

Characterization of a Single Genomic Locus Encoding the Clustered Protocadherin Receptor Diversity in *Xenopus tropicalis*

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ABSTRACT Clustered protocadherins (cPcdhs) constitute the largest subgroup of the cadherin superfamily, and in mammals are grouped into clusters of α -, β -, and γ -types. Tens of tandemly arranged paralogous Pcdh genes of the Pcdh clusters generate a substantial diversity of receptor isoforms. cPcdhs are known to have important roles in neuronal development, and genetic alterations of cPcdhs have been found to be associated with several neurological diseases. Here, we present a first characterization of cPcdhs in *Xenopus tropicalis*. We determined and annotated all cPcdh isoforms, revealing that they are present in a single chromosomal locus. We validated a total of 96 isoforms, which we show are organized in three distinct clusters. The *X. tropicalis* cPcdh locus is composed of one α - and two distinct γ -Pcdh clusters (*pcdh- γ 1* and *pcdh- γ 2*). Bioinformatics analyses assisted by genomic BAC clone sequencing showed that the *X. tropicalis* α - and γ -Pcdhs are conserved at the cluster level, but, unlike mammals, *X. tropicalis* does not contain a β -Pcdh cluster. In contrast, the number of γ -Pcdh isoforms has expanded, possibly due to lineage-specific gene duplications. Interestingly, the number of *X. tropicalis* α -Pcdhs is identical between *X. tropicalis* and mouse. Moreover, we find highly conserved as well as novel promoter elements potentially involved in regulating the cluster-specific expression of cPcdh isoforms. This study provides important information for the understanding of the evolutionary history of cPcdh genes and future mechanistic studies. It provides an annotated *X. tropicalis* cPcdh genomic map and a first molecular characterization essential for functional and comparative studies.

KEYWORDS

Xenopus tropicalis
gene cluster
neuronal development
protocadherin receptor
diversity

The term “protocadherin” (Pcdh) was coined by Suzuki and his colleagues in 1993 (Sano *et al.* 1993), and refers to a large number of cadherin-like cell surface receptors, which are expressed primarily in the nervous system. Clustered protocadherins (cPcdhs) were first dis-

covered in mammals, in which the majority of the Pcdh genes are organized in tandem in a single genomic locus, hence the name “clustered protocadherins” (Kohmura *et al.* 1998; Wu and Maniatis 1999).

Mouse cPcdh genes are organized into α , β , and γ clusters, containing 14, 22, and 22 genes, respectively. The α and γ clusters contain “variable exons” (VEs) as well as “constant exons” (CEs), the latter being located at the distal ends of the clusters. In each mature α - and γ -Pcdh transcript, a VE is spliced to three CEs. The VEs code for the six extracellular cadherin repeats (EC1-6), the transmembrane domain, and a short cytoplasmic tail. The CEs, which are shared by all the isoforms of the same cluster, code for most of the cytoplasmic domain, and are referred to as the “constant domain.” The β cluster is composed of tandem-arrayed single-exon genes lacking constant domains but still retaining a short cytoplasmic domain encoded by each of the VEs.

Based on the phylogenetic analyses of human cPcdhs, two of the 14 α -Pcdhs, and three of the 22 γ -Pcdhs, which form a distinct paraphyletic group, are referred to as the “C-type” cPcdhs. The remaining isoforms are referred to as “non C-type.” For γ -Pcdh, these non C-type

