Experimental MRI Monitoring of Renal Blood Volume Fraction Variations En Route to Renal Magnetic Resonance Oximetry

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INTRODUCTION

Kidney diseases are a global health burden with steadily increasing incidence (1-4), leading to an estimated worldwide death toll of 2 million per year from AKI (5-7). The currently available methods of assessing risk and therapeutic options for AKI are limited (5, 6, 8-10). Although a number of biochemical markers are being evaluated for use in diagnosis, risk assessment, and prognosis of AKI, there are currently no specific biomarkers that permit point-of-care diagnosis for early-stage AKI (12, 16-18). Strategies under consideration include novel imaging techniques being evaluated for use in diagnosis, risk assessment, and prognosis of AKI, and for the study of renoprotective strategies are urgently required (13-16). Strategies under consideration include novel imaging techniques that may be customized to probe early stages of AKI (12, 16-18).

Early features in the pathophysiology of AKI that could lend themselves to detection by noninvasive magnetic resonance (MR) imaging include renal tissue hypoperfusion and hypoxia—factors that are also important during the progression from AKI to chronic kidney diseases (16, 19-25). An imbalance between renal oxygen supply and demand appears to also play a prominent role in the pathophysiology of diabetic nephropathy (26). Renal oxygenation can be indirectly assessed through the blood oxygenation level-dependent (BOLD) magnetic resonance imaging (MRI) contrast (27), which can be observed through measurements of effective transversal relaxation time T2*.

Our findings provide motivation to advance multiparametric MRI for studying AKIs, with the ultimate goal of translating MRI-based renal BVf mapping into clinical practice en route noninvasive renal magnetic resonance oximetry as a method of assessing AKI and progression to chronic damage.

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Abbreviations: Acute kidney injury (AKI), blood volume fraction (BVf), free induction decay (FID), magnetic resonance imaging (MRI), oxygen saturation of hemoglobin (SO2), ultrasmall superparamagnetic iron oxide (USPIO), near-infrared spectroscopy (NIRS), signal-to-noise ratio (SNR), time-of-flight (TOF), venous occlusion (VO), radiofrequency (RF), repetition time (TR), echo time (TE), regions of interest (ROIs), multiecho gradient-echo (MGE), multi-spin echo (MSME)

Diagnosis of early-stage acute kidney injury (AKI) will benefit from a timely identification of local tissue hypoxia. Renal tissue hypoxia is an early feature in AKI pathophysiology, and renal oxygenation is increasingly being assessed through T2*-weighted magnetic resonance imaging (MRI). However, changes in renal blood volume fraction (BVf) confound renal T2*. The aim of this study was to assess the feasibility of intravascular contrast-enhanced MRI for monitoring renal BVf during physiological interventions that are concomitant with variations in BVf and to explore the possibility of correcting renal T2* for BVf variations. A dose-dependent study of the contrast agent ferumoxytol was performed in rats. BVf was monitored throughout short-term occlusion of the renal vein, which is known to markedly change renal blood partial pressure of O2 and BVf. BVf calculated from MRI measurements was used to estimate oxygen saturation of hemoglobin (SO2). BVf and SO2 were benchmarked against cortical data derived from near-infrared spectroscopy. As estimated from magnetic resonance parametric maps of T2 and T2*, BVf was shown to increase, whereas SO2 was shown to decline during venous occlusion (VO). This observation could be quantitatively reproduced in test-retest scenarios. Changes in BVf and SO2 were in good agreement with data obtained from near-infrared spectroscopy. Our findings provide motivation to advance multiparametric MRI for studying AKIs, with the ultimate goal of translating MRI-based renal BVf mapping into clinical practice en route noninvasive renal magnetic resonance oximetry as a method of assessing AKI and progression to chronic damage.
Renal BOLD MRI is based upon the $T_2^*$ dependence on $O_2$ saturation of hemoglobin (SO$_2$) and motivated by the link between $SO_2$, blood partial pressure of $O_2$ (pO$_2$), and tissue pO$_2$. However, questions have been raised regarding the interpretation of BOLD MRI data in the kidney as a surrogate of tissue oxygenation (30). These concerns were triggered by the following recent findings from renal $T_2^*$ mapping: simultaneous renal pO$_2$ and $T_2^*$ measurements showed considerable discrepancies in the quantitative relationship between changes in renal $T_2^*$ and those in renal tissue pO$_2$ for different functional regions of the kidney and for various (patho)physiological scenarios (31). The renal $T_2^*$ to tissue pO$_2$ relationship is not governed exclusively by renal blood oxygenation, but it is also heavily influenced by a number of confounders (30, 31). Of particular importance are changes in the renal blood volume fraction (BVf). Renal $T_2^*$ reflects the amount of deoxygenated hemoglobin per tissue volume, and any variation in BVf compromises the interpretation of $T_2^*$ changes as reflecting alterations in blood oxygenation, or even tissue oxygenation. This essential role of renal BVf has been largely unheeded, although it was recently highlighted by reports on changes in renal vascular conductance, local hemoglobin concentrations, and kidney size induced by physiological interventions (31, 32). The impact of renal BVf changes on $T_2^*$ exceeds that of other organs’ BVf owing to the considerable large BVf in the kidney (33). For these reasons, $T_2^*$ mapping alone cannot provide an unambiguous assessment of renal oxygenation.

