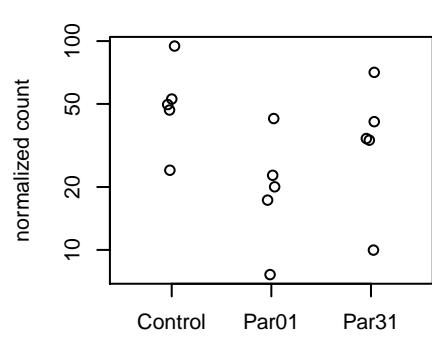
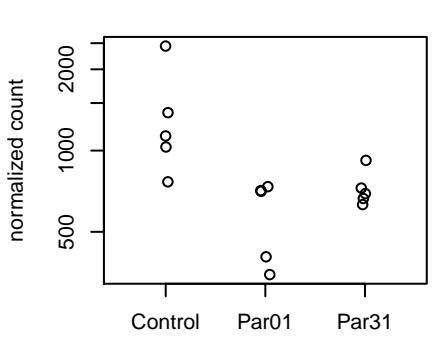
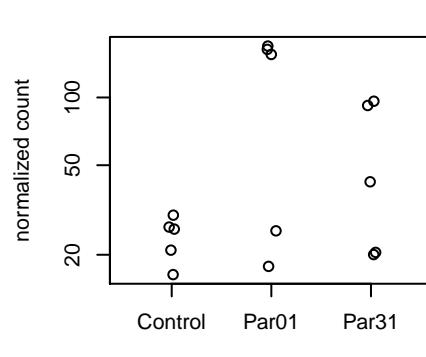


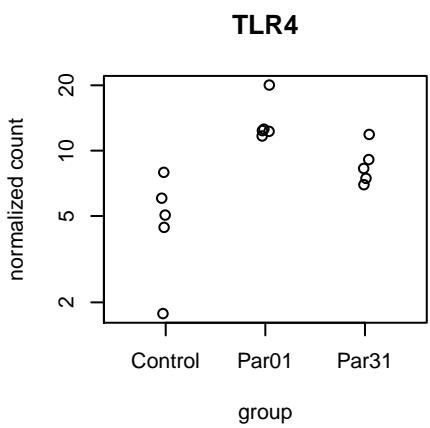
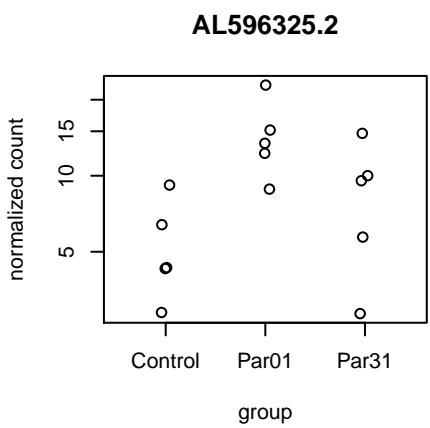
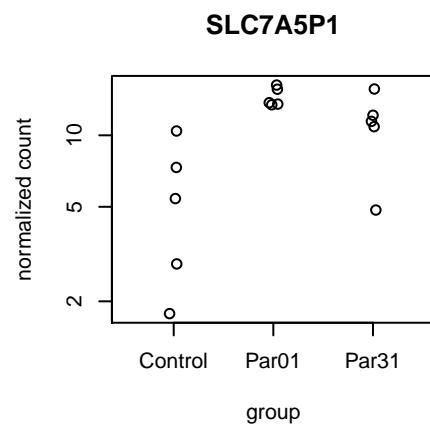
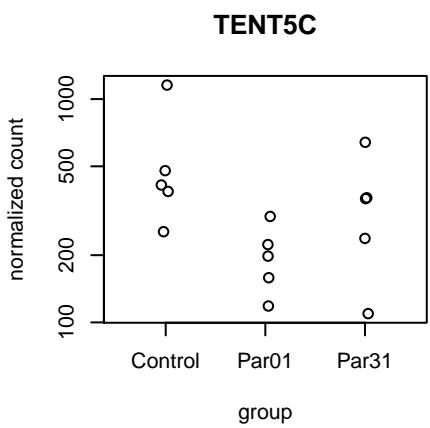
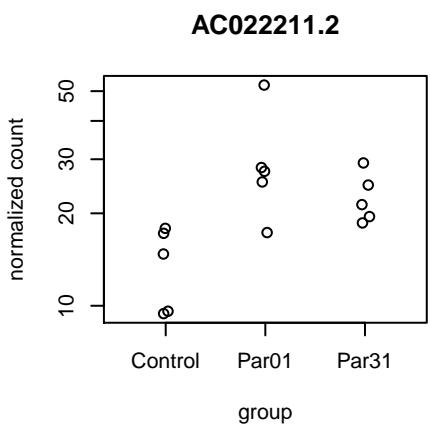
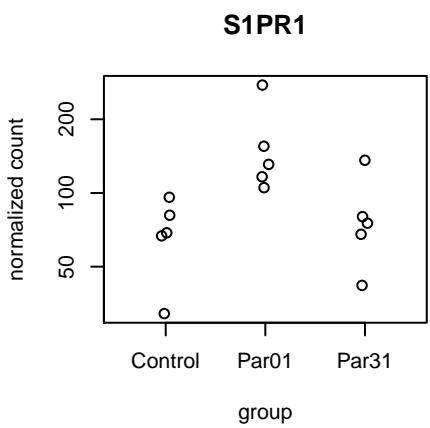
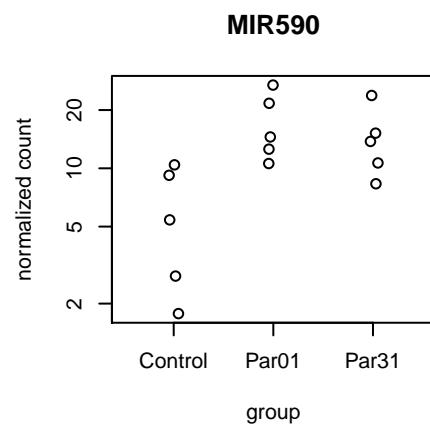
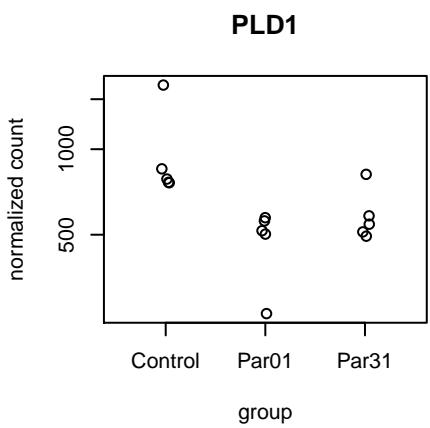
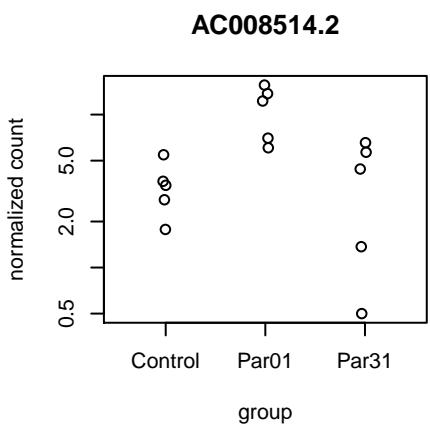