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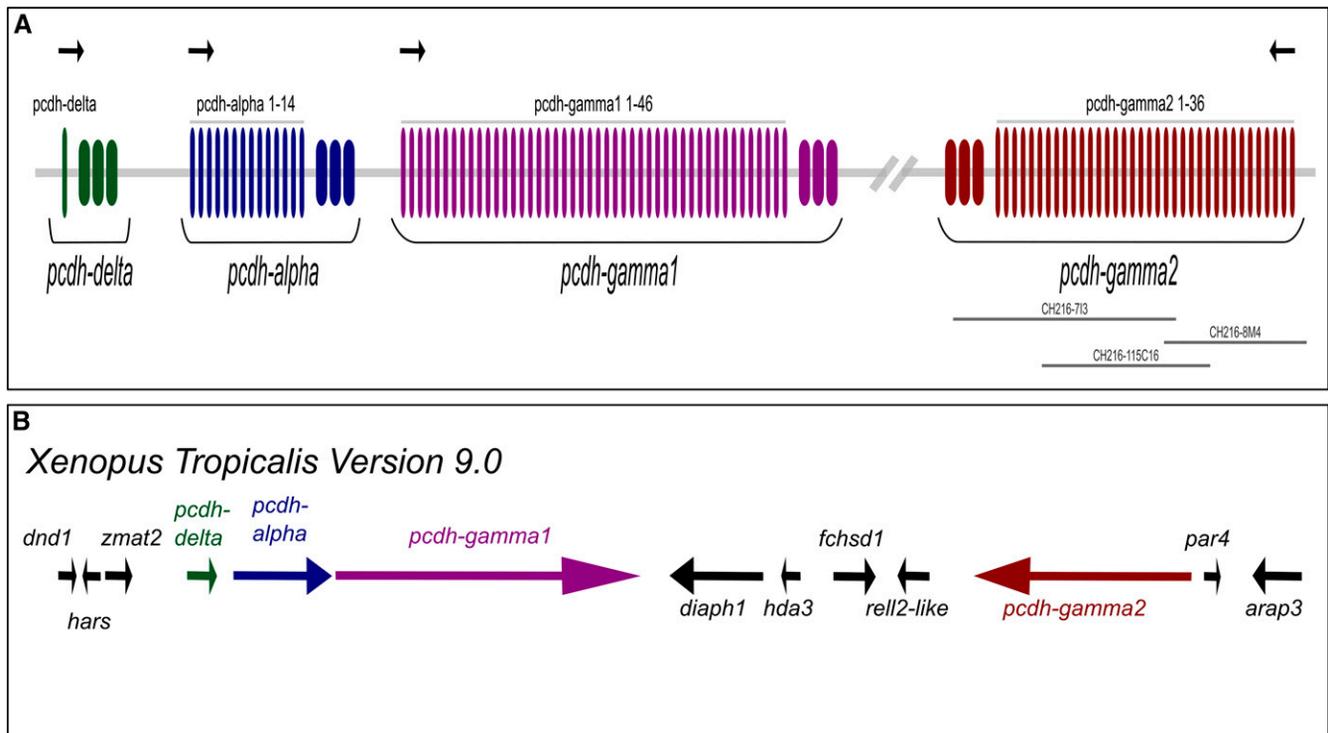


Figure 1 Genomic organization of the *X. tropicalis* cPcdhs. (A) Representative diagram showing *X. tropicalis* cPcdhs gene models. Longer and thinner rectangles denote variable exons, shorter and thicker rectangles denote constant exons. Green: *pcdh- δ* , blue: *pcdh- α* , purple: *pcdh- γ 1*, red: *pcdh- γ 2*. Black arrows above the gene models denote the orientation of the clusters. Gray bars below the gene models correspond to the sequenced BAC clones. (B) Genomic map of *X. tropicalis* cPcdhs loci based on genome build 9.0.

cPcdhs are grouped as either “A-type” or “B-type” (Wu and Maniatis 1999). Single-cell PCR studies of cerebellar Purkinje cells revealed that the five C-type isoforms are expressed biallelically and broadly, while the remaining α - and γ -Pcdhs, as well as all of the β -Pcdhs, are expressed monoallelically and in a restricted fashion (Chess 2005; Kaneko *et al.* 2006; Hirano *et al.* 2012). The work of Chen *et al.* (2012) suggested differential roles for the three C-type γ -Pcdhs, as the deletion of these genes resulted in apoptotic loss, or reduced numbers, of some neuronal populations in the spinal cord and in the retina, respectively. In contrast, the deletion of three A-type γ -Pcdhs did not cause any detectable phenotypes (Chen *et al.* 2012).

Studies on the expression of different cPcdhs have discovered several distinct promoter elements, enhancers, as well as intra and intergenic regulatory elements (Wu *et al.* 2001; Golan-Mashiach *et al.* 2012; Ribich *et al.* 2006; Yokota *et al.* 2011; Guo *et al.* 2012; Tan *et al.* 2010). A conserved sequence element (CSE), which is present in most mammalian cPcdh promoters, was shown to constitute a binding site for the CCCTC-binding factor (CTCF) / cohesin complex, which regulates transcription via chromatin modifications and looping (Kehayova *et al.* 2011).

cPcdhs were initially thought to be a vertebrate innovation since they are absent in the invertebrates, while nonclustered Pcdhs, which predate bilaterians, are present in the invertebrate genomes (Hulpiau and van Roy 2009). Recently, Pcdh clusters were identified in octopus; however, vertebrate cPcdhs and clustered octopus Pcdhs do not appear to be direct orthologous genes, but likely result from convergent evolution (Albertin *et al.* 2015).