A comprehensive renal MR oximetry protocol could have a quantitative value for the characterization of renal hemodynamics and tissue oxygenation if it integrates an assessment of renal BVf (30). Dynamic contrast-enhanced methods combined with tracer kinetic principles allow the quantification of renal perfusion and blood volume (34, 35) by using a bolus injection of exogenous agents like gadolinium chelates or iron oxide nanoparticles (36, 37). Analyzing the dynamic susceptibility contrast changes during bolus passage necessitates fast imaging with a temporal resolution of about 1 s, as well as measurement of the arterial concentration–time curve and its deconvolution from the tissue time curves. Renal dynamic susceptibility contrast was successfully implemented in dogs (36) and rats (37), but the methodological requirements limit the achievable spatial resolution and make this approach particularly challenging in small animals.

More recently, steady-state renal BVf measurements were performed by taking advantage of blood pool markers such as ultrasmall superparamagnetic iron oxide (USPIO) agents (38, 39); the change in transverse relaxation rate $R_2$ was examined in the kidney of mice (40) and rats (38). The steady-state approach comprises the simple subtraction of pre- and postcontrast maps of $R_2$ (41) (or $R_2$ if only small vessels are of interest), and it has the further benefit of facilitating the continuous monitoring of renal BVf (38). The USPIO nanoparticle preparation ferumoxytel has proven very useful for this (38, 42–45), as it can be administered intravenously without the risk of impaired renal oxygenation or perfusion (46), and it exhibits a long intravascular half-life of >14 h in humans and ~2 h in small rodents (43, 44, 47).

The goal of the current study was to determine whether intravascular contrast-enhanced MRI can be used as a means of monitoring BVf in physiological settings, in which significant variations in both renal BVf and renal oxygenation are expected, and, furthermore, to explore the possibility of correcting renal BOLD measurements for BVf variations. A previous study by Storey et al. (38) used steady-state BVf monitoring during the administration of vasoactive drugs, but the induced $R_2$ variations in the absence of USPIO were attributed to changes in tubular volume fraction rather than BVf or oxygenation. Here, we investigate the feasibility of renal BVf monitoring in rat kidneys at 9.4 T as a means of achieving a comprehensive renal MR oximetry protocol. For this purpose, ferumoxytol-enhanced renal $T_2^*$ and $T_2$ mappings were performed under baseline conditions and after a short-term reversible intervention of renal vein occlusion. This intervention is known to cause marked changes in renal blood pO$_2$ and renal BVf. Local BVf was calculated on the basis of changes in $T_2^*$, which is equally sensitive to vessels of all sizes, unlike previous works that used $T_2$ (38, 40), which is more sensitive to small vessels (48). Local $SO_2$ of the kidney was estimated using a model-based multiparametric MR technique (49), and variations in BVf and $SO_2$ were benchmarked against reference data obtained from near-infrared spectroscopy (NIRS). Given the lack of information in the literature on a suitable ferumoxytol dose for renal BVf measurements at 9.4 T, we performed dosage experiments to establish a useful level of sensitivity to changes in BVf and $SO_2$.

**METHODOLOGY**

**Animal Preparation and Control of Vital Functions**

All experiments were approved by the Animal Welfare Department of the Berlin State Office of Health and Social Affairs and were performed in accordance with the German Animal Protection Law. The procurement of animals, husbandry, and experiments conformed to the European Convention for the Protection of Vertebrate Animals used for Experimental and Other Scientific Purposes (Council of Europe No 123, Strasbourg 1985). Male Wistar rats (age, 12–13 weeks; body weight [BW], 288–330 g; n = 4; Harlan-Winkelmann, Borchern, Germany) underwent surgical preparation and MRI under urethane anesthesia (20% in H$_2$O; 6 mL/kg BW intraperitoneal; Sigma-Aldrich, Steinheim, Germany). In this approach, anesthesia is provided for several hours, leaving cardiovascular reflexes largely undisturbed.

To monitor arterial blood pressure, a catheter was placed into the femoral artery with its tip pointing towards the aorta. The catheter was connected to a pressure transducer (DT-XX, Viggo-Spectramed, Swindon, UK) and an amplifier (TAM-A Plugsys Transducer; Hugo Sachs Elektronik—Harvard Apparatus GmbH, March-Hugstetten, Germany). A second catheter was inserted into the right jugular vein, which permitted the administration of isotonic saline and USPIO. Core body temperature was monitored within the abdominal cavity by means of a fiber-optic temperature probe (OTP-M, AccuSens, Opsens, Québec City, Canada). Body temperature was maintained at 37°C with a pad supplied with steady warm water circulation.

**Renal VO as Test Intervention**

To induce renal VO during the MR study, a remotely operated inflatable cuff (occluder) was positioned around the left renal vein. Time-of-flight (TOF)-based MR angiography was performed immediately after cuff inflation and deflation to confirm the suc-
cessful occlusion and reperfusion of the kidney. In the rare case of a failed occlusion, we could detect the problem quickly and could perform a second attempt of the stimulus without a substantial delay.

