

**Supplementary material for**

**Alterations to mTOR signaling impacts metabolic stress resistance in BRAF and KRAS mutated  
colorectal carcinomas**

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## Supplementary Table

**Supplementary Table 1:** pSIRM approach was performed using CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup> and CaCO<sub>2</sub>-KRAS<sup>G12V</sup> cells (in minimum n=3). Mean of quantities (pmol/L 000 000 cells), <sup>13</sup>C-glucose incorporation (LI [%]) and normalized labeled quantities (NLQ [pmol/L 000 000 cells]) of metabolites (product of quantities and incorporation). Cells were cultivated and labeled for 5 min with physiological amounts of glucose (1.0 g/L). Ala: Alanine, Cit: Citric acid, Fum: Fumaric acid, Lac: Lactic acid, Mal: Malic acid, Pyr: Pyruvic acid, Ser: Serine, Suc: Succinic acid.

	Cell line	CaCO <sub>2</sub> -control	CaCO <sub>2</sub> -BRAF <sup>V600E</sup>	CaCO <sub>2</sub> -KRAS <sup>G12V</sup>
<b>quantities</b>	<b>Ser</b>	18935	15076	71435
	<b>Pyr</b>	2805	969	2568
	<b>Lac</b>	41227	45932	32545
	<b>Ala</b>	4097	865	688
	<b>Cit</b>	2493	3110	4167
	<b>Suc</b>	1641	1766	3635
	<b>Fum</b>	763	901	1565
	<b>Mal</b>	1843	1189	4354
	<b>LI</b>	<b>Ser</b>	0.3	0.7
<b>Pyr</b>		20.9	24.3	26.4
<b>Lac</b>		16.7	18.5	10.7
<b>Ala</b>		3.3	6.3	5.4
<b>Cit</b>		10.8	18.5	11.0
<b>Suc</b>		9.1	9.2	9.3
<b>Fum</b>		1.7	1.9	0.5
<b>Mal</b>		13.6	14.2	13.2
<b>NLQ</b>		<b>Ser</b>	46	120
	<b>Pyr</b>	527	241	864
	<b>Lac</b>	6716	8538	3745
	<b>Ala</b>	98	60	35
	<b>Cit</b>	343	650	448
	<b>Suc</b>	160	190	377
	<b>Fum</b>	14	19	8
	<b>Mal</b>	262	180	629

## Supplementary Figures

**Supplement Figure 1:** **A)** Cells were cultivated in the presence of Doxycycline and indicated glucose concentrations for 16 d. **B)** CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup> and CaCO<sub>2</sub>-KRAS<sup>G12V</sup> cells were cultivated in medium containing 1.0 g/L glucose and measured with LC-MS (shot gun proteomics). Shown are log<sub>2</sub> fold changes (fc) to CaCO<sub>2</sub>-control cells for proteins associated with epithelial to mesenchymal transition, migration and actin remodeling. Significant regulations (p<0.05 unpaired two-tailed *t* Test) comparing CaCO<sub>2</sub>-control and CaCO<sub>2</sub>-BRAF<sup>V600E</sup> or CaCO<sub>2</sub>-control and CaCO<sub>2</sub>-KRAS<sup>G12V</sup> cells were indicated with asterisks or crosses, respectively. **C)** CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup> and CaCO<sub>2</sub>-KRAS<sup>G12V</sup> cells cultivated in medium containing physiological (1.0 g/L) glucose were stained with periodic acid-Schiff (PAS) and alcian blue (AB) to verify neutral and acidic Mucin expression in vacuoles, respectively. **D)** MUC5AC expression was analyzed using ELISA assay. Cells were cultivated in physiological (1.0 g/L) glucose. Data were quantified and shown as log<sub>2</sub> fold changes (fc). **B, C, E)** Shown are standard deviation of n=3 replicates. p<0.05 was indicated with asterisk (unpaired two-tailed *t* Test).

**Supplement Figure 2:** **A, B)** Flow cytometry profiles of CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup>, CaCO<sub>2</sub>-KRAS<sup>G12V</sup>, HT29 and SW480 cells cultivated with indicated glucose amounts stained for cleaved Caspase 3. Shown are percent (%) of apoptotic cells. Shown are standard of n=2 replicates. p<0.05 was indicated with asterisk (unpaired two-tailed *t* Test)). **C, D)** CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup>, CaCO<sub>2</sub>-KRAS<sup>G12V</sup>, HT29 and SW480 cells grown in physiological (1.0 g/L) glucose concentrations were treated with 10 μM Rapamycin (+), 10 μM OSI027 (+) or DMSO (-) for 24 h and analyzed with antibody against phosphorylated 4eBP1 (Thr70). Vinculin served as loading control. Samples for each cell line were loaded on separate gels. **E, H)** Phosphorylation of S6-kinase after treatment with 10 μM Rapamycin (+), 10 μM OSI027 (+) or DMSO (-) and AKT after treatment with 1 μM (+) MK2206 or DMSO (-) for 24 h was analyzed with ELISA bead-based phosphoproteomics technology (BioPlex). Shown are log<sub>2</sub> fold changes (fc) to DMSO control (per cell line). **F, G)** CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup>, CaCO<sub>2</sub>-KRAS<sup>G12V</sup>, HT29 and SW480 cells grown in physiological glucose concentrations

were treated with 1  $\mu$ M MK2206 (+) or DMSO (-) for 24 h. Shown are viable cells compared to DMSO. Shown are standard deviation of n=3 replicates.  $p < 0.05$  was indicated with asterisk (unpaired two-tailed *t* Test). **I, J**) CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup>, CaCO<sub>2</sub>-KRAS<sup>G12V</sup>, HT29 and SW480 cells grown in physiological (1.0 g/L) glucose concentrations were treated with 1  $\mu$ M MK2206 (+) or DMSO (-) for 24 h and analyzed with antibody against phosphorylated 4eBP1 (Thr70). Vinculin served as loading control. Samples for each cell line were loaded on separate gels. **G**) BRAF was immunoprecipitated and the immunocomplexes were blotted using antibodies against BRAF and RAPTOR (different plots). IP and lysates were loaded on different gels. Vinculin served as loading control.