cPcdh loci are among the most dynamic in vertebrate genomes. Even the slowly evolving genomes of elephant shark and coelacanth have rather dynamic Pcdh loci (Noonan *et al.* 2004a; Yu *et al.* 2008). It was reported that Pcdh clusters experienced repetitive gene duplication, gene loss, gene degeneration, adaptive variation, and gene conversion

events (Noonan *et al.* 2004b; Wu 2005). Despite the dynamic nature of Pcdh clusters, a synteny of genes flanking the Pcdh loci is still present, even between mammals and cartilaginous fishes (Yu *et al.* 2008). The elephant shark genome was shown to contain δ -, ϵ -, μ -, and ν -type cPcdhs, but not α -, β -, or γ -Pcdhs, suggesting that the ancestral vertebrate genome contained a vast repertoire of different Pcdh subclusters, which then formed the modern Pcdh clusters via repetitive lineage-specific gene duplications and losses (Yu *et al.* 2008). The δ -Pcdh subcluster is composed of a single gene. It was shown to be an ancient member of the cPcdhs, being present in diverse vertebrate clades but lost in the mammalian lineage (Yu *et al.* 2008). Zebrafish and fugu, two representative species from the teleost fish lineage, were shown to contain two unlinked Pcdh loci in their genomes (Noonan *et al.* 2004a; Yu *et al.* 2008), due to the whole genome duplication event of teleost fishes. On the other hand, there is only a single cPcdh locus in the ancient genome of coelacanth, a representative of the nonteleost lineage (Noonan *et al.* 2004a).

The complex genomic organization and diverse regulation mechanisms of cPcdhs account for their remarkable diversity in the vertebrate nervous system. cPcdhs were shown to play roles in neuronal self avoidance thanks to their combinatorial expression patterns and strictly homophilic interactions. In this respect, cPcdhs demonstrate striking functional similarity with the *Drosophila* Dscam1 (Zipursky and Sanes 2010; Kise and Schmucker 2013).

The frog, *Xenopus tropicalis*, serves as a model organism, especially for developmental biology, and significant effort has been put into developing molecular tools for genetic analysis analogous to the powerful genetics available for the analysis of mouse. Importantly, the full diploid genome of *X. tropicalis* was recently sequenced, although some gene annotations and gene models still need to be established (Hellsten *et al.* 2010). This applies also to the full *X. tropicalis* cPcdh locus.