The occlusion of the renal vein is known to induce both deoxygenation of the intrarenal blood and a substantial increase in intrarenal blood volume (32). Cessation of the venous outflow of blood increases blood pressure in intrarenal veins, thus resulting in their circular distension. The increased amount of deoxygenated blood in the renal tissue amplifies the BOLD effect, which induces a reduction in the $T_2^*$-weighted MR signal in response to VO. A further reduction in the MR signal due to $T_2^*$ shortening induced by the USPIO might lower the signal-to-noise ratio (SNR) to a critical level at which the MR image/data quality might become insufficient. Therefore, VO in combination with USPIO represents an extreme scenario with respect to the expected $T_2^*$ shortening.

**BVf Measurement**

BVf was measured using the USPIO ferumoxytol (Feraheme®, AMAG Pharmaceuticals, Inc., Lexington, MA). Ferumoxytol is approved in the USA and the EU as an intravascular Fe supplement therapy for patients with iron deficiency anemia related to chronic kidney disease. The intravenous injection of ferumoxytol does not have any measurable effects on renal physiology at doses up to 41 mg Fe/kg BW in rats; the presence of ferumoxytol also does not significantly alter the control of renal hemodynamics and oxygenation as studied by aortic occlusion and hypoxia (46).

Renal BVf was calculated by comparing pre-ferumoxytol data ($R_2^*$-maps) with post-ferumoxytol data ($R_2^*$USPIO-maps) (41):

$$BVf = \frac{3}{4\pi} \cdot \frac{(R_{2,USPIO}^* - R_{2}^*)}{\Delta \chi_{USPIO} \cdot B_0 \cdot \gamma} = \frac{3}{4\pi} \cdot \frac{\Delta R_{2,USPIO}^*}{\Delta \chi_{USPIO} \cdot B_0 \cdot \gamma}$$

(1)

where $\gamma$ is the gyromagnetic ratio, which is $2.675 \times 10^8$ rad/(s T); $B_0 = 9.4$ T; and $\Delta \chi_{USPIO}$ is the susceptibility difference between blood with and without added USPIO: $\Delta \chi_{USPIO} = 0.024$ ppm (cgs units) × [USPIO dose in milligram Fe per kilogram BW] (50).

**Ferumoxytol Dose-Finding Study**

Considering the lack of information on a suitable ferumoxytol dose for renal BVf measurements at 9.4 T, we performed a dose-finding study. To accomplish this, the following 3 competing effects had to be balanced:

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**Figure 1.** Illustration of experimental protocol. Magnetic resonance (MR) monitoring of a baseline–stimulus–recovery cycle was performed 5 times: once without ultrasmall superparamagnetic iron oxide (USPIO) (phase 1) and 4 successive times (phases 2–5) each with an incremental dose of USPIO.
(1) sensitivity of the BVf measurements increases with the USPIO dose;
(2) the SNR of MR images decreases with the USPIO dose; and
(3) sensitivity to changes in BVf also decreases at high USPIO doses because $T_2^*$ tends to zero.

For these reasons, a ferumoxytol dose ranging from 2 to 8 mg Fe/kg BW was used.

**Experimental Protocol**

The experimental protocol comprised 5 phases with increasing cumulative doses of ferumoxytol as illustrated in Figure 1:

(1) *Phase 1*: Baseline–stimulus–recovery without USPIO

(a) Baseline $T_2^*$ and $T_2$ mapping and TOF MR angiographic confirmation of renal blood flow.

(b) VO performed by remote inflation of the cuff.

(c) The absence of renal blood flow was checked immediately using TOF MR angiography.

(d) $T_2^*$ and $T_2$ mapping during VO.

(e) Release of the venous cuff. Effective duration of VO was $\sim$3 min (depended on acquisition time of respiration-gated MRI scans performed in steps c and d).

(f) Restoration of renal blood flow was checked by TOF angiography.

(g) Recovery phase, during which $T_2^*$ and $T_2$ mapping was continued: 5 repetitions, approximately every 3 mins covering a 12- to 14-min period (dependent of respiratory rate because scans were respiration-triggered).

(2) *Phase 2*: Baseline–stimulus–recovery with first USPIO dose

(h) The intravascular contrast agent (USPIO; 2 mg of Fe/kg body mass) was administered using a power injector at a rate of 0.25 mL/min via the jugular vein catheter.

(i–o) Following the conclusion of the injection and an additional 3-min mixing time, the steps of phase 1 were repeated to obtain matching post-USPIO data for phase 1.

(3) *Phase 3*: Baseline–stimulus–recovery with second USPIO dose

Same as phase 2 but with a different USPIO dose: the additional injection of 2 mg of Fe/kg USPIO yielded the approximate cumulative dose of 4 mg of Fe/kg USPIO (owing to the long half-life of the USPIO).

(4) *Phase 4*: Baseline–stimulus–recovery with third USPIO dose

Same as phase 2 but with a different USPIO dose; the additional injection of 2 mg of Fe/kg USPIO yielded an approximate cumulative dose of 6 mg of Fe/kg USPIO.

(5) *Phase 5*: Baseline–stimulus–recovery with fourth USPIO dose

Same as phase 2 but with a different USPIO dose; the additional injection of 2 mg of Fe/kg USPIO yielded the approximate cumulative dose of 8 mg of Fe/kg USPIO.