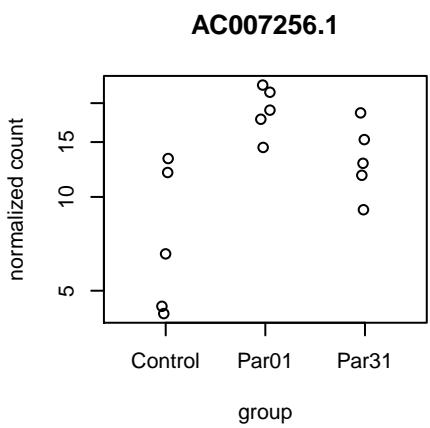
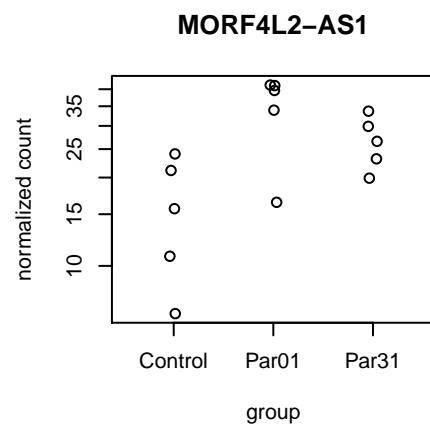
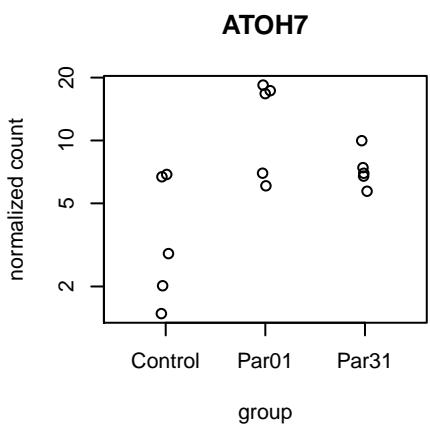
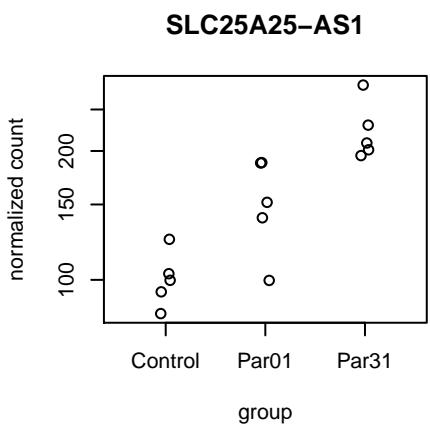
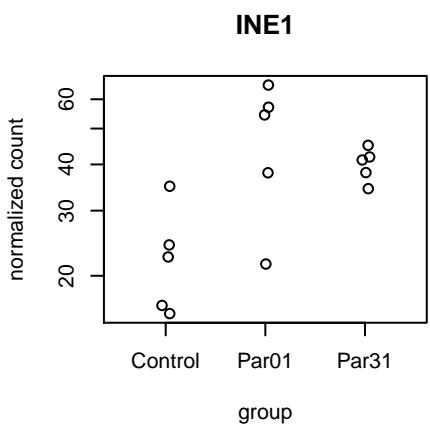
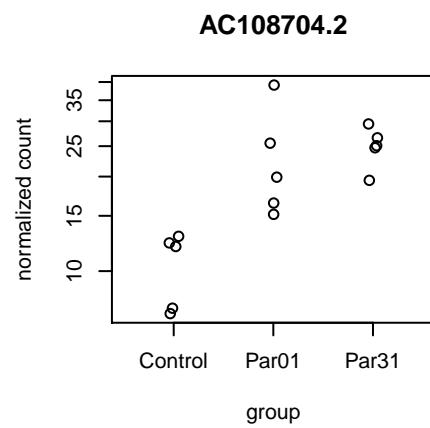
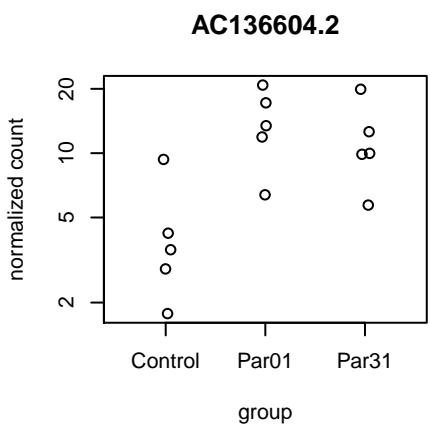


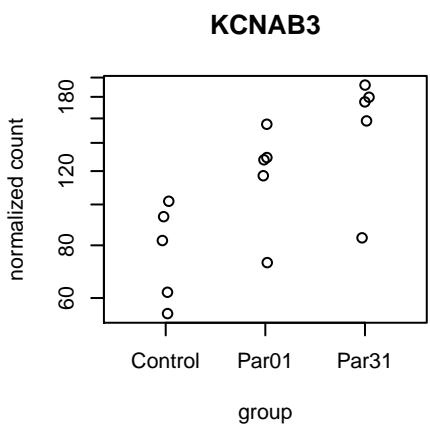
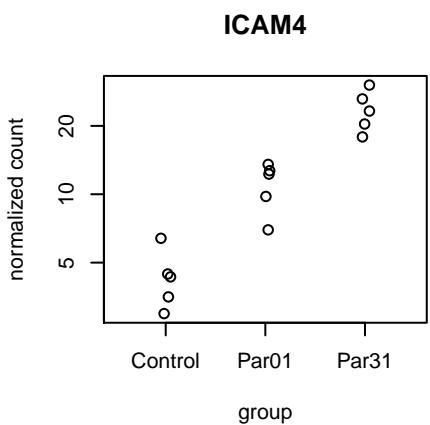
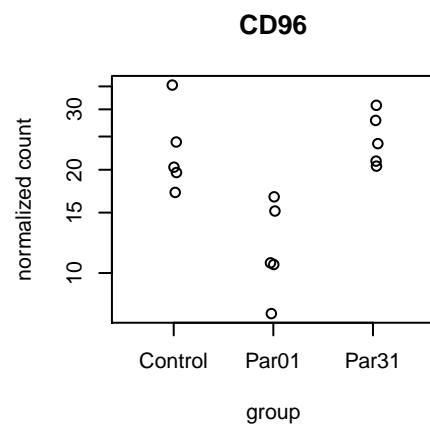
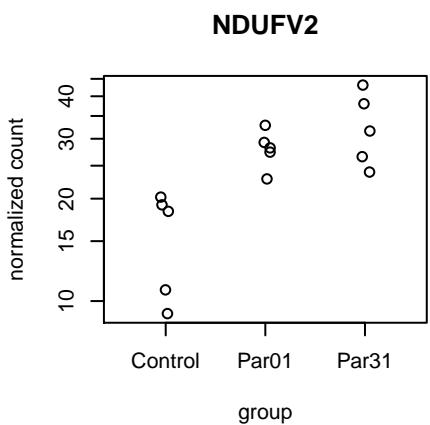
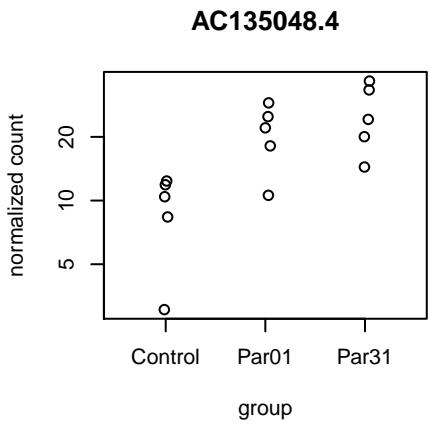
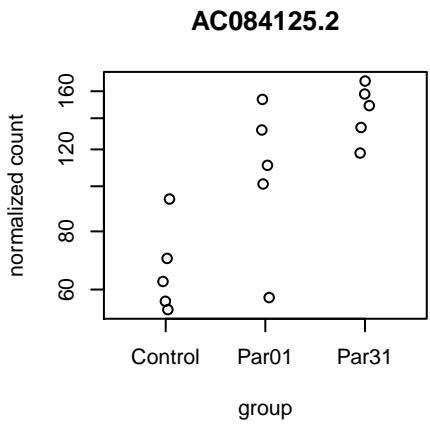