**Supplement Figure 3:** **A**) Hierarchical clustering of relative protein quantities (z-score) were shown for CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup> and CaCO<sub>2</sub>-KRAS<sup>G12V</sup> cells cultivated in physiological (1.0 g/L) glucose. Proteins associated to central carbon metabolism were indicated per cluster. Enrichment analysis was done using gene ontology biological process terms. **B-D**) CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup> and CaCO<sub>2</sub>-KRAS<sup>G12V</sup> cells were exposed to 1.0 g/L (B, C) or 2.5 g/L (D) of <sup>13</sup>C-glucose for 5 min, harvested and measured with GC-MS. The ratio (log<sub>2</sub> fold changes, fc) of B) CaCO<sub>2</sub>-BRAF<sup>V600E</sup> to CaCO<sub>2</sub>-control, C) CaCO<sub>2</sub>-KRAS<sup>G12V</sup> to CaCO<sub>2</sub>-control or D) CaCO<sub>2</sub>-KRAS<sup>G12V</sup> to CaCO<sub>2</sub>-BRAF<sup>V600E</sup> for labeled (metabolites) quantities are shown. **E**) CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup> and CaCO<sub>2</sub>-KRAS<sup>G12V</sup> cells were exposed to <sup>13</sup>C-glucose (physiological and intermediate amounts) for 5 min, harvested and measured with GC-MS. Extracellular lactic acid quantities were depicted. Shown are standard deviation of n=3 replicates.  $p < 0.05$  was indicated with asterisk (unpaired two-tailed *t* Test).

**Supplement Figure 4:** **A**) CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup> and CaCO<sub>2</sub>-KRAS<sup>G12V</sup> cells were cultivated with indicated glucose concentrations and analyzed with quantitative real time PCR for *MCT1* and *MCT4* expression. *PGK1* served as loading control. Relative mRNA expression to CaCO<sub>2</sub>-control cultured with 1.0 g/L glucose was shown. **B, C**) CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup>, CaCO<sub>2</sub>-KRAS<sup>G12V</sup>, HT29 and SW480 cells cultivated in medium containing 1.0 g/L or 2.5 g/L glucose

concentrations and treated with 0.1  $\mu$ M SR13800 for 24 h were analyzed using quantitative real time PCR for *MCT1* and *MCT4* expression. *PGKI* served as loading control. Relative mRNA expression to DMSO was shown. glc: glucose. A-C) Shown are standard deviation of (in minimum) n=3 replicates. p<0.05 was indicated with asterisk (unpaired two-tailed *t* Test).

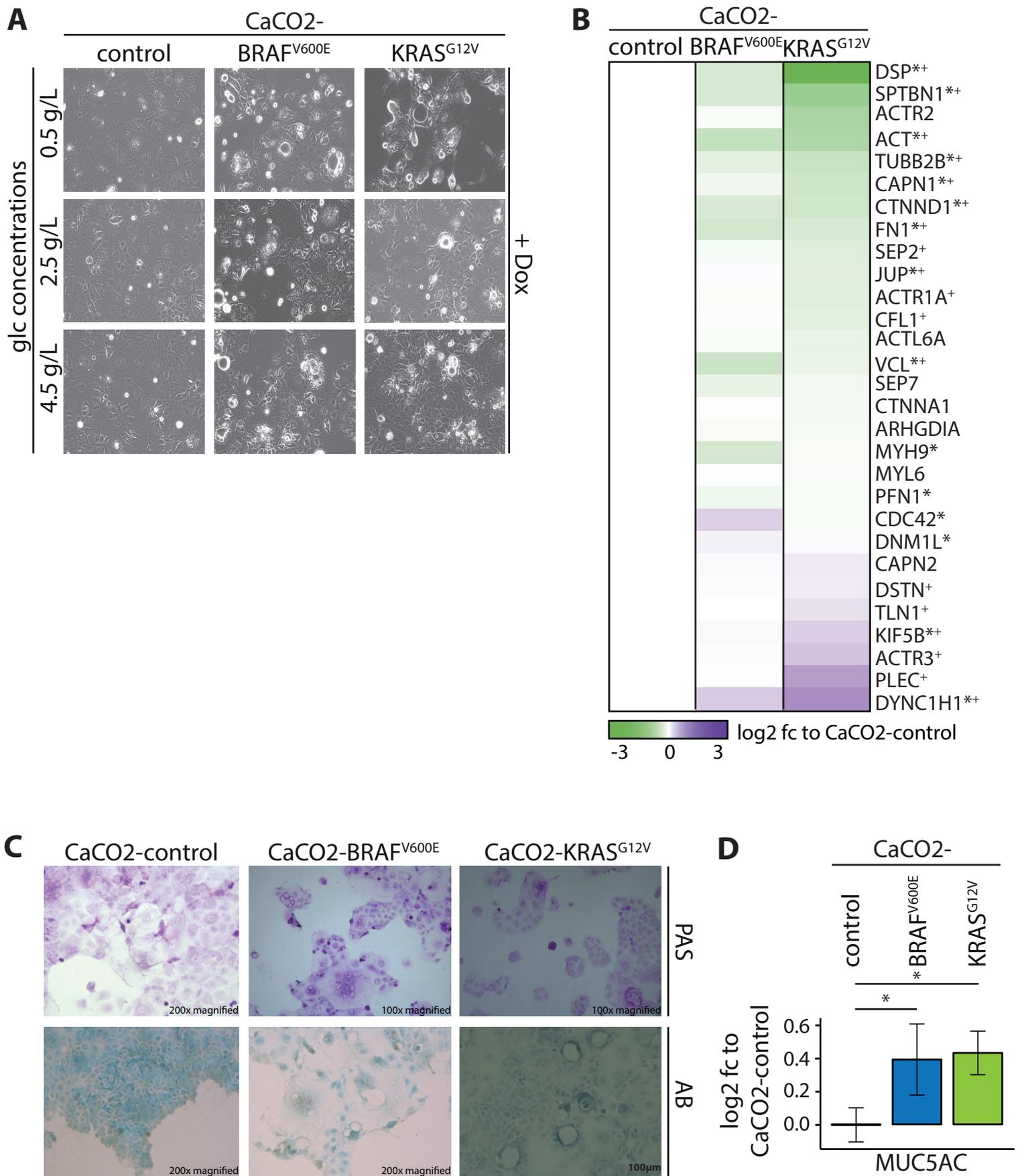
**Supplement Figure 5:** **A)** CaCO<sub>2</sub>-control, CaCO<sub>2</sub>-BRAF<sup>V600E</sup> and CaCO<sub>2</sub>-KRAS<sup>G12V</sup> cells were subcutaneously injected into the right flank of mice in the presence of Doxycycline (Dox) every 2 d by intraperitoneal injection. Mean of tumor volume is shown for n=3 mice per group up to 14 d. **B)** CaCO<sub>2</sub>-KRAS<sup>G12V</sup> were subcutaneously injected into the right flank of mice receiving 200  $\mu$ L PBS or BrPy (8 mg/kg) in the presence of Doxycycline treatment every 2 days (starting from day 8) by intraperitoneal injection. Hematoxylin eosin staining (H&E) staining was performed from paraffin embedded sections.