The duration of the experimental phase was $\sim$20 min, which is short in comparison to the duration of the plasma half-life of ferumoxytol in rats (up to 3 h) (38).

**MRI Imaging Experiments**

MRI experiments were performed on a 9.4 T animal MR system (Biospec 94/20, Bruker Biospin, Ettlingen, Germany) using a radiofrequency (RF) coil setup established for renal imaging (linear polarized birdcage RF resonator for transmission in conjunction with a curved 4-channel receive RF coil array; Bruker Biospin, Ettlingen, Germany).

$T_2^*$-weighted pilot scans for geometrical planning and section positioning were acquired. We conducted a local volume-selective shimming of the magnetic field homogeneity on a voxel enclosing only the kidney, using an automatic optimization algorithm based on the FID length.

Interleaved $T_2^*$ and $T_2$ mapping was performed with respiratory-gated (Model 1025, SA Instruments, Stony Brook, NY) imaging protocols. For $T_2$ mapping, a multiecho gradient-echo
mapping, a multi-spin echo correction in renal T2 inhomogeneities, and then first-order local shimming and R2 map (in-house by pixelwise monoexponential fitting to and SNR measured in vivo at different Fe doses. In vivo, the B0 map of the kidney. This B0 map of the kidney for different shim settings and |

versus the true T2 and T1 uniform across the kidney. The latter was performed 3 times map; (iii) calculating intravoxel dephasing; and (iv) using this correction involves was performed to correct the first TE 10 milliseconds at the beginning of the expiratory plateau, which resulted in the acquisition of several k-space lines within this window. For MGE measurements, the respiratory trigger window included the entire expiratory plateau, which allowed for the acquisition of 1 k-space line per breath, with the effective TR being equal to the respiratory interval.

A coronal oblique section was placed such that it covered the kidney centrally at its maximum extension. An in-plane spatial resolution of (226 x 445) μm², field of view = (38.2 x 50.3) mm², matrix size = 169 x 113 zero-filled to 169 x 215, and a section thickness of 1.4–1.5 mm was used.

For TOF angiography, we used a spoiled gradient echo technique (2D FLASH; TR = 11 milliseconds; TE = 3 milliseconds; flip angle = 80°) with a spatial in-plane resolution of (200 x 268) μm² and 15 sections (section thickness = 1.0 mm).

**Region of Interest Analysis**

We extended the semiautomated kidney segmentation approach used in our previous studies (31, 51) to provide 5 regions of interest (ROIs) in the renal cortex and 5 ROIs in the renal outer medulla (Figure 2). In brief, a rectangle was manually placed around the kidney borders, followed by drawing lines at the crossing of the kidney border and 2 automatically placed diagonals. Further, 10 ROIs were placed automatically at locations in the cortex and outer medulla that had been predefined with respect to the reference rectangle.

**Effects of Magnetic Field Inhomogeneity on Renal T2**

Three-dimensional mapping of B0 was performed to correct the B0 inhomogeneity effects on T2* (49). This B0 correction involves (i) acquiring a 3D B0 map of the kidney; (ii) estimating intra-voxel field dispersion across the kidney by fitting a 3D polynomial to the B0 map; (iii) calculating intravoxel dephasing; and (iv) using this for correcting the measured T2* map of the kidney.

To assess the need for B0 correction in renal T2* mapping, we performed B0 correction on a kidney for different shim settings and compared the corrected renal T2*-maps with the measured, uncorrected T2*-maps. All shims were first set to zero to allow for significant B0 inhomogeneities, and then first-order local shimming was performed on a voxel tightly enclosing the kidney to make B0 uniform across the kidney. The latter was performed 3 times to account for any variability. A 3D polynomial function of the third order was fitted to the measured 3D B0 map (in-house developed program; MATLAB, MathWorks, Natick, WA). This polynomial function was used to estimate the spin dephasing on a subvoxel grid of 2 x 2 x 4 voxels per image voxel (voxel size = 0.22 x 0.23 x 1.40 mm³). Subvoxel grids finer than 2 x 2 x 4 did not yield significantly different results.

**Model-Based MRI Data Analysis**

Multiecho MR data were converted into parametric maps of absolute T2* and T2 by pixelwise monoexponential fitting to the signal intensities of denoised (SANLM filter, VBM8 toolbox, SPM8; www.fil.ion.ucl.ac.uk/spm) series of T2*- and T2-weighted images acquired as a function of the TE (in-house developed program; MATLAB, MathWorks, Natick, WA). The relaxation rates were calculated as R2* = 1/T2* and R2 = 1/T2. The subtraction of precontrast maps from corresponding postcontrast maps yielded ΔR2* -maps, which were used for the calculation of BVf-maps using equation (1).
Renal blood SO₂ was estimated by applying a multiparametric MR technique (49) to the R₂*, R₂ and BVf data from the kidney. This approach was originally proposed for human brain MRI (52) and is based on a theoretical model of the BOLD contrast (53). This approach was refined by replacing a complex model fit with a simpler single parameter fit combined with actual MR measurements of R₂, BVf, and B₀ (49). Assuming that the magnetic field inhomogeneity is negligible and constant throughout the duration of the experiment, we solved the model equation
\[ s(t) = f(t, BVf, SO₂, \ldots) \]
for SO₂:
\[ SO₂ \approx 1 - \frac{3}{4\pi} \cdot \frac{(R₂^* - R₂)}{γ \cdot Δχ₀ \cdot BVf \cdot Hct \cdot B₀} \] (2)