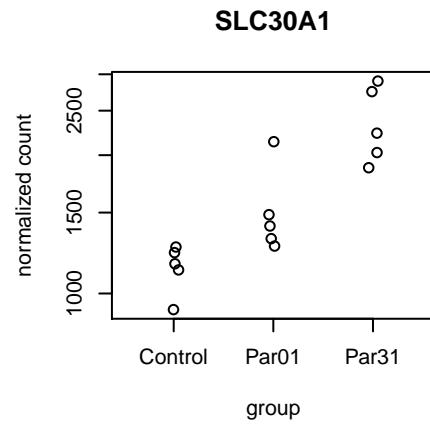
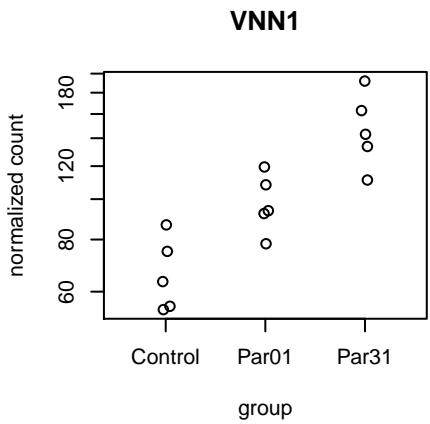
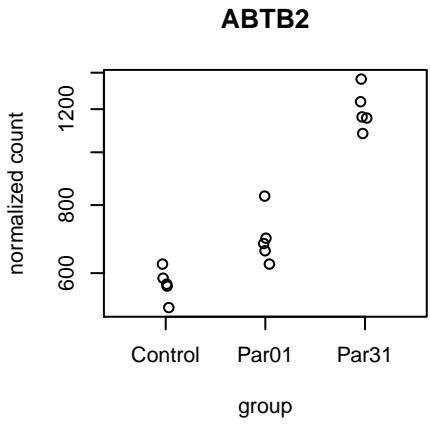
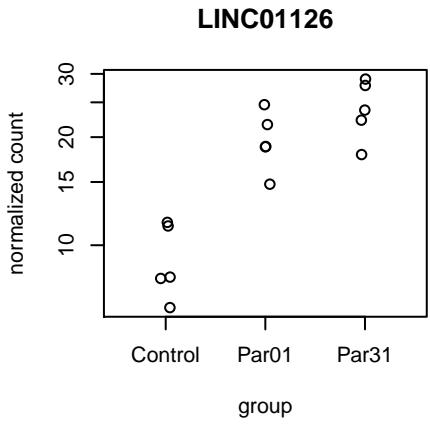
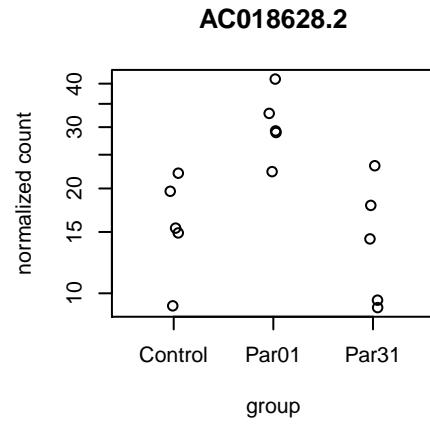
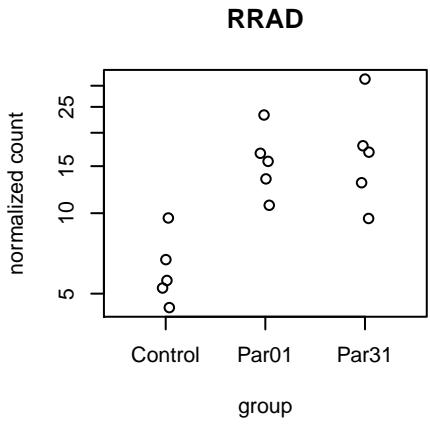
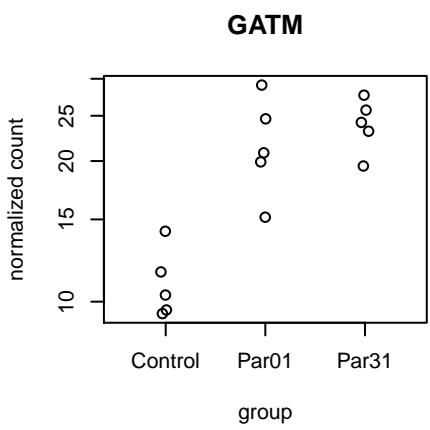
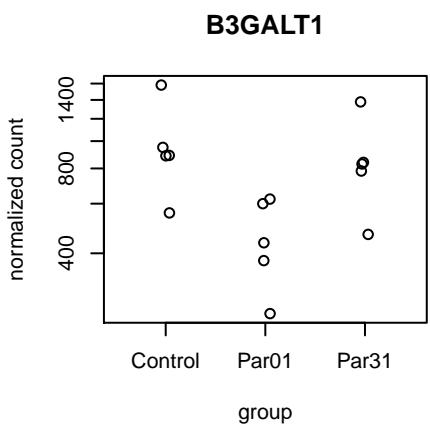
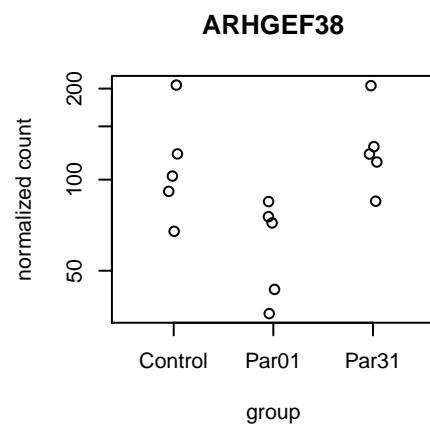
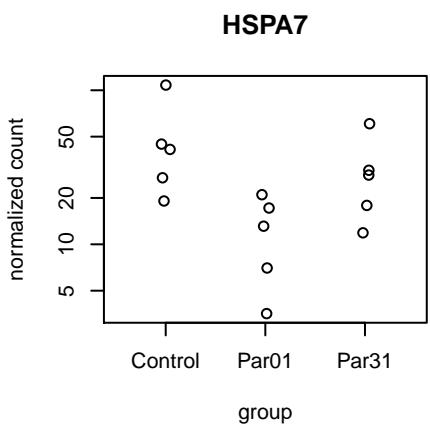


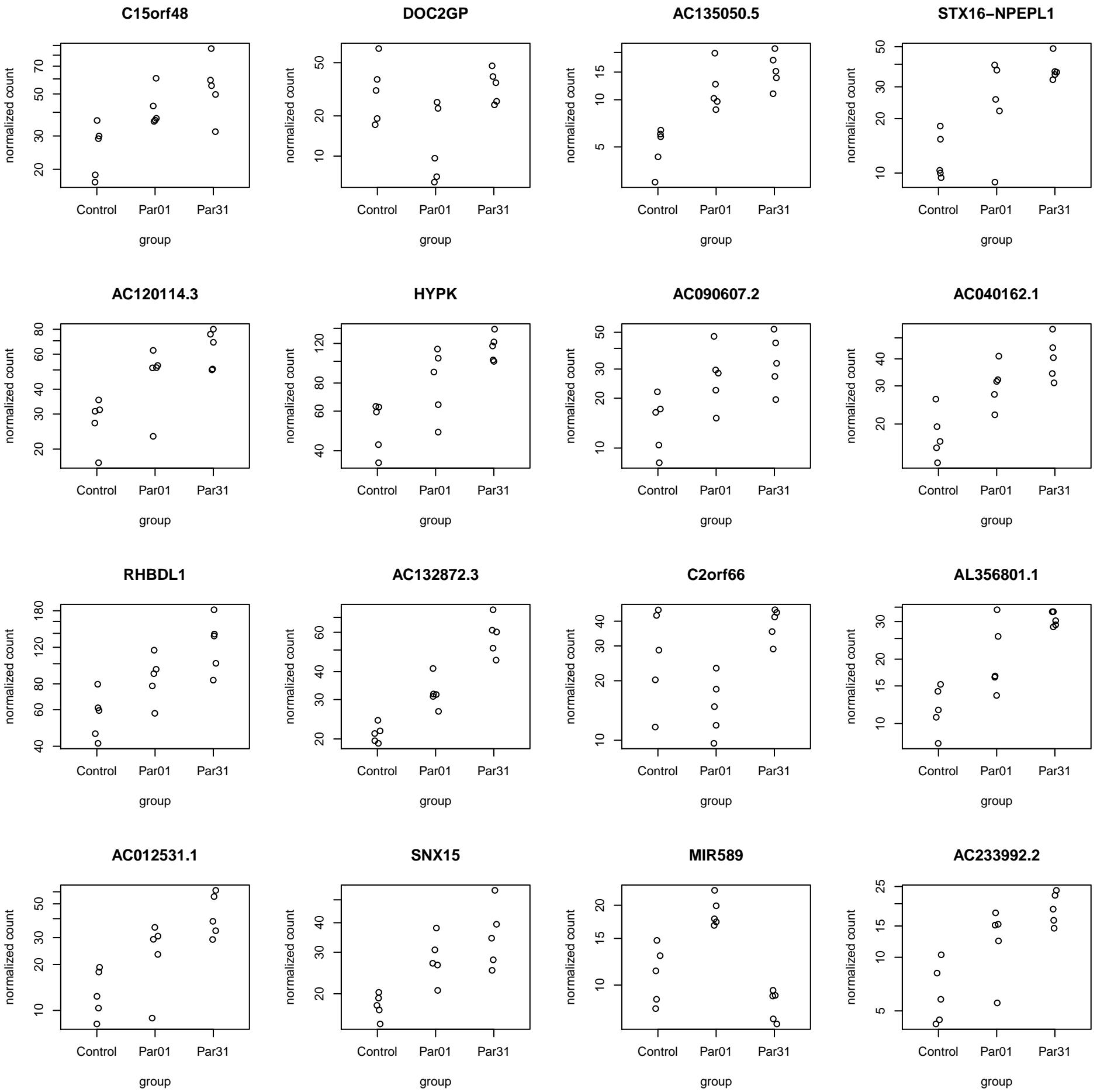


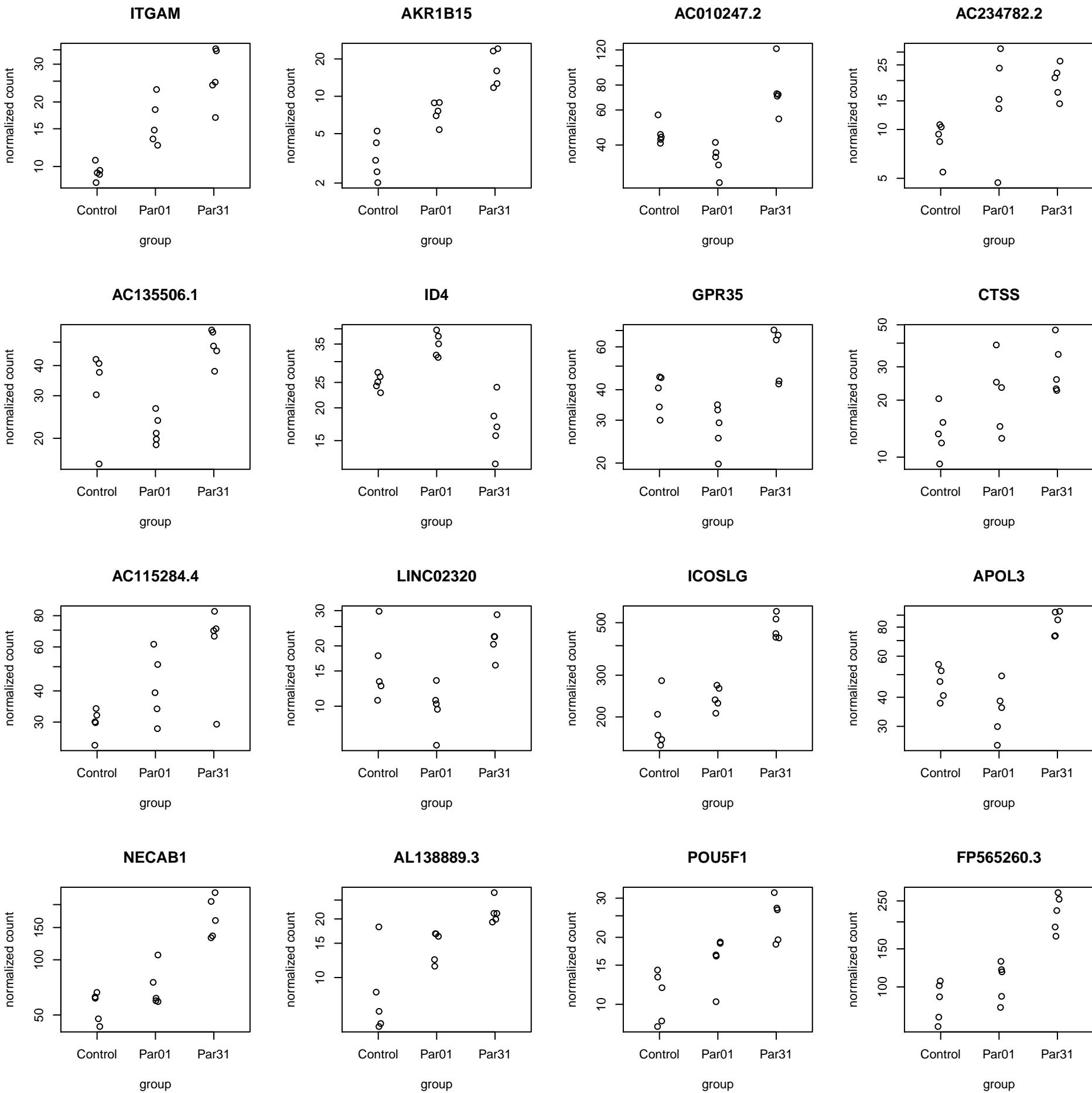