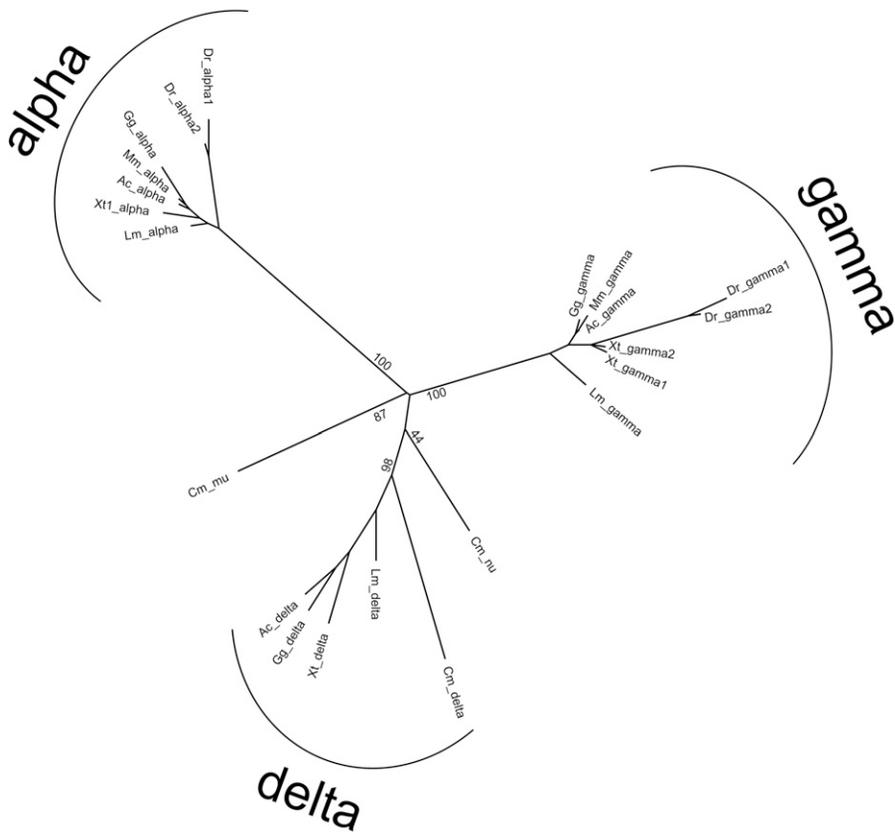


Figure 2 Phylogenetic analysis of cPcdhs at the cluster level. The tree was generated using ML and is based on the multiple sequence alignment of the constant domains of different Pcdh clusters. % Bootstrap values of major nodes are shown. Ac, anole; Cm, elephant shark; Dr, zebrafish; Gg, chicken; Lm, coelacanth; Mm, mouse; Xt, *X. tropicalis*.

In this study, we used cDNA and BAC sequencing, RNA *in situ* expression analysis, as well as bioinformatics analyses, to characterize *X. tropicalis* cPcdhs. We show that there is a single cPcdh locus in the *X. tropicalis* genome containing three clusters, analogous to the mammalian genome organization. In contrast to the mouse locus, however, the *X. tropicalis* genome lacks β -Pcdh and instead contains a novel additional γ -cluster (Figure 1). The genomic locus consists of one α , one γ 1, and one γ 2 cluster. *X. tropicalis* and mouse have the same number of α -Pcdh isoforms (14 in both), while the two *X. tropicalis* γ clusters are significantly larger, with γ 1 containing 46 and γ 2 containing 36 isoforms. The expansion appears to reflect extensive lineage-specific gene duplication events (58 isoforms in mouse and 96 in *X. tropicalis*). We conducted SMRT sequencing of a BAC clone targeting the γ 2 locus (Figure 1), and validated that our sequencing results are in good agreement with sequences released by the *X. tropicalis* sequencing project. We detected low-level gene conversion events exercising on the *X. tropicalis* cPcdhs, at levels comparable to those of mouse cPcdhs.

MATERIALS AND METHODS

Gene annotation

X. tropicalis Genome Build 7 (JGI 7.1/xenTro7) was used for the gene annotation. The annotation was performed manually; gene models were predicted by open reading frame analysis and GENSCAN (Burge and Karlin 1997), and were validated experimentally by the analysis of *X. tropicalis* EST clones from public databases, analysis of cDNA clones from our cluster specific 5'RACE reactions, and re-analysis of publicly available RNA-seq data (Tan *et al.* 2013; Paranjpe *et al.* 2013). Gene-specific 5'RACE primers were designed so as to detect the 3'UTR regions of each cluster in order to be able to detect all of the isoforms of a given cluster. The 5'RACE sequences for the α , γ 1 and γ 2 clusters are 5'-GGAAGGTGCATCAACAGTAG

GAAGAA-3', 5'-TGCCCTGTTGGTGTTCAGCCAATC-3', and 5'-ACCAATTCGCTTGGGGAATTCTTCTGGGG-3', respectively.

BAC sequencing

X. tropicalis BAC clone CH216-115C16 was obtained from the CHORI BACPAC Resource Center and purified with the Epicenter BACMAX DNA Purification Kit. Sequencing was performed with Pacific Biosciences SMRT technology using the standard 5-kb template preparation protocol, C2 chemistry, and P4 polymerase, with Magbead loading on SMRT cells, and with a 120-min movie time.

Phylogenetic analysis

Multiple sequence alignments were produced with CLUSTALW (Sievers *et al.* 2011). The alignments were used to estimate the maximum likelihood (ML) phylogeny with PhyML (Guindon *et al.* 2010), with parameters determined by ModelGenerator (Keane *et al.* 2006).

Gene conversion

Synonymous substitution rates were estimated with the yn00 program of the PAML package (Yang 2007). Nucleic acid multiple sequence alignments were generated by the RevTrans program (Wernersson and Pedersen 2003) based on amino acid multiple sequence alignments generated with CLUSTALW (Sievers *et al.* 2011).

Promoter motif discovery

Promoter regions (2 kb upstream of the translational start codon) were scanned with the MEME suite (Bailey *et al.* 2009).