Here \( Δχ₀ = 0.264 \) ppm (54), which represents the susceptibility difference between deoxygenated and oxygenated red blood cells, B₀ = 9.4 T, and SO₂ is given in arbitrary units (because B₀ is uncorrected). A hematocrit of 0.40 was used for the cortex and the outer medulla (85%–95% of systemic hematocrit, which was assumed to be 0.45) (55). In this feasibility study, the data analysis was performed using equations (1) and (2), as these permit access to both parameters of interest, that is, SO₂ and BVf.

To evaluate intrasubject reproducibility, we compared results from the baseline with those obtained after 10 min of recovery and also repeated the baseline–occlusion–recovery experiment (~25 min later). Moreover, before repeating the experiment, we increased the USPIO dose by 2 mg Fe/kg BW to assess the dependency of the results on the USPIO dose.

**NIRS**

Quantitative information on the relative changes of total cortical hemoglobin per tissue volume (as a surrogate for BVf) and cortical SO₂ during VO were obtained using multidistance continuous wave NIRS in a separate cohort of 10 male Wistar rats. The NIRS measurements are based on recording spatially resolved diffuse reflectance with a linear fiber probe. This probe permitted sampling of the renal tissue up to a maximum depth of ~2 mm and provided information about the renal cortex. The results presented here were obtained by a refined model analysis of data reported in Grosenick et al. (32), with improved separation of tissue absorption changes from changes in tissue scattering. To reference our results against these data, the high-temporal-resolution NIRS data were averaged over the duration of each MGE MRI scan.

**Figure 4.** T₂*-weighted images (echo time [TE] = 3.6 milliseconds, spatial resolution = 226 × 422 μm) of a rat kidney at baseline, during venous occlusion (VO), and at the beginning of the recovery phase. Without USPIO (top row), the short-term VO induces a very strong signal decrease in the renal cortex and the outer medulla. The impact of USPIO administration on T₂*-weighted image contrast (left column) lends itself to an estimation of blood volume fraction (BVf). A hypointense area stretching from the papilla via the inner medulla to the central outer medulla becomes prominent with increasing USPIO dose. This hypointensity might represent the influence of large Fe-rich vessels located close to the image section rather than medullary tissue properties (see Discussion).
RESULTS

Ferumoxytol Dose-Finding Study

The dose-finding study was performed to ensure that BOLD effects were detectable after USPIO administration taking into account that SNR may be further reduced by changes in $T_2^*$ owing to the deoxygenation of blood. By performing Monte Carlo simulations for a wide range of $T_2^*$ and SNR, we estimated the SNR-dependence of the $T_2^*$ mapping error (Figure 3).

\[
\Delta T_{2,\text{error}} = T_{2,\text{calc}} - T_{2,\text{true}}
\]

with “true” indicating the true $T_2^*$ value used as an input and “calc” indicating the $T_2^*$ value calculated by curve fitting to the noisy signal intensity data versus TE. For cortical and outer medullary $T_2^*$ the absolute error in $T_2^*$ was $<1.0$ milliseconds.

$T_2^*$-Weighted MR Imaging

We observed a significant change in contrast and image quality following short-term VO and after increasing USPIO concentrations (Figure 4). VO caused a very strong signal decrease in the renal cortex and in the outer medulla. Even after a USPIO administration of 4 or 6 mg of Fe/kg BW, the strength of this decrease of the signal during VO remained similar and was easily detectable. Although the signal loss suggested renal hypoxia, that is, a decrease in oxygen saturation of hemoglobin, this remained speculative because, at this point, possible changes in BVf had not been considered. Comparing the $T_2^*$-weighted images at baselines for different USPIO doses shows the impact of USPIO administration on image contrast, which lends itself to an estimation of BVf.

Important for estimating the BVf is the sensitivity for detecting the change in $T_2^*$ caused by labeling the blood with USPIO. As expected, a larger USPIO dose provided more sensitivity for the measurement of:

\[
\Delta T_{2,\text{USPIO}} = T_{2,\text{USPIO}} - T_2
\]

because the larger the cumulative USPIO dose, the larger the change in $T_2^*$ compared with the baseline (Figure 5A). This $T_2^*$ change was more pronounced in the outer medulla than in the renal cortex. In contrast to the sensitivity to USPIO effects, the sensitivity to changes in $T_2^*$ induced by VO decreased rapidly with the Fe dose (Figure 5B):

\[
\Delta T_{2,\text{VO}} = T_{2,\text{VO}} - T_{2,\text{baseline}}
\]

The sensitivity to changes in $T_2^*$ induced by VO fell below $\Delta T_2^* = 2.0$ milliseconds in the outer medulla for USPIO doses of 6 mg of Fe/kg and more. The relative $\Delta T_2^*$ as percentage of baseline $T_2^*$, did not change much and stayed between $\sim 60\%$ and $80\%$.