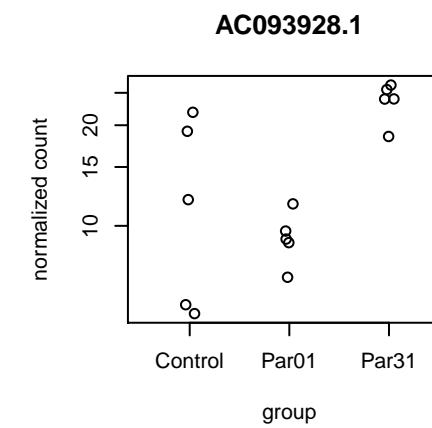
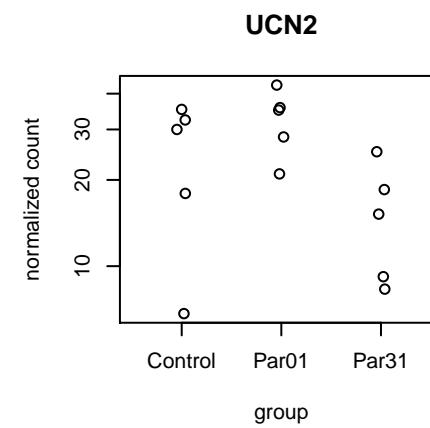
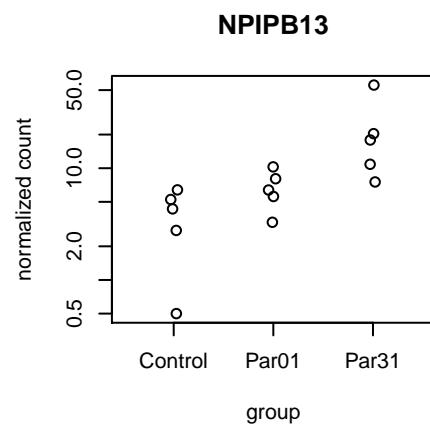
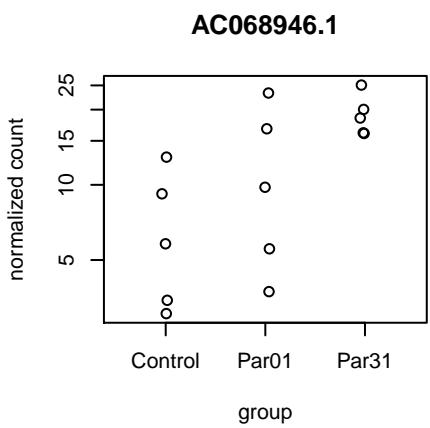
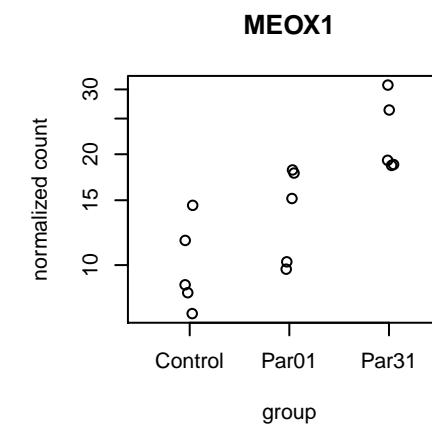
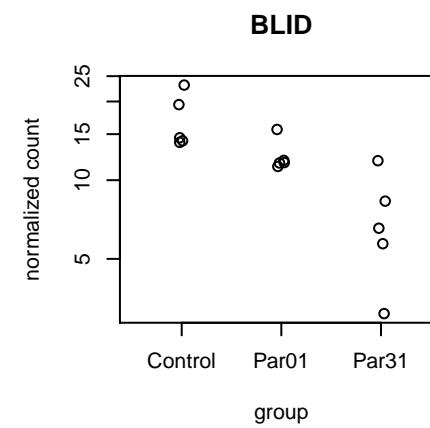
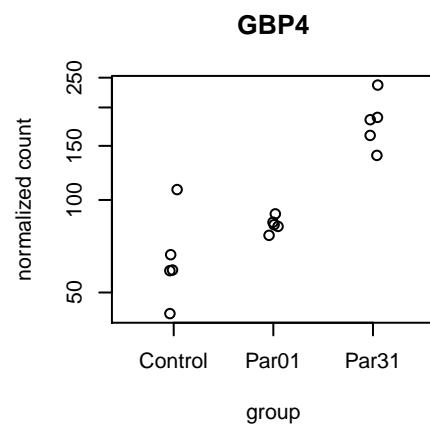
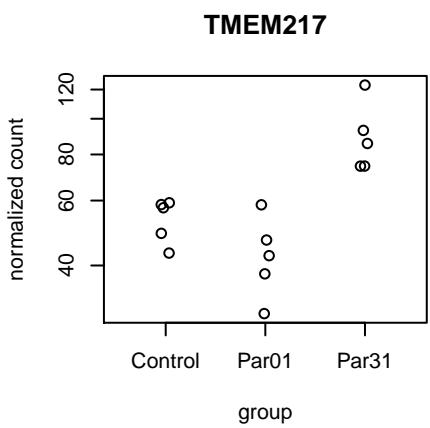
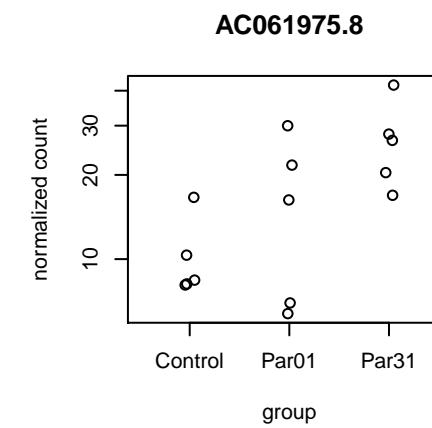
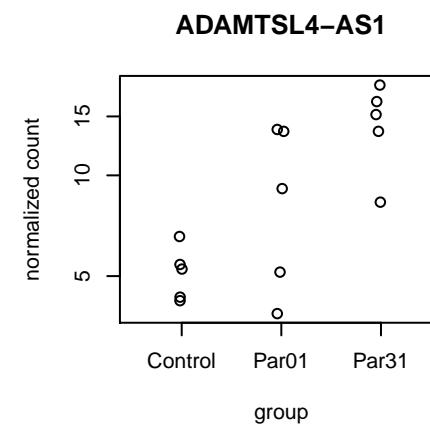
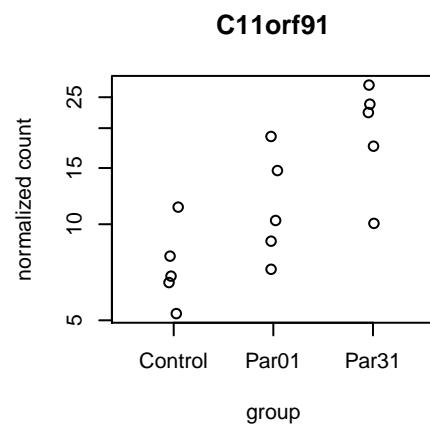
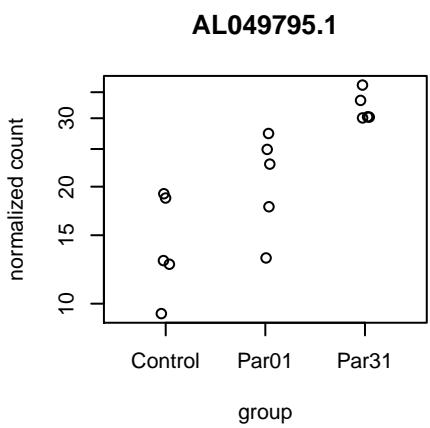
