MCT1 is a predictive marker for lenalidomide maintenance therapy in multiple myeloma

Jacob Stroh,1,2,* Anja Seckinger,3,4,* Michael Heider,1,2,* Martina Rudelius,5 Ruth Eichner,1,2 Markus Schick,6 Jolanta Slawska,1,6 Martina Emde-Rajaratanam,3,4 Hans Salwender,7-9 Uta Bertsch,10 Hartmut Goldschmidt,10,11 Katja Weisel,12 Christof Scheid,1,3,14 Ulrich Keller,6,15-17 Dirk Hose,3,4,† and Florian Bassermann1,2,15,16,†

1Department of Medicine III, Klinikum rechts der Isar, and 2TranslaTUM, Center for Translational Cancer Research, Technical University of Munich, Munich, Germany; 3Department of Hematology and Immunology, Myeloma Center Brussels, and 4Labor f€ur Myelomforschung, Vrije Universiteit Brussel (VUB), Jette, Belgium; 5Institute of Pathology, Ludwig Maximilians University, Munich, Germany; 6Department of Hematology, Oncology, and Tumor Immunology, Charit€-Universitatsmedizin, Berlin, Germany; 7Asklepios Tumorzentrum Hamburg, Hamburg, Germany; 9AK Altona, Hamburg, Germany; 9AK St. Georg, Hamburg, Germany; 10Department of Internal Medicine V, and 11National Center for Tumor Diseases (NCT), Heidelberg University Hospital, Heidelberg, Germany; 12Department of Medicine II, University Medical Center Hamburg-Eppendorf, Hamburg, Germany; 13Department of Internal Medicine I, and 14Center of Integrated Oncology Cologne Bonn, University of Cologne, Cologne, Germany; 15German Cancer Consortium (DKTK), Heidelberg, Germany; 16German Cancer Research Center (DKFZ), Heidelberg, Germany; and 17Max-Delbr€uck Center for Molecular Medicine, Berlin, Germany

Biomarkers that predict response to lenalidomide maintenance therapy in patients with multiple myeloma (MM) have remained elusive. We have shown that immunomodulatory drugs (IMiDs) exert anti-MM activity via destabilization of MCT1 and CD147. In this study, cell samples of 654 patients with MM who received lenalidomide (n = 455), thalidomide (n = 98), or bortezomib (n = 101) maintenance were assessed by gene expression profiling and RNA sequencing, followed by correlation of MCT1 and CD147 expression with data for progression-free survival (PFS) and overall survival (OS). Patients with high expression levels of MCT1 showed significantly reduced PFS (31.9 months vs 48.2 months in MCT1 high vs MCT1 low; P = .03) and OS (75.9 months vs not reached [NR] in MCT1 high vs MCT1 low; P = .001) in cases with lenalidomide maintenance, whereas MCT1 expression had no significant impact on PFS or OS in cases with bortezomib maintenance. We validated the predictive role of MCT1 for IMiD-based maintenance in an independent cohort of patients who received thalidomide (OS, 83.6 months vs NR in MCT1 high vs MCT1 low; P = .03). Functional validation showed that MCT1 overexpression in human MM cell lines significantly reduced the efficacy of lenalidomide, whereas no change was observed with bortezomib treatment, either in vitro or in a MM xenograft model. Our findings have established MCT1 expression as a predictive marker for response to lenalidomide-based maintenance in patients with MM.

Introduction

Multiple myeloma (MM) is a heterogenous clonal plasma cell malignancy that accounts for ~10% of all hematologic malignancies.1 The introduction of novel substances, such as the immunomodulatory drugs

*J.S., A.S., and M.H. contributed equally to this study.
†D.H. and F.B. contributed equally to this study.
The expression data were deposited in ArrayExpress (accession numbers E-MTAB-4715, E-MTAB-4717, E-MTAB-5212, E-TABM-937, and E-TABM-1088).