Considering the sensitivity of $T_2^*$ to USPIO injection (relevant for BVf estimation) and the $T_2^*$ sensitivity to VO (physiological stimulus) at different cumulative USPIO doses. Plots of changes in cortical and outer medullary $T_2^*$ induced by the injection of USPIO (A). Absolute $\Delta T_2^*$ in milliseconds (left panel) and relative $\Delta T_2^*$ as percentage of $T_2^*$ without USPIO (right panel) increased rapidly with USPIO dose. Plots of changes in cortical and outer medullary $T_2^*$ induced by VO ($T_2^*$ values during a given VO versus $T_2^*$ before the respective VO) (A). Although absolute $\Delta T_2^*$ in milliseconds (left panel) decreased with USPIO dose, the percentage $\Delta T_2^*$ (right panel) remained between $\sim 60\%$ and $80\%$. Group means and SEMs (n = 4) of cortical and outer medullary $\Delta T_2^*$ obtained from the ROIs are determined as shown in Figure 2.

Effects of Magnetic Field Inhomogeneity on Renal $T_2^*$

Macroscopic gradients in the magnetic field $B_0$—for instance, owing to imperfect $B_0$ shimming—may cause undesirable spin dephasing and could artificially shorten $T_2^*$. Before mapping renal $R_2^*$, BVf, and $S_O_2$, we tested whether the susceptibility weighting and $T_2^*$ were dominated by microscopic $B_0$ susceptibility gradients. We investigated whether $B_0$ correction could reverse unwanted magnetic field inhomogeneity effects on $T_2^*$ by comparing measured and corrected renal $T_2^*$-maps acquired in vivo for different magnetic field shim settings as shown in Figure 6.

The 3D polynomial fits to the measured $B_0$-maps described the magnetic field inhomogeneity with high fidelity. A notable
Macroscopic magnetic field inhomogeneity was present with the shim settings adjusted to zero. For this shim setting, \( B_0 \) correction markedly increased \( T_2^* \), particularly in the inner medulla, where \( T_2^* \) is typically large. \( B_0 \) inhomogeneity across the kidney was very small after local shimming on the kidney. After \( B_0 \) correction, the renal \( T_2^* \)-maps obtained for all 3 \( B_0 \) shim settings displayed high agreement. For local shimming, which we perform routinely in renal MR studies, the effect of \( B_0 \) correction on \( T_2^* \) was negligible. This confirms that macroscopic intravoxel dephasing causes only minor \( T_2^* \) effects for the TE range and the voxel size used and ensures that \( T_2^* \) is governed by microscopic \( B_0 \) susceptibility gradients.

Renal \( R_2 \), \( R_2^* \), BVf, and \( SO_2 \) Mapping
Parametric maps were calculated for renal \( T_2 \) and \( T_2^* \), which were then converted to \( R_2 \)- and \( R_2^* \)-maps, respectively. Such quantitative maps permit comparisons between animals and over time, as they are not biased by external factors such as RF coil sensitivity \( (B_{1}) \) or the position of the subject under investigation with respect to the receive RF coil. The difference between \( R_2^* \)- and \( R_2 \)-maps \( (\Delta R_2) \) and \( \Delta R_2 \), respectively, acquired before and after USPIO administration is closely related to the local BVF. The blood volume measurement procedure is illustrated in Figure 7, which shows parametric maps of renal \( R_2 \) and \( R_2^* \) relaxation rates under baseline conditions, without and with USPIO (4 mg of Fe/kg), along with their difference. Dissimilarities between \( \Delta R_2 \) and \( \Delta R_2^* \) are expected owing to their different sensitivities to large vessels (see Discussion).

Analysis of BVF-maps obtained at baseline and during renal VO (Figure 8) revealed an increase in cortical and medullary BVF upon VO. Renal BVF returned to baseline after 10 min of recovery. Next, maps of renal \( SO_2 \) were calculated using the multiparametric BOLD model outlined in equation (2), which requires \( T_2^* \) and BVF as input data to analyze \( SO_2 \) at baseline, upon VO and on recovery (Figure 8). The reduction in renal \( T_2^* \) during VO was associated with a decrease in \( SO_2 \) in the cortex and outer medulla combined with a substantial increase in blood volume.

To show the necessity of monitoring BVF (rather than using a fixed literature value), we also calculated \( SO_2 \)-maps that assume renal BVF to remain constant and identical to the baseline condition during the entire experiment (Figure 8, lower panel). Almost everywhere in the cortex and outer medulla, \( SO_2 \) values were considerably lower in the experiments neglecting BVF changes compared with experiments taking BVF changes into account.

Reproducibility of results was very high: the results obtained for the cortical and outer medullary BVF and \( SO_2 \) at baseline conditions and after 10 min of recovery were almost
was much higher than cortex and outer decrease during renal VO. The results of this assess would be much less susceptible to the measured by NIRS were compared obtained from MRI were benchmarked against quantitative derived from MRI were decreased markedly during the short-term VO for both VOLUME 3 NUMBER 4 tissue concentration, as measured by NIRS. As for the injected USPIO, hemoglobin effectively serves as a naturally occurring label in the blood, and hence, its tissue concentration may be regarded as a surrogate for BVf, as long as the hematocrit remains unchanged.