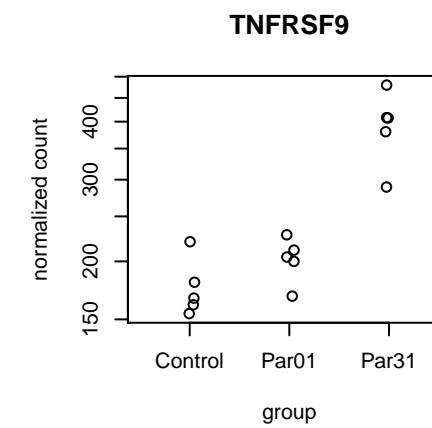
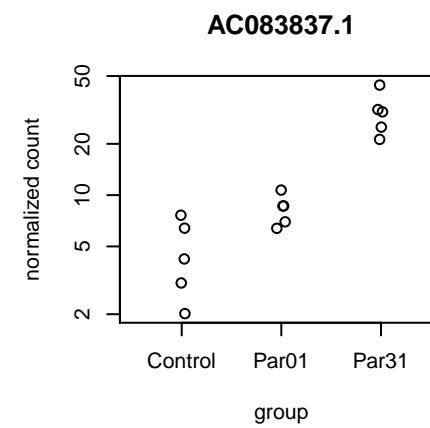
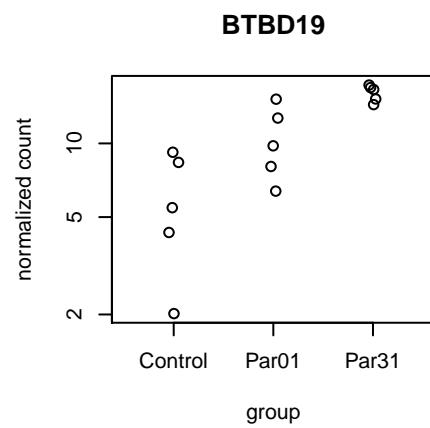
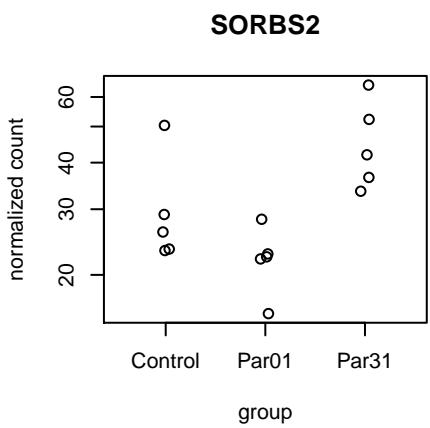
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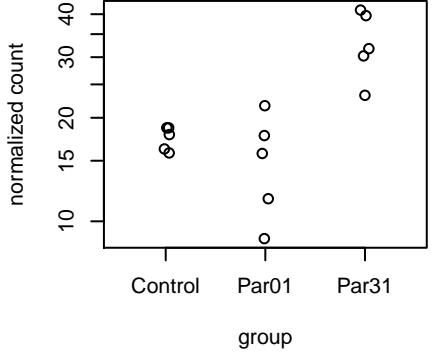
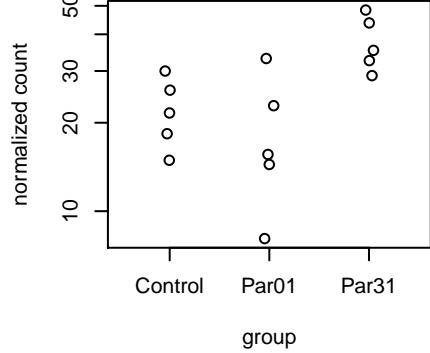
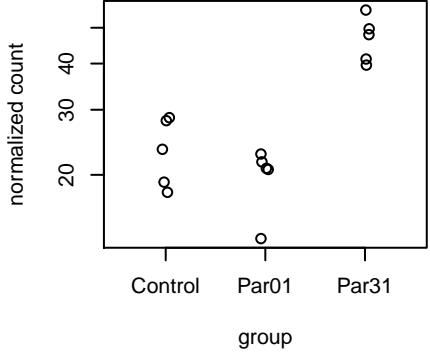
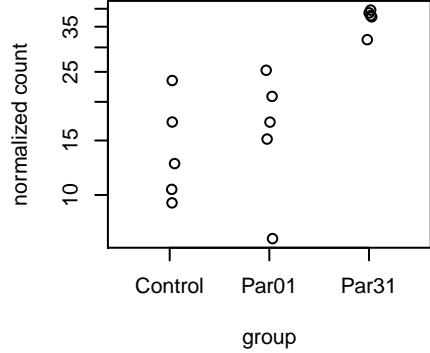
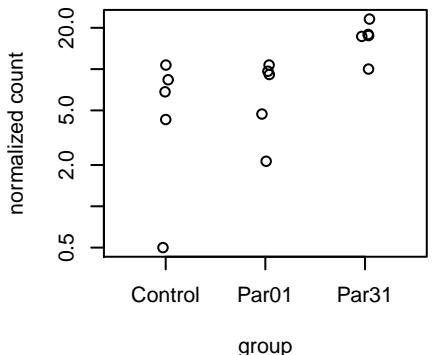
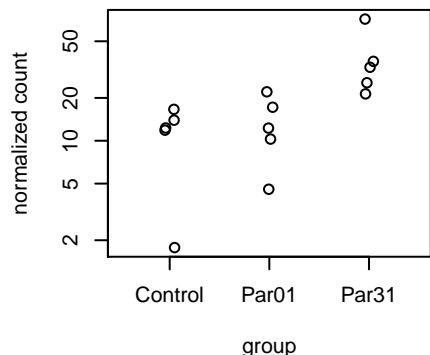
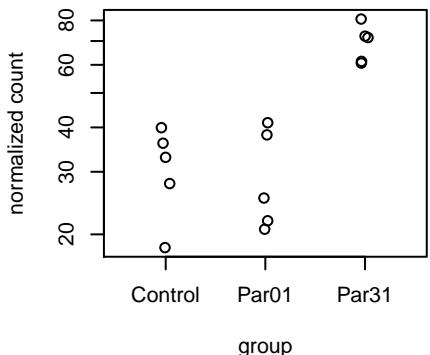
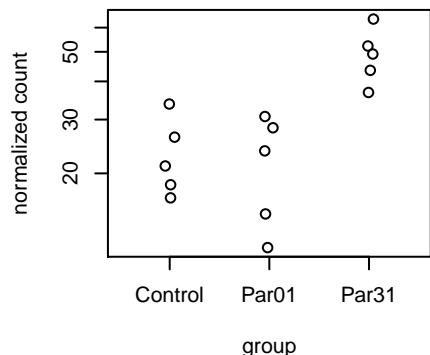