**Supplementary Information for blots:**

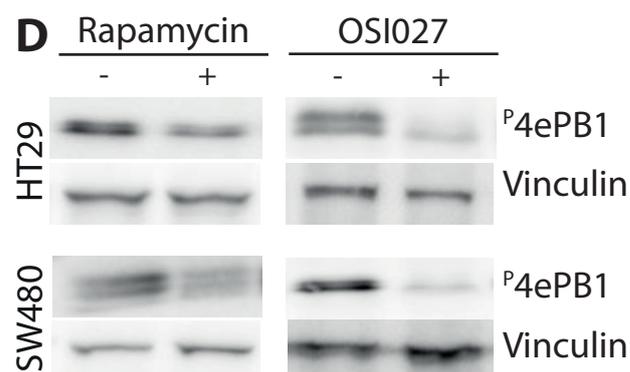
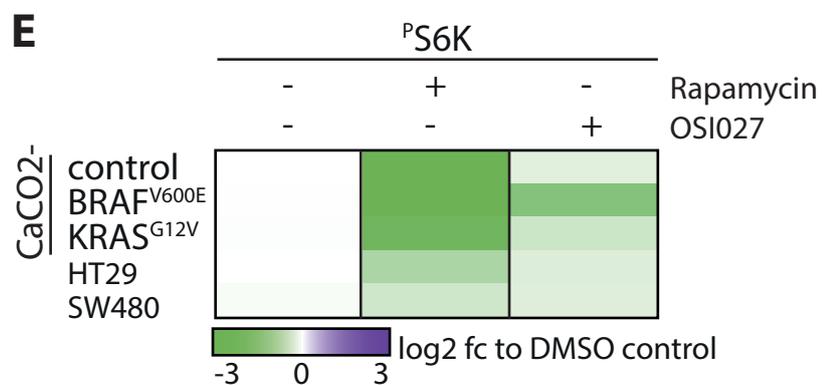
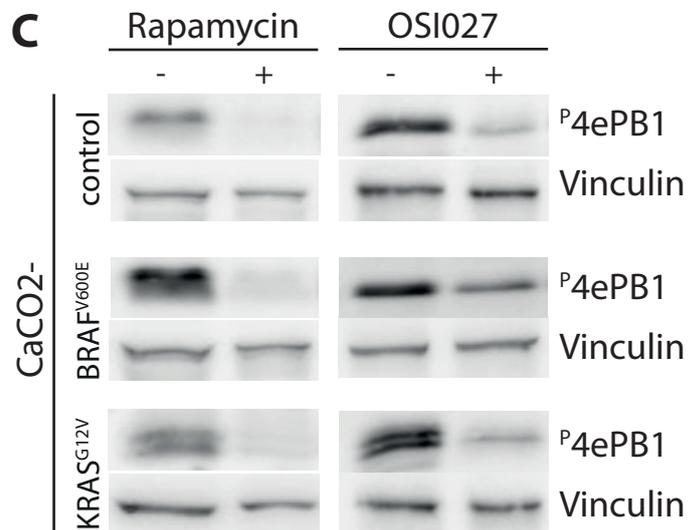
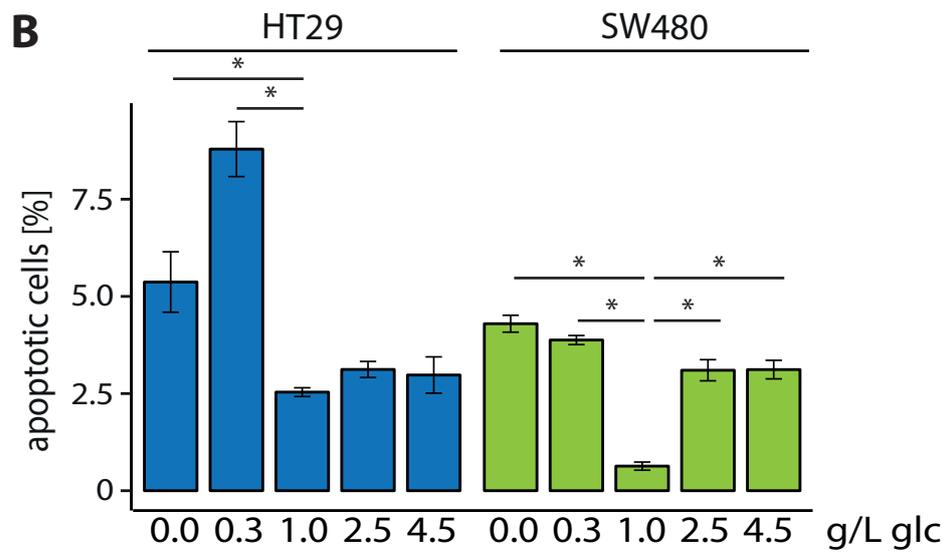
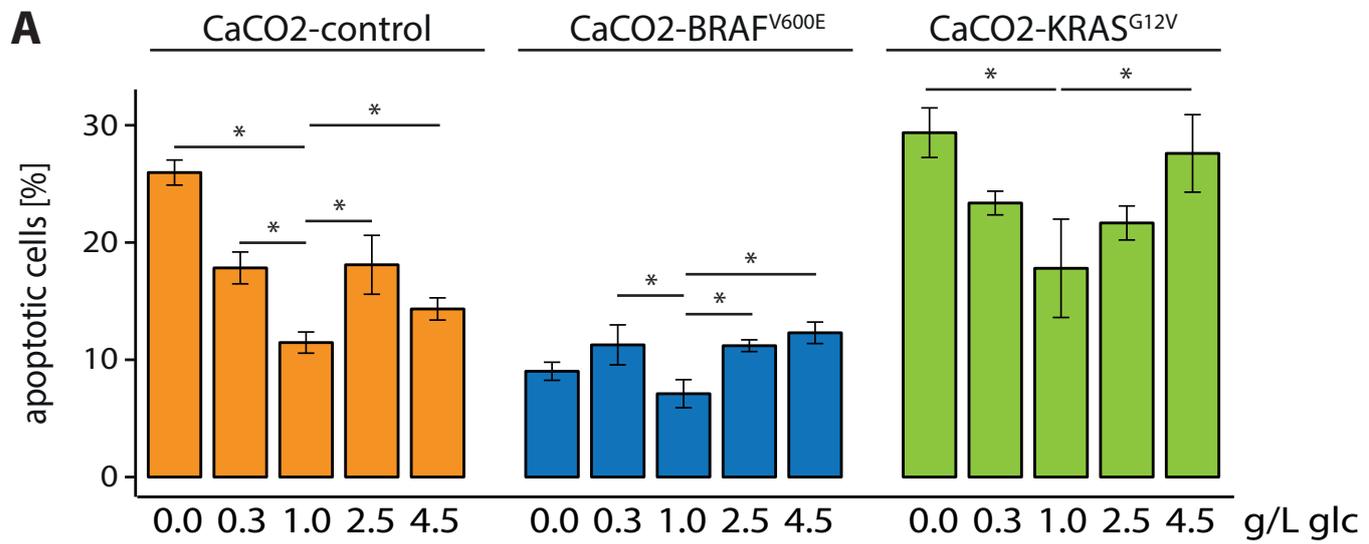
Full length blot to Figure 3A

