Tissue guidance without filopodia

Andrin Wacker, Holger Gerhardt & Li-Kun Phng

To cite this article: Andrin Wacker, Holger Gerhardt & Li-Kun Phng (2014) Tissue guidance without filopodia, Communicative & Integrative Biology, 7:3, e28820, DOI: 10.4161/cib.28820

To link to this article: https://doi.org/10.4161/cib.28820

Copyright © 2014 Landes Bioscience

Published online: 09 Apr 2014.

Submit your article to this journal

Article views: 317

View related articles

View Crossmark data

Citing articles: 6 View citing articles
Tissue guidance without filopodia

Andrin Wacker1,2,†, Holger Gerhardt1,2,3, and Li-Kun Phng1,2*

1Department of Oncology; Vascular Patterning Laboratory; VRC; Leuven, Belgium; 2VIB; Vascular Patterning Laboratory; VRC; Leuven, Belgium; 3Vascular Biology Laboratory; London Research Institute; Cancer Research UK; London, UK

*Current address: Swiss Center for Regenerative Medicine; University Hospital Zurich and University of Zurich; Zurich, Switzerland

Keywords: cell guidance, cell migration, endothelial tip cell, filopodia, neuronal pathfinding, vascular patterning

Filopodia are highly dynamic, rod-like protrusions that are found in abundance at the leading edge of migrating cells such as endothelial tip cells and at axonal growth cones of developing neurons. One proposed function of filopodia is that of an environmental probe, which serves to sense guidance cues during neuronal pathfinding and blood vessel patterning. However, recent studies show that tissue guidance occurs unhindered in the absence of filopodia, suggesting a dispensability of filopodia in this process. Here, we discuss evidence that support as well as dispute the role of filopodia in guiding the formation of stereotypic neuronal and blood vessel patterns.

Many biological processes depend on the guided migration of single cells or cell collectives. Directed migration is based on gradients of attractive or repulsive molecular cues that are either diffusible or surface bound. Guidance mechanisms share some characteristic hallmarks that distinguish them from tissue patterns generated by random processes. One of the most striking hallmarks resulting from directed migration is stereotypic tissue patterns such as that observed in neuronal and blood vascular networks. However, the final pattern is highly adapted to function, where neuronal activity fine-tunes synaptic connectivity and blood flow shapes the blood vessel pattern.

Axonal growth cones navigate to their target over large distances with a precision and timing that is highly conserved from animal to animal. Modulation of attractive and repulsive gradients leads to erroneous pathfinding with misprojections. For example, Netrin is an axonal guidance cue acting from the neural tube midline that, if presented from an ectopic source, causes axonal growth cone turning toward the ectopic site of presentation within explant cultures. Furthermore, its genetic deletion induces misguidance in vivo. Although it is difficult to experimentally show protein gradients of diffusible cues, a tissue gradient of Netrin in the developing neural tube has been described.

The blood vascular network is also highly organized and displays branching patterns that are very similar between animals. This is particularly true for major arteries and veins. Live imaging of intersegmental vessel (ISV) development in the zebrafish embryo reveals that directed migration of endothelial cells lays the foundation of the vascular network (Fig. 1A).

Ectopic expression of the vascular cue vascular endothelial growth factor, VEGF-A, in muscle cells leads to altered ISV trajectories from one side of the trunk to the other (Fig. 1B), suggesting misguidance of the vessels. This observation suggests that physical barriers provided by tissues are insufficient for steering vessel patterning and that guidance cues may direct endothelial cell migration. Indeed, the VEGF, Netrin1/Un5B, and Semaphorin/PlexinD1 signaling pathways regulate ISV patterning.

In mouse hindbrain and retina, VEGF-A mRNA production shows a tissue gradient and there is some evidence for graded protein distribution in the hindbrain. The c-terminal heparin-binding domain of VEGF-A is important for VEGF-A protein distribution such that its deletion results in more diffused VEGF-A distribution, perturbed vessel morphology and mispatterning of the vasculature. These data suggest that extracellular VEGF-A resembles axonal guidance cues by steering vascular sprouts by means of a protein gradient. This concept is further supported by the presence of a gradient of internalized VEGF-A and VEGF-C in the developing retina vasculature, with endothelial cells at the migrating front displaying the highest level of endocytosed VEGF protein. Although a gradient of VEGF-A protein has not been demonstrated in the zebrafish embryo, indirect evidence exists. For instance, loss of soluble decoy receptor sFlt1 (VEGFR1) expression in Plexin-D1 mutants leads to ISV misprojections, presumably via the modulation of VEGF-A availability and therefore gradient formation. sFlt1 has also been proposed to locally shape the VEGF gradient in the mouse retina and thereby ensures appropriate angles of vascular sprouting. In addition, the CXCL12-CXCR4 chemokine signaling pathway plays an essential role in the patterning of arteries and nerve-artery alignment in the skin of the embryonic mouse limb. Hence, it seems likely that vascular sprouts, like axonal growth cones, are guided by gradients of diffusible cues. However, the mechanism of how blood vessels integrate signaling cues to form a hierarchical network of vessels is as yet unknown.