RNA-seq analysis

Publicly available RNA-seq data were retrieved from the European Nucleotide Archive. Adapter and quality trimming were performed with

■ **Table 1** Percent amino acid sequence identity values of the extracellular and cytoplasmic domains of mouse and *X. tropicalis* cPcdhs

Domain	A Average of All Mouse α Isoforms	B Average of All Xt α Isoforms	C Mouse Pcdha9 vs. Xt pcdh- α 1	D Average of All Mouse γ Isoforms	E Average of All Xt γ 1 Isoforms	F Average of All Xt γ 2 Isoforms	G Mouse Pcdhga3 vs. Xt pcdh- γ 1 3	H Mouse Pcdhga3 vs. Xt pcdh- γ 2 35
EC1	95.85	70.79	57.83	55.80	71.01	56.37	19.85	15.44
EC2	69.29	68.06	52.48	57.63	68.65	58.63	23.91	26.21
EC3	58.52	59.30	45.00	50.48	64.69	51.81	17.65	14.60
EC4	85.49	61.60	49.48	59.32	71.25	57.08	17.16	16.42
EC5	98.01	73.40	46.08	62.76	68.97	59.17	23.74	19.42
EC6	84.31	56.22	38.89	81.62	79.67	62.59	19.35	24.19
CD	100.00	100.00	68.39	100.00	100.00	100.00	75.00	73.60

Columns A, B, D, E, and F show average percent sequence identity values of mouse α , Xt α , mouse γ , Xt γ 1, and Xt γ 2 clusters, respectively. Columns C, G, and H show percent sequence identity values of mouse Pcdha9–Xt pcdh- α 1, mouse Pcdhga3–Xt pcdh- γ 1 3, and mouse Pcdhga3–Xt pcdh- γ 2 35 pairs, respectively. EC, extracellular domain; CD, constant domain.

Trimmomatic (Bolger *et al.* 2014), and reads were mapped to xenTro7 with Tophat2 (Trapnell *et al.* 2009) using the option “–b2-very-sensitive.” Uniquely mapped read-pairs/reads were retrieved with Samtools (Li *et al.* 2009) and were fed to HTSeq for count-based expression analysis (Anders *et al.* 2013). The expression was reported as FPKM or RPKM for paired-end or single-read sequencing, respectively.

RNA in situ hybridization

Whole mount RNA *in situ* hybridization experiments were performed according to the protocol introduced by Harland (1991). The *pan-gamma* probe targets the constant exons of the γ 1 cluster, while the *gamma*1, *gamma*2, and *alpha* probes recognize the 3'UTR regions of the γ 1, γ 2, and α clusters, respectively. The PCR primer sequences are as follows: *pan-gamma* forward: 5'-CAGGCGCAGCCGAACG-3', *pan-gamma* reverse: 5'-CAGGCGCAGCCGAACG-3'; *gamma*1 forward: 5'-CAGGCGCAGCCGAACG-3', *gamma*1 reverse: 5'-GTTCCACTCAGACCCAACTCT-3', *gamma*2 forward: 5'-CAGGCGCAACCTAACGCAGATTGGCGGT-3', *gamma*2 reverse: 5'-TCATTGTTTTGCGTTCTGCGT-3'; *alpha* forward: 5'-CCCAAATCATCCTCATCCA-3'; *alpha* reverse: 5'-TCATTTATCGTTGTTGT CAGCT-3'.

Data availability

The authors state that all data necessary for confirming the conclusions presented in the article are represented fully within the article.

RESULTS

X. tropicalis locus containing three variable Pcdh clusters

The gene models of *X. tropicalis* Genome Builds 7 (xenTro7) and 8 (xenTro8) related to proposed cPcdhs had been incomplete at the outset of this analysis. In order to identify genomic regions corresponding to cPcdh genes, we performed TBLASTN searches using mouse α -, β -, and γ -Pcdh, and lizard δ -Pcdh, sequences as queries and the *X. tropicalis* Genome Build 7 (xenTro7) as the subject. We identified an α cluster, two γ clusters, and a single Pcdh- δ gene in a single scaffold (scaffold_3). We did not identify a β cluster, which, in mouse, is situated between α and γ clusters.

The newly identified clusters were annotated manually on Genome Build 7 (xenTro7). First, we created gene models using open reading frame analysis (cutoff: 700 amino acids), and the GENSCAN ab initio gene prediction algorithm (Burge and Karlin 1997). Gene models that fail to code for six cadherin repeats were filtered out.

Experimental support for the gene models came from different sources: 1) we performed RACE experiments in order to generate *X. tropicalis* cPcdh cDNA clones. RACE reactions were performed with total RNA from various developmental stages, and the cluster-specific 5'RACE primers targeted the constant exons of the cluster of interest. 2) We screened the *X. tropicalis* EST databases for cPcdh EST clones. 3) We obtained publicly available *X. tropicalis* RNA-seq studies, and reanalyzed the raw data.