Full length blot to Figure 3B

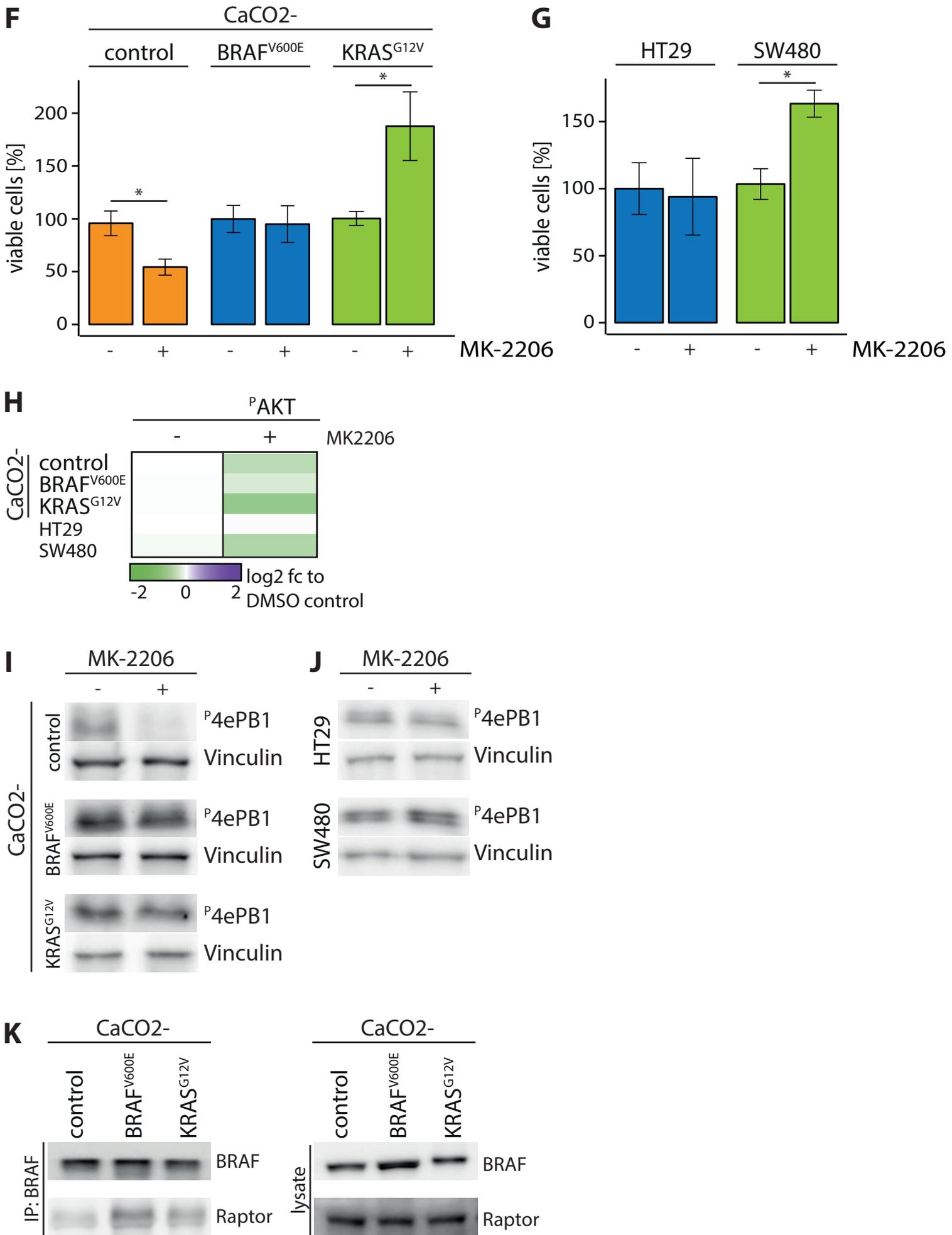
Full length blot to Figure 6D



Supplement Figure 1

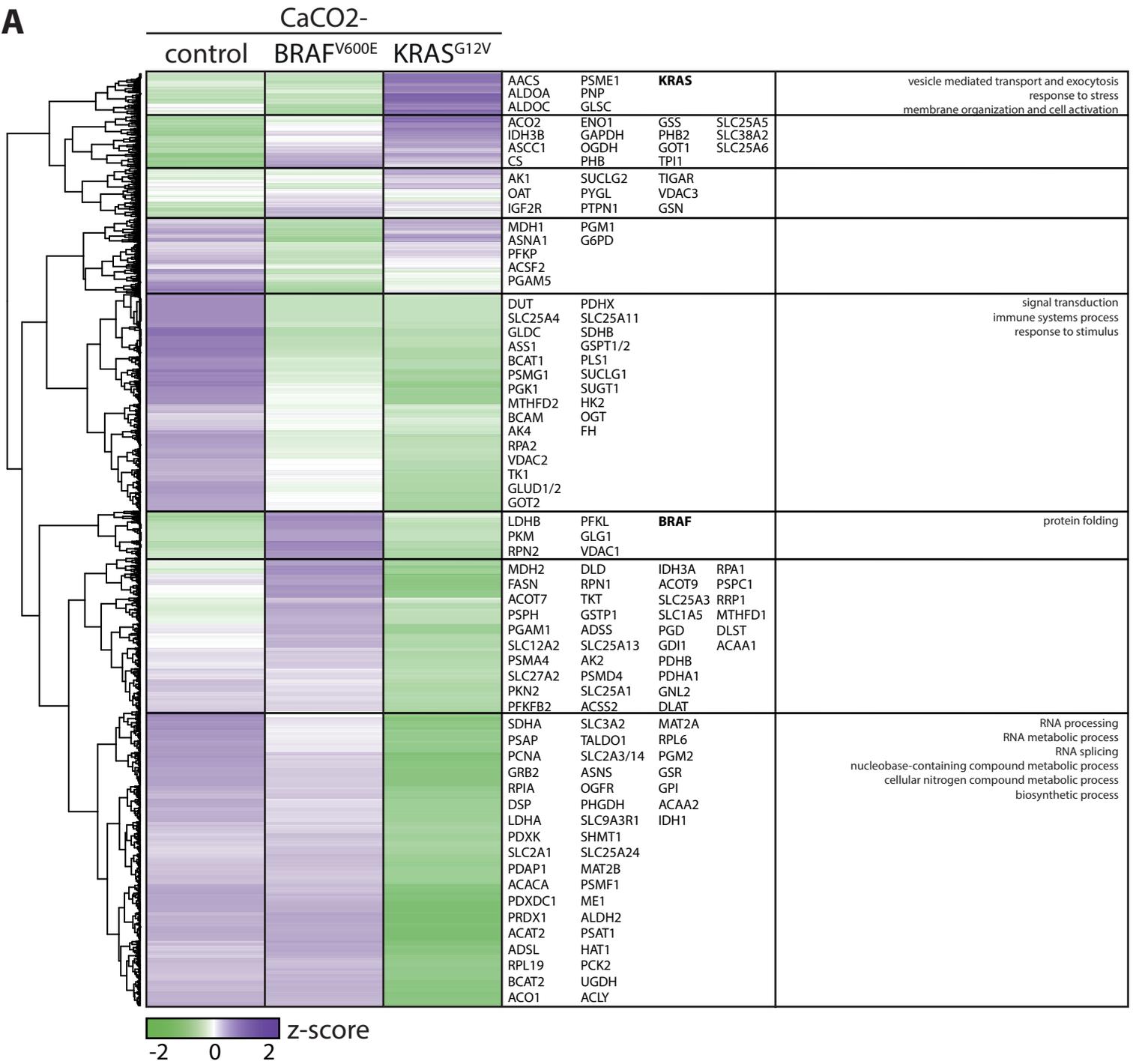