**DISCUSSION**

This work makes an important contribution to the literature on renal functional MRI by assessing changes in the renal BVf and in the renal SO₂ in response to VO. To achieve this goal, BVf measurements were implemented for rat kidneys at 9.4 T, including a dose-finding study for the intravascular contrast agent ferumoxytol. Multiparametric analysis was performed to estimate renal SO₂. Occlusion-induced changes in BVf and SO₂ derived from MRI were benchmarked against BVf and SO₂ references obtained from NIRS. Our main findings are that (1) a 4 mg of Fe/kg dose of ferumoxytol is suitable for BVf measurements at 9.4 T using baseline, VO and recovery; (2) the proposed approach provides high reproducibility for BVf and SO₂ assessment as demonstrated by the test–retest experiments; (3) relative changes in cortical BVf and cortical SO₂ derived from MRI were in accordance with relative changes in BVf and SO₂ deduced from NIRS; and (4) without the monitoring of BVf, MRI overestimates the SO₂ decrease during renal VO. The results of this work permitted a noninvasive detection of BVf increase upon VO and a removal of its effects on blood oxygenation-sensitized renal MR.

We found BVf to be higher in both outer and inner medulla than in the cortex, as shown by both ΔR₂- and ΔR₂*-maps. Although these observations do not agree with recent results obtained by 3D microcomputed tomography (where cortical BVf was reported to be larger than medullary BVf) (56), they are in alignment with previous reports on ΔR₂*-based BVf estimates in rats and mice (38, 40), as well as with a series of earlier reports that measured renal BVf by means of (51) Cr-labelled red cells and ¹²⁵I-γM-immunoglobulin: Rasmussen (55) reported medullary BVf in rats to be approximately twice the cortical BVf. There is no gold standard method, and the results provided by different techniques differ significantly. A hypointensity in T₂*-weighted images had already suggested that BVf was higher in the medulla, but the ΔR₂*-maps depicted this even more clearly. The area with apparently very high BVf stretches from the papilla via the inner medulla to the central outer medulla. We hypothesize that these phenomena represent the influence of large Fe-rich vessels (57) located close to the image section, rather than the actual medullary tissue properties. The dissimilarity between ΔR₂- and ΔR₂*-maps in revealing this area of unexpectedly high BVf further support our hypothesis: unlike ΔR₂, ΔR₂ is predominantly sensitive to small vessels and capillaries, because it relies on water diffusion within the near environment of the vessel walls, and the surface-to-volume ratio is highest for small vessel diameters. Hence, ΔR₂ would be much less susceptible to the long-distance effects of a high amount of USPIO within extremely large blood vessels. Indeed, the locations of apparently high BVf in the papilla and inner medulla colocalize with the renal artery and vein, and the interlobar arteries and veins (56).

In the outer medulla, BVf displayed a rather high spatial heterogeneity and large spatial gradients along the longitudinal
(rostral–caudal) axis. Here, large blood vessels (interlobar arteries and veins) surrounding the image section, as well as arcuate arteries and veins that run along the border between cortex and medulla (56), may play a role. Owing to the relatively large section thickness, the $T_2^*$-weighted images are considerably susceptible to magnetic field and frequency dispersions perpendicular to the image plane, created by large USPIO-loaded vessels. This hinders the assessment of the renal medulla. The choice of an axial section orientation or much thinner section thickness (if permitted by the SNR) could help resolve this issue. Despite these limitations, the variations in cortical BVf during renal VO were in good agreement with the results obtained from NIRS.

Our approach of parametric mapping of renal $T_2^*$ and $T_2$ made use of MGE and MSME techniques including respiratory triggering for respiratory motion compensation. The duration of each scan was 60–90 s, permitting a TR of ~3 min for interleaved $T_2^*$ and $T_2$ mapping. These protocols are available on clinical MR scanners, and hence it will be easily translatable to clinical use. The translation of our approach into the clinic is fueled by an increasing number of reports that eloquently speak of the off-label use of ferumoxytol for a broad spectrum of preclinical and diagnostic imaging applications (32, 43, 44, 58, 59). In the clinical setting, renal $T_2^*$ and $T_2$ mapping will most likely be used at time points that are at least hours, if not days or months, apart. In such a context, a 90-s delay between $T_2^*$ and $T_2$ scans appears to be short enough. Protocols affording breath-hold acquisitions could make respiratory triggering unnecessary, and parallel imaging capabilities are readily available on human MR scanners, both permitting acceleration of data acquisition.

In a laboratory setting, a higher temporal resolution might be needed to study some acute stimuli applied to animals. Fast physiological changes demand shorter scan durations and higher repetition rates. Imaging techniques that provide $T_2^*$- and $T_2$-weighted data (52, 60) provide an alternative to these preclinical applications. Fast spin-echo variants for $T_2$ and $T_2^*$ mapping permit choosing any desired $T_2^*$-weighting (including ultrashort times down to zero) and provide the extra benefit of being almost immune to image distortion (61, 62). Simultaneous dual-contrast 2-in-1 rapid acquisition with relaxation enhancement