Filopodia are highly dynamic, rod-like protrusions that are found in abundance at the leading edge of migrating cells such as endothelial tip cells and at axonal growth cones of developing neurons. One proposed function of filopodia is that of an environmental probe, which serves to sense guidance cues during neuronal pathfinding and blood vessel patterning. However, recent studies show that tissue guidance occurs unhindered in the absence of filopodia, suggesting a dispensability of filopodia in this process. Here, we discuss evidence that support as well as dispute the role of filopodia in guiding the formation of stereotypic neuronal and blood vessel patterns.

Many biological processes depend on the guided migration of single cells or cell collectives. Directed migration is based on gradients of attractive or repulsive molecular cues that are either diffusible or surface bound. Guidance mechanisms share some characteristic hallmarks that distinguish them from tissue patterns generated by random processes. One of the most striking hallmarks resulting from directed migration is stereotypic tissue patterns such as that observed in neuronal and blood vascular networks. However, the final pattern is highly adapted to function, where neuronal activity fine-tunes synaptic connectivity and blood flow shapes the blood vessel pattern.

Axonal growth cones navigate to their target over large distances with a precision and timing that is highly conserved from animal to animal. Modulation of attractive and repulsive gradients leads to erroneous pathfinding with misprojections. For example, Netrin is an axonal guidance cue acting from the neural tube midline that, if presented from an ectopic source, causes axonal growth cone turning toward the ectopic site of presentation within explant cultures. Furthermore, its genetic deletion induces misguidance in vivo. Although it is difficult to experimentally show protein gradients of diffusible cues, a tissue gradient of Netrin in the developing neural tube has been described.

The blood vascular network is also highly organized and displays branching patterns that are very similar between animals. This is particularly true for major arteries and veins. Live imaging of intersegmental vessel (ISV) development in the zebrafish embryo reveals that directed migration of endothelial cells lays the foundation of the vascular network (Fig. 1A).

Ectopic expression of the vascular cue vascular endothelial growth factor, VEGF-A, in muscle cells leads to altered ISV trajectories from one side of the trunk to the other (Fig. 1B), suggesting misguidance of the vessels. This observation suggests that physical barriers provided by tissues are insufficient for steering vessel patterning and that guidance cues may direct endothelial cell migration. Indeed, the VEGF, Netrin1/Un5B, and Semaphorin/PlexinD1 signaling pathways regulate ISV patterning.

In mouse hindbrain and retina, VEGF-A mRNA production shows a tissue gradient and there is some evidence for graded protein distribution in the hindbrain. The c-terminal heparin-binding domain of VEGF-A is important for VEGF-A protein distribution such that its deletion results in more diffused VEGF-A distribution, perturbed vessel morphology and mispatterning of the vasculature. These data suggest that extracellular VEGF-A resembles axonal guidance cues by steering vascular sprouts by means of a protein gradient. This concept is further supported by the presence of a gradient of internalized VEGF-A and VEGF-C in the developing retina vasculature, with endothelial cells at the migrating front displaying the highest level of endocytosed VEGF protein. Although a gradient of VEGF-A protein has not been demonstrated in the zebrafish embryo, indirect evidence exists. For instance, loss of soluble decoy receptor sFlt1 (VEGFR1) expression in Plexin-D1 mutants leads to ISV misprojections, presumably via the modulation of VEGF-A availability and therefore gradient formation. sFlt1 has also been proposed to locally shape the VEGF gradient in the mouse retina and thereby ensures appropriate angles of vascular sprouting. In addition, the CXCL12-CXCR4 chemokine signaling pathway plays an essential role in the patterning of arteries and nerve-artery alignment in the skin of the embryonic mouse limb. Hence, it seems likely that vascular sprouts, like axonal growth cones, are guided by gradients of diffusible cues. However, the mechanism of how blood vessels integrate signaling cues to form a hierarchical network of vessels is as yet unknown.
Is There a Role for Filopodia in Cell Guidance?