**Supplement Figure 2**



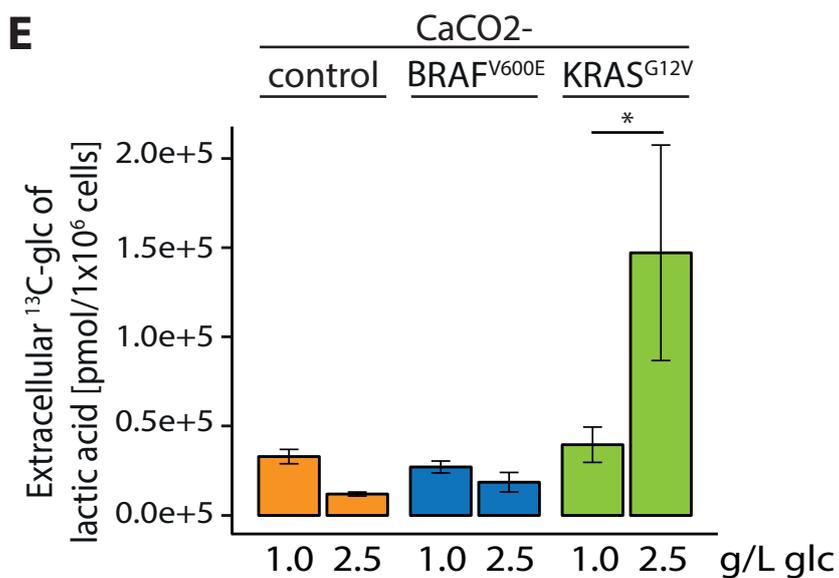
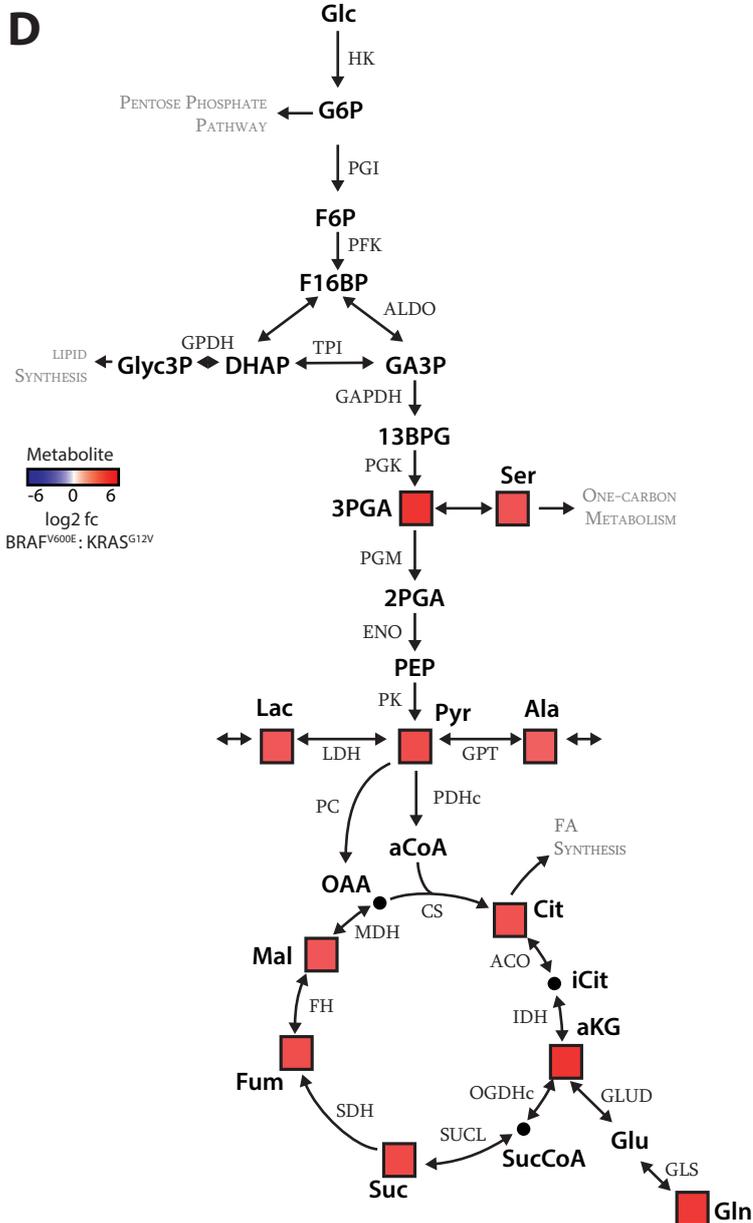
**Supplement Figure 2**