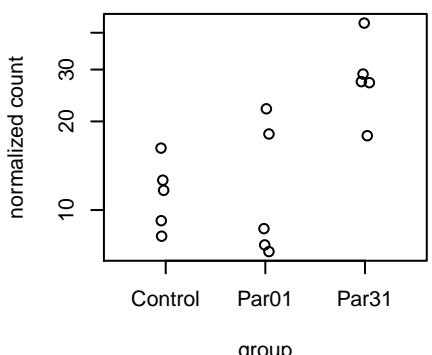
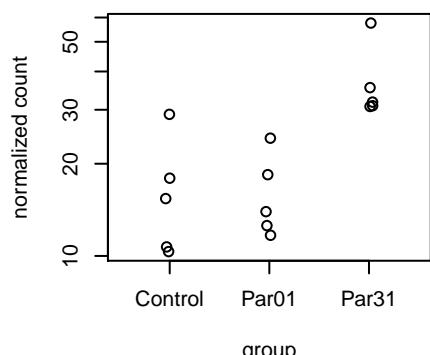
AP000944.5**SPACA6P-AS****TBX4****AC010735.1****UNC13A****TRIM31****AC107959.1****AC011379.2****UCKL1-AS1****AP001057.1****Figure S2:** Boxplots of all 298 genes differentially expressed in the PC-3 PF RNAseq experiment.



Figure S3. Experiment hardware setup. **A:** Airbus 310 aircraft used for the 34th DLR parabolic flight campaign. **B:** View in the aircraft, equipped with the experiment racks of several science teams. **C:** The rack with the incubator used for the experiment. **D:** Inside of the incubator prior to take off of the flight. **E:** The Multi Sample Incubator Centrifuge (MuSIC) of DLR (Gravitational Biology) Cologne, placed in an incubator on ground. **F:** Samples within the swing-out device on the MuSIC centrifuge during acceleration. **G:** The Vibraplex device built by DLR (Gravitational Biology), Cologne, Germany.

Table S1 Demultiplexing statistic of the 10 PF and 5 control samples.