Guided cell migration requires the generation of polarized plasma membrane protrusions such as filopodia, lamellipodia, blebs and invadopodia. Although these protrusions differ structurally and in their composition, they each contribute to cell movement by pushing the leading edge forward. Both axonal growth cones and endothelial tip cells display abundant filopodia protrusions (Fig. 2). Their dynamic behavior of extension and retraction is reminiscent of antennae probing their environment and are generally thought to serve as environmental sensors that integrate extracellular signals during directed migration. Growth cone morphology, reflected for instance by filopodia number, has been correlated with guidance decisions at choice points\(^\text{21-24}\) and filopodia preferentially contact target cells, sometimes over long distance.\(^\text{23,25}\) In vivo experiments with titrated doses of an inhibitor of F-actin polymerization, cytochalasin B, blocked growth cone filopodia formation in grasshopper T11 neurons and in Xenopus retinal ganglion cell neurons.\(^\text{26,27}\) While pathfinding was disrupted in conditions without filopodia, only minor effects were seen on axonal extension, suggesting that filopodia are dispensable for growth but essential for guidance. Similarly, in vitro treatment with cytochalasin B abolished glutamate-induced growth cone turning with only slight effects on neurite extension.\(^\text{28}\) However, a little known study on retinal ganglion cells suggests that axon guidance can occur without filopodia.\(^\text{29}\) Here, the authors demonstrate that growth cones extend few or no filopodia after inhibiting Ena/VASP function.\(^\text{29}\) While axonal elongation was slowed down, retinal ganglion cell axons devoid of filopodia showed normal trajectories in vivo and growth cone navigation across several choice points was unaffected. Thus, the role of filopodia in mediating growth cone navigation is unclear.

Similar to growth cones, endothelial tip cells at the leading edge of vascular sprouts produce many long filopodia that extend toward the direction of vascular growth during angiogenesis. These filopodia express the VEGF-A receptor, VEGFR2, and become misdirected and shorter upon disruption of VEGF-A distribution in the mouse retina.\(^\text{16}\) As filopodia have been proposed to act as sensors of the extracellular milieu and endothelial cells respond to axon guidance molecules such as Slits and Roundabouts, Netrins and Unc5 receptors, Semaphorins, Plexins and Neuropilins, and Ephrins and Eph receptors,\(^\text{30}\) it has been widely assumed that they sense and integrate pro-angiogenic and repulsive cues in tip cells to enable guided migration and stereotypic vessel patterning. However, the role of filopodia in vessel guidance has never been proven nor questioned.

Recently, work from our laboratory demonstrated that filopodia are not essential for mediating endothelial tip cell guidance.\(^\text{31}\) By using low concentrations of Latrunculin B

---

**Figure 1.** Intersegmental vessels form a highly stereotypic vessel pattern in the zebrafish embryo (A). Misexpression of Vegfa165 in the embryonic trunk leads to misguidance of an ISV (arrows) toward cells expressing ectopic Vegfa165 (green, arrowheads) (B). DA, dorsal aorta; ISV, intersegmental vessel. Scale bars, 20µm. Plasmid encoding Vegfa165 is a gift from Nathan Lawson.

**Figure 2.** Axonal growth cones (A, image courtesy of Isabelle Brunet) and endothelial tip cells (B) extend long filopodia (arrowheads) in the direction of migration. Scale bars, 10µm.
Filopodia are Dispensable for Tip Cell Guidance

In summary, our study shows that during angiogenesis, endothelial filopodia are dispensable for tip cell guidance. This finding complements that of Dwivedy et al., who showed that filopodia are also not essential for axonal growth cone navigation and challenges the long-standing notion that filopodia are required for guided migration. In fact, filopodia or filopodia-like structures have been ascribed many other functions. These include facilitating cell-cell matching and epithelial sheet adherence during dorsal closure in Drosophila, transmitting signals such as Delta-Notch and Sonic Hedgehog signaling between non-neighboring cells, inducing cell shape changes required for preimplantation embryonic development by providing tension and positioning nuclei in nurse cells during oogenesis in Drosophila. In endothelial cells, we propose that filopodia serve as templates from which lamellipodia emerge and that both protrusive structures coordinate to allow efficient migration and expansion of new vascular sprouts. In addition, tip cell filopodia facilitate the process of anastomosis, a process whereby tip cells meet, fuse and establish new junctions to form a connected vascular network. Thus, the mechanism(s) by which blood vessels are guided by extracellular cues is still unresolved.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

References
23. Bovolenta P, Mason C. Growth cone morphology varies with position in the developing mouse visual pathway from retina to first targets. J Neurosci 1987; 7:1447-60; PMID:3572487
31. Phng LK, Stanchi F, Gerhardt H. Filopodia are dispensable for endothelial tip cell guidance. Development 2013; 140:4031-40; PMID:24046339; http://dx.doi.org/10.1242/dev.097352
35. Sanders TA, Llagostera E, Barna M. Specialized filopodia direct long-range transport of SHH during vertebrate tissue patterning. Nature 2013; 497:628-32; PMID:23624372; http://dx.doi.org/10.1038/nature12457