**A**

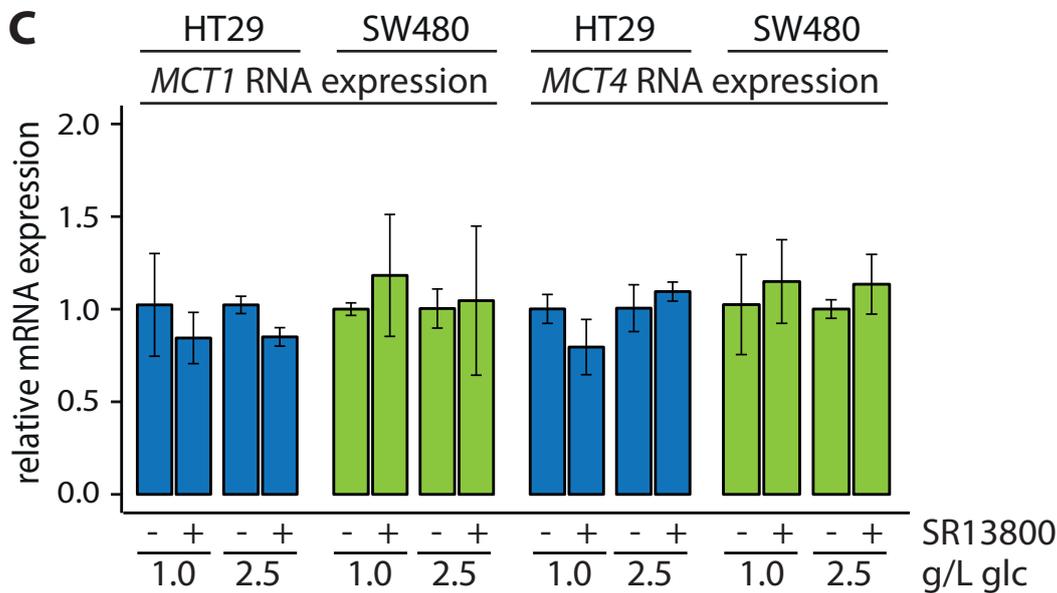
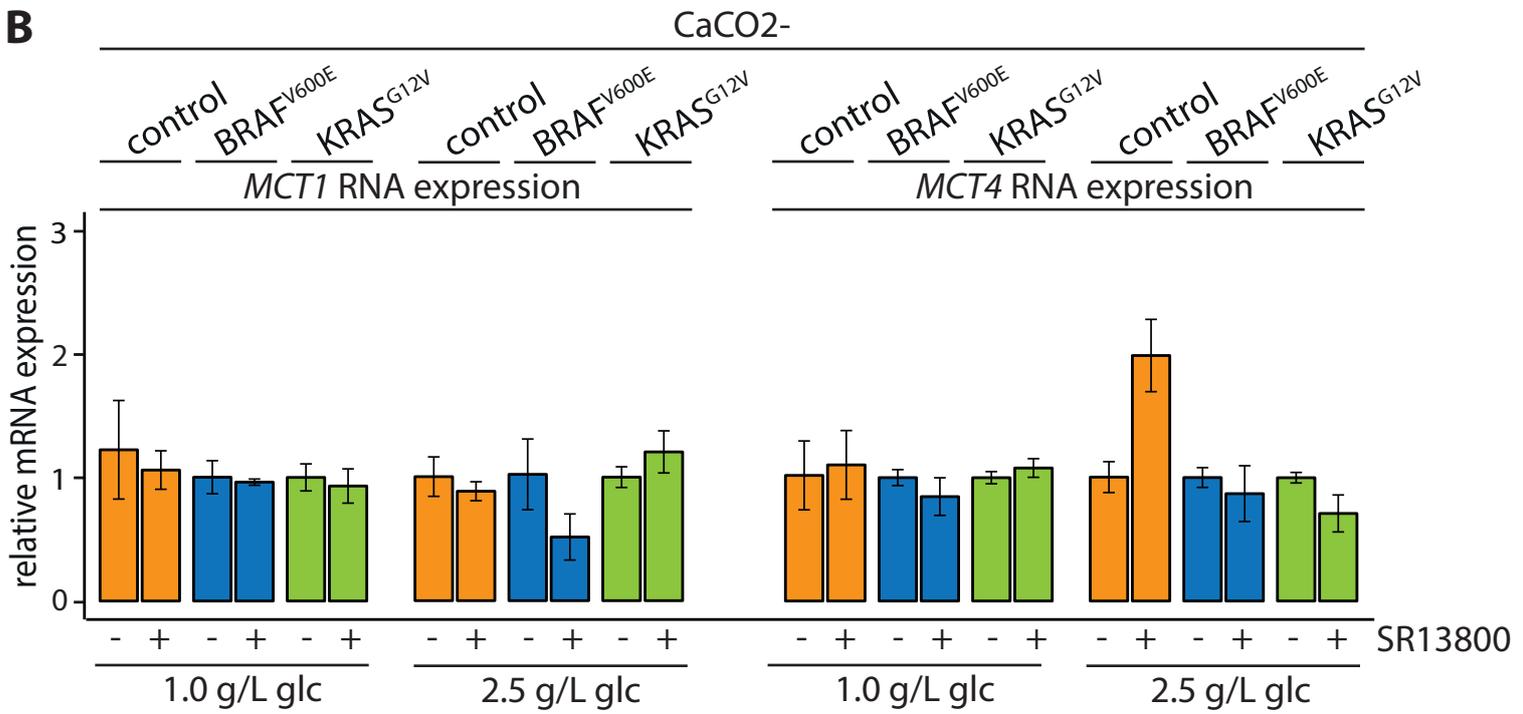
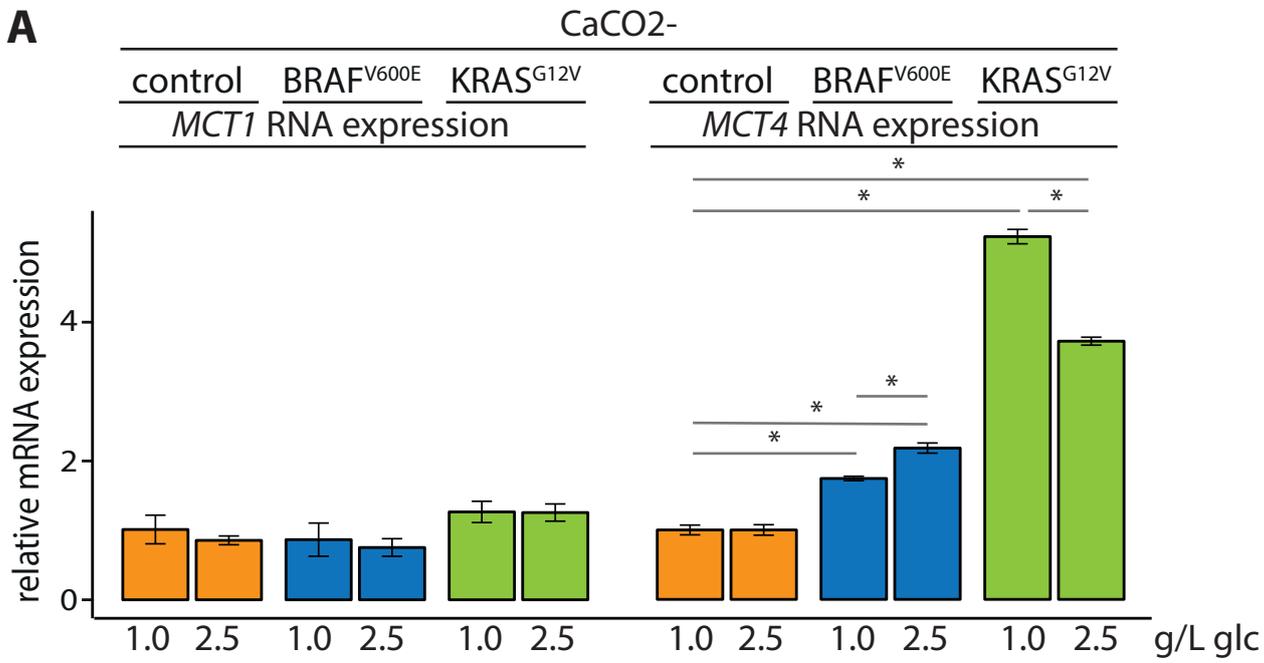


