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Supplemental information

A computational model of the DNA damage-induced

IKK/ NF-κB pathway reveals a critical dependence

on irradiation dose and PARP-1

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Fig S1. Experimental data and simulations of calibrated model, related to Figure 1 and STAR Methods.

Dots represent experimental data and the yellow line represents simulation data. The grey area surrounding the yellow line shows the fitted standard deviation. Otherwise the standard deviation is represented by grey bars which is true for the MRN and ATM recruitment data. For the recruitment data of the PARP-1 WT and mutant two similar data sets exist which differ in the temporal resolution and time period. For those experiments, the irradiation dose is not available (n/a). For 0.8, 12, and 17 Gy for the recruitment data of the MRN complex, additional biological replicates exist. Details about the experimental data and settings are given in STAR methods. The estimated parameter values for the best fit are shown in Table S1. Abbreviation: sig: signalosome.



Fig S2. Influence of irradiation dose and PARP-1 inhibition on cumulated amount of parPARP, related to Figure 4 and Figure 5.

The cumulated amount of parPARP was quantified by integrating the flux of PARylated PARP-1 dissociating from DNA (corresponding parameter: koff_act) in the time frame of signalosome formation for various irradiation doses with and without 90% inhibition of PARylation (corresponding parameter: vact).



Fig S3. Profile likelihoods for estimated model parameters, related to STAR Methods.

The black star represents the estimated parameter value of the best fit and the corresponding fit quality given as negative log likelihood. To compute the profile likelihood, the specified parameter is fixed to an increased or decreased value compared to the calibrated value and the remaining parameters are refitted. The resulting log likelihood is represented by a grey x. To identify the 95% confidence interval of a parameter, a threshold for the log likelihood is calculated based on a χ^2 distribution with one degree of freedom (dashed red line).

log₁₀ label values of description log₁₀ value boundaries -7.0 m⁻²·s⁻¹ [-10, -2] SFM k1 formation of signalosome SFM k2 -1.3 s⁻¹ [-10, 3] dissociation of signalosome poly-ubiquitination of TRAF6 and -4.5 m⁻¹.s⁻¹ TM k1 [-10, 3] binding of pATM activation of TAK1 and recruitment -10.2 m⁻¹·s⁻¹ TM k2 [-12, 3] to TRAF6 complex integration of IKKy into IKK complex, -3.8 m⁻¹·s⁻¹ mono-ubiguitination of IKKy and TM k3 [-10, 3] phosphorylation of IKKβ 3.9 m total concentration of ATM ATM_tot [2, 7] [2, 7] IKK_Y_tot 5.0 m total concentration of IKKy MRN_tot 6.9 [2, 7] total concentration of MRN m PARP1 tot 5.3 m [-, -] total concentration of PARP-1 [2, 7] total concentration of TAK1 TAK1 tot 6.2 m TRAF6 tot 2.0 m [2, 7] total concentration of TRAF6 positive feedback of chromatin--9.0 m⁻¹·s⁻¹ k1 [-10, 3] bound MRN on MRN recruitment k2 -5.2 m⁻¹·s⁻¹ [-10, 3] recruitment of ATM to MRN positive feedback of pATM on k3 -3.2 s⁻¹ [-10, 1] activation of ATM -9.5 m⁻¹·s⁻¹ k4 recruitment of MRN to DNA lesion [-10, 3] activation of ATM and its k5 -3.4 s⁻¹ [-10, 1] dissociation from MRN k DNAb $3.6 \text{ m} \cdot \text{s}^{-1}$ generation of binding sites [2, 4] 0.0 s⁻¹ kdepar dePARylation of PARP-1 [-4, 0] parameter of Hill term for positive 3.8 m² km [-4, 9] feedback of MRN parameter of Hill term for positive 1.9 m² km2 [-4, 9] feedback of ATM dissociation of PARP-1 from DNA koff -1.4 s⁻¹ [-10, 3] lesion dissociation of PARP-1 from 2.9 s⁻¹ koff2 [-10, 3] periphery of lesion dissociation of PARylated PARP-1 koff act 1.1 s⁻¹ [-10, 3] from chromatin -8.9 m⁻¹·s⁻¹ kon [-9,-2] binding of PARP-1 to lesion

Table S1. Parameter description with estimated parameter values, related to STAR methods.

kr	-2.1 m ^{-1.} s ⁻¹	[-9, 3]	PARylated PARP-1-induced recruitment of PARP-1 to periphery of lesions
n	log10(2)	[-, -]	Hill coefficient for positive feedback of MRN and ATM
vact	0.8 s ⁻¹	[-0.4, 1]	automodification of PARP-1

Abbreviations: m: molecules, s: seconds