Requests for data sharing may be submitted to Florian Bassermann (florian.bassermann@tum.de).
The full-text version of this article contains a data supplement.
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Figure 1. Expression of MCT1 determines PFS and OS in patients receiving IMiD-based maintenance therapy. (A) MCT1 gene expression in plasma cell disorders (n = 1486) and normal bone marrow plasma cells (BMPC, n = 19). MGUS, monoclonal gammopathy of unknown significance; AMM, asymptomatic MM; MM, previously untreated, therapy-requiring MM; MMR, relapsed/refractory MM; and HMCL, human myeloma cell line. Numbers in blue (red) indicate the number of patients expressing (not expressing) MCT1. (B-E) Landmark analysis of patients with MM who underwent maintenance treatment after autologous stem cell transplantation. Expression of MCT1 was assessed by gene expression profiling in CD138-purified myeloma cell samples and correlated with PFS and OS data. P-values are log-rank. (B) PFS of 455 patients who underwent lenalidomide maintenance therapy in the GMMG-MM5-trial (31.9 months vs 48.2 months; MCT1$^{high}$ vs MCT1$^{low}$). (C) OS of 455 patients who underwent lenalidomide maintenance therapy in the GMMG-MM5-trial (75.9 months vs not reached months; MCT1$^{high}$ vs MCT1$^{low}$). (D) PFS of 62 patients who underwent bortezomib maintenance therapy in the GMMG-MM4-trial (39.8 months vs 32.6 months; MCT1$^{high}$ vs MCT1$^{low}$). (E) OS of 101 patients who underwent bortezomib maintenance therapy in the GMMG-MM4-trial, 125.8 months vs 129.8 months; MCT1$^{high}$ vs MCT1$^{low}$).
Figure 2. Overexpression of MCT1 attenuates lenalidomide cytotoxicity in vitro and in vivo. (A-B) Cell proliferation analysis of MM1S (A) and U266 (B) cells, which were infected with lentivirus via control constructs (empty vector [EV]) or constructs to induce MCT1 or CD147 expression, treated with dimethyl sulfoxide (DMSO) or 10 μM lenalidomide for 72 h. (C) Quantification of tumor growth during lenalidomide or vehicle control treatment using caliper measurements (Ctrl, Len: n = 4 tumors). Xenograft tumors from U266 cells expressing MCT1 or the EV control. (D) Positron emission tomography (PET) images, using 18F-fluorodeoxyglucose, of representative immunocompromised NOD-SCID mice bearing xenograft tumors expressing MCT1 or the EV control. Images were taken after 7 days of respective treatment. Arrows
(IMIDs) lenalidomide and thalidomide, proteasome inhibitors, and monoclonal antibodies, has greatly improved treatment response and survival within the past 2 decades.1 Transplantation-eligible, fit patients undergo 1 or 2 cycles of high-dose melphalan and autologous stem cell transplantation subsequent to several cycles of induction therapy.2 Lenalidomide maintenance therapy has improved progression-free survival (PFS) in this setting; however, a beneficial effect of lenalidomide maintenance therapy on overall survival (OS) has been inconsistent in several studies.3–8 Despite the high need for predictive markers of response to lenalidomide maintenance therapy, they have remained elusive.

We have shown that IMIDs exert anti-myeloma activity via destabilization of different, mostly metabolically active transmembrane complexes by means of interfering with the quality control function of CRBN, in addition to the established degradation of neosubstrates, such as IKZF1 and IKZF3.9–12 Among these, CD147 and MCT1 form a heterodimeric transmembrane complex that functions as a transporter of monocarboxylates, such as lactate, a function of vital importance for the survival of cancer cells, particularly within the hypoxic environment of the bone marrow.13,14 MCT1 and CD147 are upregulated in MM and have been shown to be essential for proliferation and survival of MM cells.15–17 Persistence of MCT1 and CD147 expression upon lenalidomide treatment is associated with IMiD resistance in samples from patients with primary MM samples, and overexpression of the complex attenuates IMiD efficacy in MM xenograft models.9 Against this background, we investigated gene expression levels of MCT1 and CD147 in 654 patients with MM treated with lenalidomide, thalidomide, or bortezomib maintenance and found that high MCT1 expression significantly correlated with short PFS and OS in patients who receive IMiD maintenance therapy. These findings were functionally validated in human multiple myeloma cell lines (HMCLs) and in MM xenograft models.