**Supplement Figure 3**

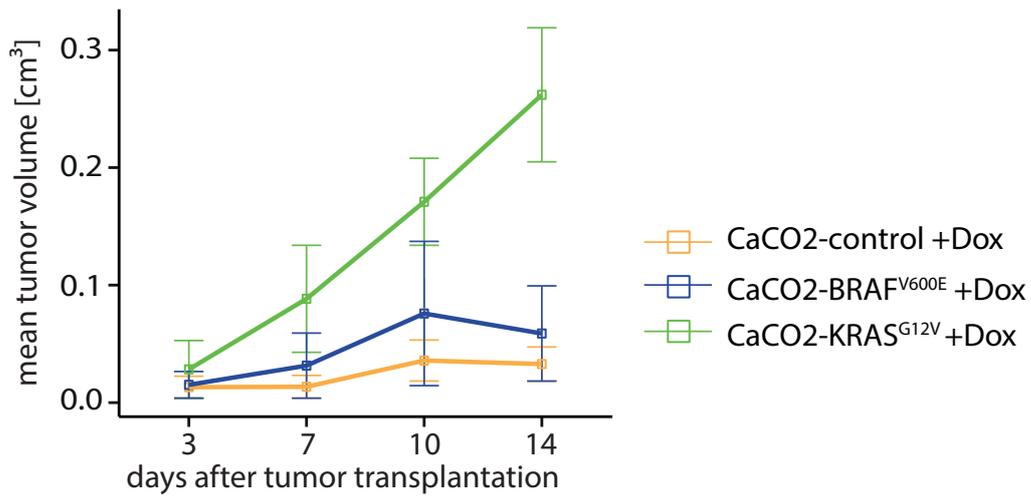
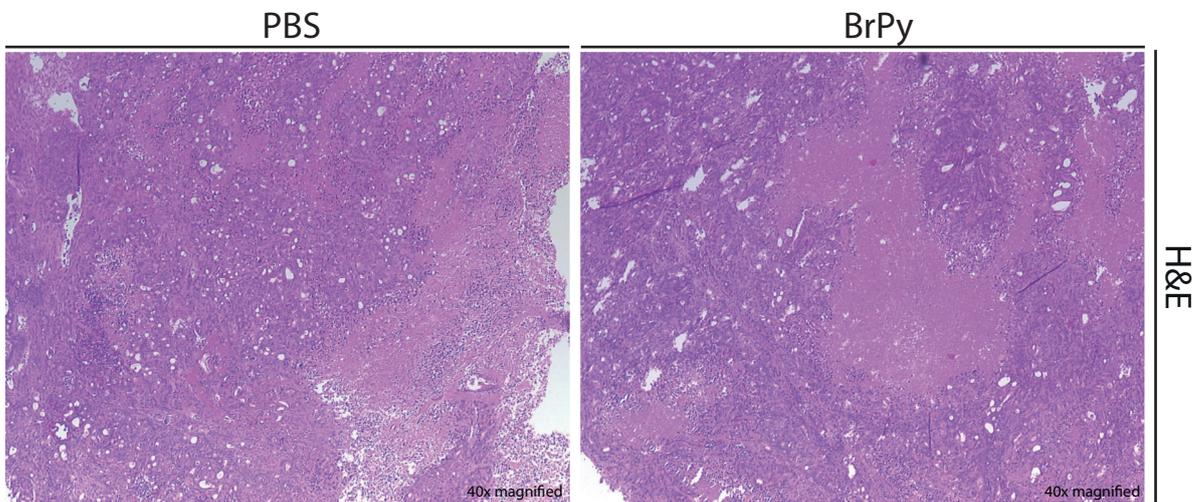