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**Figure 8.** Maps of renal cortical and outer medullary $T_2^*$, together with estimated maps of BVf and oxygen saturation of hemoglobin ($SO_2$) at baseline, during VO and the recovery phase. The maps at baseline and after 10 min of recovery are almost indistinguishable, confirming that the effects of VO are reversible. Within-subject repeatability is demonstrated for renal BVf and $SO_2$ by comparing the maps derived from 2 different experimental phases, namely, phase 3 (4 mg of Fe/kg USPIO) and phase 4 (6 mg of Fe/kg USPIO). Test-retest reliability (repeatability) was high—the differences between both iterations are nearly negligible, even though more USPIO had been injected in between them. In addition, $SO_2$ maps were calculated assuming that renal BVf remains constant and identical to the baseline condition (bottom row); the missing compensation for BVf changes during VO results in $SO_2$ values being considerably lower than when BVf was monitored.
presents a valuable alternative to sequential $T_2^*$ and $T_2$-weighted fast-spin echo acquisitions and promises to eliminate section-misregistration artifacts induced by bulk or physiological motions (63). Further enhancements of the SNR and spatial resolution could be gained by using cryogenically cooled RF coils in a preclinical setting (64, 65).

The multiparametric BOLD approach based on combining the monitoring of $T_2^*$ and $T_2$ with BVF has not previously been attempted for renal MR blood oxymetry. This study demonstrates the feasibility of this approach in rats. A high reproducibility of $T_2^*$ and $T_2$ mapping and BVF measurements translated into SO$_2$ estimates with similarly high reproducibility.

The impact of the assumptions embedded in the model has previously been studied and it indicates that the model is sufficiently realistic (66) for the brain. The assumptions about the microvascular architecture should also hold for the kidney, except that the BVF used in the numerical study (4%) should be higher in renal tissue (literature values are contradictory but typically exceed 10%). However, because the model does not take into account the effects of renal tubuli, the presented study results may serve as only an approximate indicator for the usefulness of such a model-based approach and further investigation will be needed to advance the model for the kidney.

The current implementation of the model-based analysis has some methodological constraints that can be improved in future work. For the current study, intrarenal hematocrit was assumed to be 0.40, but, in fact, this value is known to vary throughout the kidney. Strategies to account for this may be necessary. To account for the susceptibility difference between blood with and without added ferumoxytol, a literature value of $\Delta X$ for a different USPIO of the same size and at the same magnetic field strength was used (50). It would be an added refinement to measure the $\Delta X$ for ferumoxytol and include it in the BVF calculations in the future. Because of our focus on fast, short-term changes in renal oxygenation, we assumed a constant magnetic field homogeneity, which might change modestly with respect to the kidney owing to respiratory motions and changes in renal size (renal tissue moves within B$_0$). SO$_2$ results were reported in arbitrary units because B$_0$ effects were not accounted for in this study. Our initial experiments (Figure 6) had shown that the effects of magnetic field inhomogeneity on renal $T_2^*$ were negligible after local shimming on the kidney, which we performed routinely. Generally, acquiring B$_0$-maps for each subject are recommended to permit a B$_0$ correction, if needed, and exclude a possible bias. Alternatively, acquiring high-resolution $T_2^*$ data makes a separate B$_0$ correction unnecessary owing to the reduced intravoxel dephasing (49, 67, 68), and it could represent an attractive way forward, presuming that the scan time penalty for increasing the spatial resolution is either of no relevance for the application or can be counteracted by strategies to accelerate acquisition. This can be accomplished, for example, by using combined acquisition techniques that integrate a minimum of 2 imaging strategies for $T_2^*$ and $T_2$ mapping (69).

Benchmarking BVF and SO$_2$ results against a quantitative reference is an obvious means of validating experimental results obtained from a novel MR technique. Currently, a perfect quantitative counterpart for MR-derived BVF and SO$_2$ is not available, as is the case for most MR techniques. Invasive tissue pO$_2$ probes can sample only very small regions, and they measure tissue pO$_2$ rather than blood oxygenation. NIRS allows measurements of the oxygen saturation of hemoglobin and tissue concentration of hemoglobin—a surrogate for BVF—but is currently limited to probing the cortex due to its low penetration depth. Comparisons between the occlusion-induced changes in cortical BVF and SO$_2$ obtained by MRI with the NIRS results for the cortex yield very good agreement, which should motivate the
further development of the MR approach proposed here into a comprehensive renal MR oximetry protocol.

In conclusion, this work established ferumoxytol-based steady-state MR measurements of renal BVfs for rats at a magnetic field strength of 9.4 T. Combining the BVf measurements with the monitoring of $T_2^*$ and $T_2$ allowed us to implement multiparametric quantitative BOLD MRI for the kidney as a promising approach en route renal blood oximetry. The findings are encouraging and should stimulate efforts to further improve this multiparametric technique, with the ultimate aim of translating it into clinical practice for the evaluation of AKIs and the development of chronic damage. Accomplishing this goal will require calibrations through simultaneous quantitative measurements with invasive physiological probes and NIRS in the same kidney. Once available to the clinicians, multiparametric renal MR oximetry will represent the first noninvasive method to reliably measure renal blood oxygenation. It could be combined with MR techniques for perfusion and diffusion (to probe tubular volume fraction) en route a comprehensive characterization of renal hemodynamics and tissue oxygenation, which may be an important biomarker for the early stages of a range of kidney diseases.

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