Sample	Barcode sequence	PF Clusters	% Perfect barcode	% One mismatch barcode	Yield (Mbases)	% PF Clusters	% >= Q30 bases	Mean Quality Score
1P 1	GCCAAT	54,026,813	98.33	1.67	10,913	100	93.96	35.94
1P 2	CAGATC	65,954,683	98.59	1.41	13,323	100	93.68	35.87
1P 3	ACTTGA	59,237,051	98.95	1.05	11,966	100	93.85	35.92
1P 4	GATCAG	50,967,453	98.98	1.02	10,295	100	94.05	35.95
1P 5	TAGCTT	32,965,164	98.95	1.05	6,659	100	93.54	35.85
31P 1	GGCTAC	61,988,136	99.24	0.76	12,522	100	93.25	35.8
31P 2	CTTGTA	42,205,855	98.4	1.6	8,526	100	93.36	35.82
31P 3	AGTCAA	67,427,596	98.51	1.49	13,620	100	93.49	35.86
31P 4	AGTTCC	64,146,346	98.84	1.16	12,958	100	93.37	35.81
31P 5	ATGTCA	56,824,184	98.38	1.62	11,478	100	93.14	35.79
Control 1	ACAGTG	42,430,335	98.87	1.13	8,571	100	93.92	35.93
Control 2	ATCACG	58,111,319	98.8	1.2	11,738	100	91.96	35.49
Control 3	CGATGT	82,935,298	98.83	1.17	16,753	100	94.14	35.98
Control 4	TTAGGC	42,112,474	99.13	0.87	8,507	100	93.29	35.82
Control 5	TGACCA	67,660,339	98.73	1.27	13,667	100	93.53	35.85

Table S3. Primer selection for the 32 genes and the housekeeper gene 18S in qPCR

Factor	Primer Name	Sequence 5'-3'
18S rRNA	18s-F	GGAGCCTGCGGCTTAATT
	18s-R	CAACTAAGAACGCCATGCA
Actin beta (<i>ACTB</i>)	ACTB-F	TGCCGACAGGATGCAGAAG
	ACTB-R	GCCGATCCACACGGAGTACT
RAC alpha serine / threonine protein kinase (<i>AKT1</i>)	AKT1-F	CTTCTATGGCGCTGAGATTGTG
	AKT1-R	CAGCATGAGGTTCTCCAGCTT
Caspase 3 (<i>CASP3</i>)	CASP3-F	CTCCAACATCGACTGTGAGAAGTT
	CASP3-R	GCGCCAGCTCCAGCAA
Caspase 8 (<i>CASP8</i>)	CASP8-F	TGCAAAAGCACGGGAGAAAG
	CASP8-R	CTCTTCAAAGGTCGTGGTCAAAG
Caspase 9 (<i>CASP9</i>)	CASP9-F	CTCCAACATCGACTGTGAGAAGTT
	CASP9-R	GCGCCAGCTCCAGCAA
Caveolin 2 (<i>CAV2</i>)	CAV2-F	GATCCCCACCCGGCTAAC
	CAV2-R	CACCGGCTCTGCGATCA
Cadherin-1 (<i>CDH1</i>)	CDH1-F	GCTGGACCAGAGAGAGTTCC
	CDH1-R	CAGCTGTTGCTGTTGTGCTT
Collagen 1 alpha 1 (<i>COL1A1</i>)	COL1A1-F	ACGAAGACATCCCACCAATCAC
	COL1A1-R	CGTTGTCGCAGACGCAGAT
Endothelial growth factor (<i>EGF</i>)	EGF-F	TGCCAGCTGCACAAATACAGA
	EGF-R	TCTTACGGAATAGTGGTGGTCATC
Endothelial growth factor receptor (<i>EGFR</i>)	EGFR-F	TTGCCGCAAAGTGTGTAACG
	EGFR-R	GAGATGCCACTGATGGAGG
Ezrin (<i>EZR</i>)	EZR-F	GCAATCCAGCCAAATACAAC
	EZR-R	CCACATACTGGAGGCCAAAGTAC
Vascular endothelial growth factor receptor 2 (<i>FLK1</i>)	FLK1-F	TCTTCTGGCTACTTCTTGTACATC
	FLK1-R	GATGGACAAGTAGCCTGTCTTCAGT
Vascular endothelial growth factor receptor 1 (<i>FLT1</i>)	FLT1-F	CCCTCGCCGGAAGTTGTAT
	FLT1-R	GATAATTAACGAGTAGCCACGAGTC
Fibronectin (<i>FN1</i>)	FN1-F	AGATCTACCTGTACACCTTGAATGACA
	FN1-R	CATGATACCAGCAAGGAATTGG
Interleukin 6 (<i>IL6</i>)	IL6-F	CGGGAACGAAAGAGAGAACGCTCTA
	IL6-R	GAGCAGCCCCAGGGAGAA
Interleukin 8 (<i>CXCL8</i>)	IL8-F	TGGCAGCCTCCTGATTCT
	IL8-R	GGGTGGAAAGGTTGGAGTATG
Cytokeratin 8 (<i>KRT8</i>)	KRT8-F	GATCTCTGAGTGAAACCGGAACA
	KRT8-R	GCTCGGCATCTGCAATGG
Laminin alpha 3 (<i>LAMA3</i>)	LAMA3-F	AAAGCAAGAAGTCAGTCAGC
	LAMA3-R	TCCCATGAAGACCATCTCGG
Laminin beta 2 (<i>LAMB2</i>)	LAMB2-F	TGCTCATGGTCAATGCTAATCTG
	LAMB2-R	TCTATCAATCCTCTCCTGGACAA
Matrix metalloproteinase 9 (<i>MMP9</i>)	MMP9-F	CCTGGAGACCTGAGAACCAATC
	MMP9-R	TTCGACTCTCCACGCATCTCT
Moesin (<i>MSN</i>)	MSN-F	GAAATTGTCATCAAGCCATTG
	MSN-R	CCATGCACAAGGCCAAGAT
Mechanistic target of	MTOR-F	ATCTTGGCCATAGCTAGCCTC

rapamycin (<i>MTOR</i>)	MTOR-R	ACAACTGGGTCAATTGGAGGG
Osteopontin (<i>OPN/SPP1</i>)	OSP-F	CGAGGTGATAGTGTGGTTATGGA
	OSP-R	CGTCTGTAGCATCAGGGTACTG
Plasminogen activator inhibitor 1 (<i>PAI1</i>)	PAI1-F	AGGCTGACTTCACGAGTCTTCA
	PAI1-R	CACTCTCGITCACCTCGATCTC
Phosphatidylinositol 4,5 bisphosphate 3 kinase catalytic subunit beta (<i>PIK3CB</i>)	PIK3CB-F	AGAAAAGTTGGCCGGTIC
	PIK3CB-R	GCAGTCAACATCAGCGCAA
Radixin (<i>RDX</i>)	RDX-F	GAAAATGCCAACCAATCAA
	RDX-R	GTATTGGGCTGAATGGCAAATT
Transforming growth factor beta (<i>TGFB1</i>)	TGFB1-F	CACCCGCGTGCTAATGGT
	TGFB1-R	AGAGCAACACGGGTTAGGTA
TIMP metallopeptidase inhibitor 1 (<i>TIMP1</i>)	TIMP1-F	GCCATCGCCCCAGATC
	TIMP1-R	GCTATCAGCCACAGCAACAACA
Talin 1 (<i>TLN1</i>)	TLN1-F	GATGGCTATTACTCAGTACAGACAATGA
	TLN1-R	CATAGTAGACTCCTCATCTCCTTCCA
Tubulin beta (<i>TUBB</i>)	TUBB-F	CTGGACCGCATCTCTGTACTAC
	TUBB-R	GACCTGAGCGAACAGAGTCCAT
Vinculin (<i>VCL</i>)	VCL-F	GTCTCGGCTGCTCGTATCTT
	VCL-R	GTCCACCAGCCCTGTCATTT
Vascular Endothelial Growth Factor A (<i>VEGFA</i>)	VEGFA	CTACCTCCACCATGCCAAGTG
	VEGFA	GCGCTGATAGACATCCATGAAC