**Supplement Figure 3**

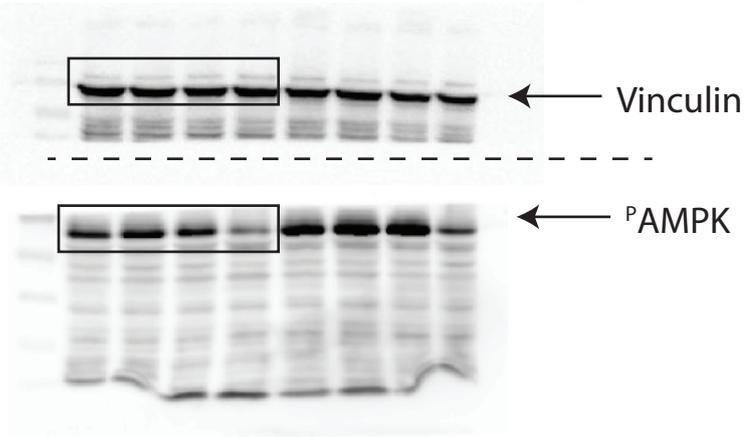


**Supplementary Figure 4**

**A****B**CaCO<sub>2</sub>-KRAS<sup>G12V</sup>

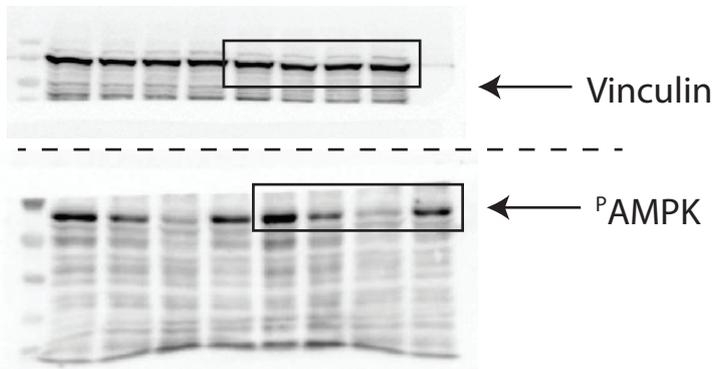
CaCO2-BRAF<sup>V600E</sup>

0.0 0.3 1.0 2.5 0.0 0.3 1.0 2.5 g/L glc



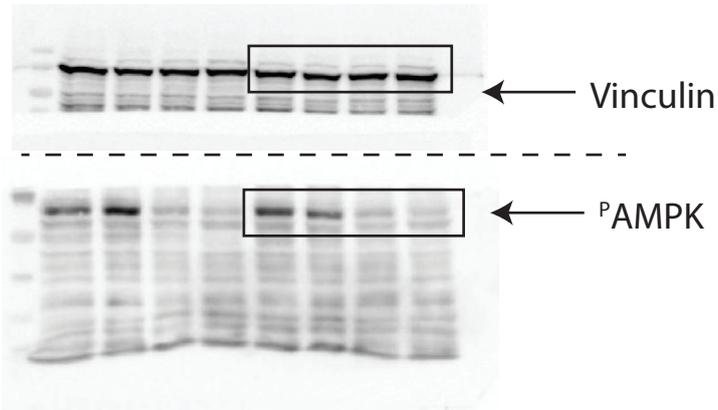
CaCO2-KRAS<sup>G12V</sup>

0.0 0.3 1.0 2.5 0.0 0.3 1.0 2.5 g/L glc



CaCO2-control

0.0 0.3 1.0 2.5 0.0 0.3 1.0 2.5 g/L glc



Full length blots to Figure 3A

SW480

0.0 0.3 1.0 2.5 0.0 0.3 1.0 2.5 0.3 1.0 2.5 g/L glc  
← Vinculin

← <sup>P</sup>AMPK

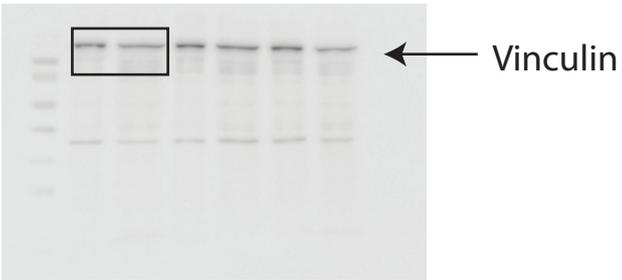
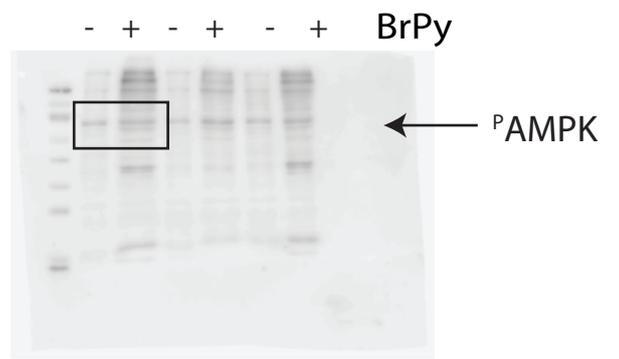
HT29

0.0 0.3 1.0 2.5 0.0 0.3 1.0 2.5 0.0 0.3 1.0 2.5 g/L glc  
← Vinculin

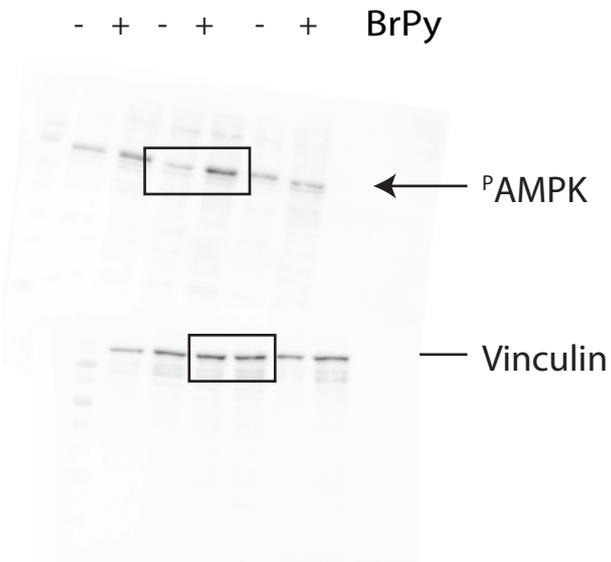
← <sup>P</sup>AMPK

Full length blots to Figure 3B

CaCO2-BRAF<sup>V600E</sup>



CaCO2-KRAS<sup>G12V</sup>



Full length blots to Figure 6D