**Methods**

CD138-purified myeloma cell samples of 1486 patients with plasma cell disorders at different stages of MM were investigated by gene expression profiling (GEP), using U133 2.0 plus DNA microarrays for expression of MCT1 (202235_at) and CD147 (208677_s_at).18,19 The presence and absence of gene expression were determined by using the presence-absence calls with negative probe sets (PANP) algorithm.20 “Low” and “high” levels of expression were determined by using maximally selected rank statistics. Gene expression from the study patients was validated by RNA sequencing (RNA-seq), which was performed as previously described.18,19,21 Of those patients, 654 received high-dose melphalan and autologous stem cell transplantation and maintenance with bortezomib (n = 101), thalidomide (n = 98), or lenalidomide (n = 455) as the first-line treatment within 2 phase 3 clinical trials of the German-speaking Myeloma Multicenter Group (GMMG) (GMMG-HD4 and GMMG-MM5; EudraCT 2004-000944-26 and 2010-019173-16). The correlation of CD147 and MCT1 with PFS and OS was determined. Survival curves and median time to progression were computed with nonparametric survival estimates for censored data using the Kaplan-Meier method. Differences between the curves were tested using the G-P log-rank test. Landmark analysis was performed by defining an alternative starting point (landmark) at 12 months. The study was in accordance with the ethical standards of the participating institutions and national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards. Written informed consent was obtained from each patient before participation in the studies.

The HMCLs MM1S (ATCC: CRL-2974) and U266 (DSMZ: ACC-9) cells were cultured in RPMI-1640 (Gibco) with 10% heat-inactivated fetal bovine serum and 1% penicillin-streptomycin. Cells with lentiviral overexpression of CD147 or MCT1 were generated as previously described.9 The cells were treated with lenalidomide (Sigma) or bortezomib (Sigma) at the concentrations indicated in the respective figure legends, and proliferation was assessed by using trypan blue exclusion.

For xenograft experiments 0.6 × 10^7 MM cells were mixed with Matrigel Basement Membrane Matrix (BD Bioscience) at a 1:1 ratio and injected subcutaneously into the opposite flanks of female NOD SCID mice at 8 to 10 weeks of age (Janvier). After tumor engraftment, the mice were randomly assigned to receive lenalidomide (30 mg/kg) in 0.5% carboxymethylcellulose and 0.25% Tween 80 in deionized water, by daily oral gavage; IV bortezomib (0.5 mg/kg every 72 h); or vehicle control and imaged by 18F-fluorodeoxyglucose positron emission tomography/magnetic resonance imaging on day 7 of treatment. Tumor growth was monitored by caliper measurements. The mice were housed in specific pathogen–free conditions, and the animal experiments were conducted with permission from the District Government of Upper Bavaria. Immunohistochemistry, cell extract preparation, and western blot analysis were performed as previously described.9,10,22

**Results and discussion**

**Expression of MCT1 determines PFS and OS in patients receiving IMiD-based maintenance therapy**

First, we performed GEP of MM samples of 1486 patient to determine MCT1 expression in plasma cell disorders at different stages of the disease. Expression was validated by RNA-seq (supplemental Figure 1A). MCT1 expression was not only elevated when compared with normal plasma cells, but increased steadily with the progression state of plasma cell disorders, with the highest levels observed in patients with relapsed MM and in MM cell lines (Figure 1A). GEP and RNA-seq correlation with PFS and OS data of 455 patients who received maintenance treatment with the IMID lenalidomide showed significantly reduced PFS (31.9 months vs 48.2 months in MCT1 high vs MCT1 low; P = .03) and OS (75.9 months vs not reached [NR] in MCT1 high vs MCT1 low; P = .001) in patients with...
high expression levels of MCT1 (Figure 1B–C). Patients treated in an independent cohort by maintenance with the IMiD thalidomide, likewise showed significantly shorter OS (83.6 vs 92.4 months in MCT1high vs MCT1low; \( P = .03 \)) with high MCT1 expression levels, and displayed a trend toward reduced PFS (34.8 vs 43.7 months in MCT1high vs MCT1low; \( P = .23 \)) that was not statistically significant (supplemental Figure 2A–B). These findings validate MCT1 as a biomarker for IMiD maintenance in an independent cohort. The somewhat lower predictive effect of MCT1 expression on thalidomide maintenance therapy, as compared with lenalidomide, corresponds well to a lower activity of this drug in terms of destabilizing MCT1.9 By contrast, MCT1 expression had no effect on PFS (39.8 vs 32.6 months in MCT1high vs MCT1low; \( P = .6 \)) and OS (125.8 vs 129.8 months in MCT1high vs MCT1low; \( P = .28 \); Figure 1D–E) in patients receiving bortezomib-based maintenance, thus underscoring a specific prognostic value of MCT1 expression on lenalidomide maintenance. MCT1 expression was not associated with high-risk chromosomal aberrations, Revised-International Staging System score, or GEP-based high-risk scores. Expression of CD147 was not predictive of the outcome of lenalidomide maintenance (supplemental Figure 3A–B). This finding is in line with a low variation of CD147 expression at the mRNA level (supplemental Figures 1B and 3C) and previous observations demonstrating a posttranslational dependency of CD147 protein expression (supplemental Figures 1B and 3C).13,16,22 It is therefore conceivable, that a dependency of lenalidomide maintenance on CD147 would become evident only on the protein level. These protein data, however, are not available for the investigated cohort of patients with MM.

Overexpression of MCT1 attenuates lenalidomide cytotoxicity in vitro and in vivo

Next, we validated the findings in cell culture models of HMCLs. Two cell lines, MM1S and U266 were infected with lentivirus to induce forced protein expression of both MCT1 or CD147 and subjected to lenalidomide or bortezomib treatment. Indeed, overexpression of MCT1 in MM cells significantly reduced sensitivity to lenalidomide treatment (Figure 2A–B), thereby functionally validating the predictive significance of MCT1. In contrast to the patient-derived data outlined thus far, high CD147 protein expression also attenuated lenalidomide efficacy in this model, again underscoring the notion that CD147 requires analysis on the protein level rather than on the mRNA level. By contrast, the efficacy of the proteasome inhibitor bortezomib was not affected by overexpression of MCT1 or CD147 (supplemental Figure 4A–B).

To confirm these findings in vivo, we established a xenograft model using U266 cells with control (empty vector) or MCT1 overexpression. After tumor engraftment mice were randomly assigned to receive lenalidomide, bortezomib, or control treatment. In line with our in vitro data, induced overexpression of MCT1 protected the tumors from the anti-myeloma activity of lenalidomide (Figure 2C–E; supplemental Figure 4D–F). Again, the cytotoxic effects of bortezomib treatment were not impaired by overexpression of MCT1 (Figure 2D–E; supplemental Figure 4C–F). Both lenalidomide and bortezomib treatment resulted in the induction of apoptosis. While the induction of apoptosis upon lenalidomide treatment was impaired in tumors with overexpression of MCT1, levels of cleaved caspase 3 remained unchanged by bortezomib treatment (Figure 2F–G).

We have identified MCT1 as a predictive marker for the efficacy of lenalidomide maintenance therapy in MM. To the best of our knowledge, MCT1 marks the first functionally validated biomarker for this therapeutic setting in MM. Based on our findings, patients with high expression levels of MCT1 have a reduced response to lenalidomide-based maintenance therapy and could benefit from other maintenance regimens, including proteasome inhibitors and CD38-targeting antibodies.

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Authorship

Contribution: F.B. and D.H. initiated the study; J. Stroh, A.S., M.H., D.H., and F.B. conceived and designed the research; J. Stroh, A.S., and M.H. performed most of the experiments and data analyses with crucial help from M.E.-R.; M.R. performed the immunohistochemical analyses; J. Sławaska helped with the xenograft experiments; H.S., U.B., H.G., K.W., and C.S. provided patient data; J. Stroh, A.S., M.H., R.E., M.S., U.K., D.H., and F.B. analyzed the results; F.B. coordinated the work; J. Stroh, A.S., M.H., D.H., and F.B. wrote the manuscript; and all authors discussed the results and commented on the manuscript.

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ORCID profiles: A.S., 0000-0002-2771-712X; R.E., 0000-0002-8433-9452; H.G., 0000-0003-0961-0035; U.K., 0000-0002-8485-1958.

Correspondence: Florian Bassermann (primary contact)
Department of Medicine III, Hematology and Oncology, Klinikum rechts der Isar, Technical University of Munich, Ismaninger Strasse 22, 81675 Munich, Germany; e-mail: florian.bassermann@tum.de; and Dirk Hose, VUB, Department of Hematology and Immunology, Myeloma Center Brussels, Laarbeeklaan 103, 1090 Jette, Belgium; e-mail: dirk.hose@vub.be.

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