Our annotation efforts resulted in the identification of 14, 46, and 36 genes for the *pcdh- α* , *pcdh- γ 1* and *pcdh- γ 2* clusters, respectively. In addition, a single *pcdh- δ* gene was also identified.

While this manuscript was in preparation, *X. tropicalis* Genome Build 9 (xenTro9) was released. All of the aforementioned Pcdh clusters are present on chromosome 3 of this genome build, and Pcdh cluster sequences in the two genome builds demonstrate > 99% sequence identity. Moreover, the *X. tropicalis* cPcdh locus is syntenic with the two *X. laevis* cPcdh loci that are located on the short and long third chromosomes XLA3S and XLA3L, respectively (data not shown). We therefore based our cPcdh genomic map on Genome Build 9 (Figure 1).

The clusters are flanked by non-Pcdh genes, suggesting that cPcdh clusters, as annotated in Figure 1, are likely to be complete Pcdh clusters. The *pcdh- γ 1* cluster can be regarded as syntenic with the mouse γ -Pcdh cluster, while the *pcdh- γ 2* cluster potentially originated from a duplication of the *pcdh- γ 1* cluster, and this duplication does not include any flanking genes.

The assembly of *X. tropicalis* Pcdh clusters is of high quality

Sequence analysis of the paralogous genes of the *X. tropicalis* Pcdh clusters revealed high similarity between some of the paralogous genes. We defined similarity as percent sequence identity of isoform pairs over their second and third extracellular domains (EC2–EC3) at the amino acid level. EC2–EC3 was chosen as these two ecto-domains were shown to be responsible for providing the specificity of the strictly homophilic interactions of α -, β -, and γ -Pcdhs (Schreiner and Weiner 2010; Thu *et al.* 2014; Rubinstein *et al.* 2015; Nicoludis *et al.* 2015), and hence are expected to be diverse in sequence. Comparison of EC2–EC3 regions of *X. tropicalis* cPcdh isoforms at the amino acid level showed that a large number of *X. tropicalis* cPcdh isoforms demonstrate high sequence identity to at least one other isoform (Supplemental Material, Figure S1). In contrast, the number of “highly similar” mouse cPcdh isoforms is relatively low (Figure S1).

The *X. tropicalis* genome was sequenced with a whole genome shotgun sequencing approach, which sometimes impairs the sequence

■ **Table 2 Average synonymous substitutions per codon (dS)**

	dS _{EC1}	dS _{EC2}	dS _{EC3}	dS _{EC4}	dS _{EC5}	dS _{EC6}	dS _{highest} /dS _{lowest}
Xt pcdh- α 1-14	1.15	2.84	2.11	1.84	1.27	1.35	2.47
Xt pcdh- γ 2 20-26	0.28	0.33	0.68	0.43	0.44	0.33	2.46
Xt pcdh- γ 2 28-35	0.46	0.45	0.79	0.47	0.82	0.15	5.35
Xt pcdh- γ 2 1-13	1.24	2.55	2.62	1.44	1.13	1.72	2.32
Xt pcdh- γ 1 42-46	0.26	1.06	0.63	0.50	1.02	0.53	4.06
Xt pcdh- γ 1 1-41	1.67	1.76	1.83	1.64	1.23	0.89	2.04
Mm Pcdha1-12	0.31	2.08	2.15	1.71	0.27	1.35	7.86
Mm Pcdhb1-22	1.08	1.66	1.43	1.60	0.79	1.36	2.10
Mm Pcdhga1-12	2.16	2.29	2.45	2.61	2.34	1.88	1.39
Mm Pcdhgb1-8	1.38	1.90	2.21	1.54	0.60	1.50	3.70
Dr Pcdh2ab1-12	1.67	1.13	2.91	1.13	0.12	0.01	231.14

Phylogenetic analysis of *X. tropicalis* cPcdhs

In order to investigate the evolutionary relationships of *X. tropicalis* cPcdhs at the cluster level, we constructed a phylogenetic tree using multiple sequence alignments of the constant domains of the cPcdhs of mouse, chicken, lizard, zebrafish, coelacanth, and elephant shark, which represent mammals, birds, reptiles, teleost fish, and lobe-finned fish, respectively, along with the constant domains of *X. tropicalis* cPcdhs (Figure 2). The results confirmed that *X. tropicalis* clusters are direct orthologs of mouse α - and γ -, as well as fish δ -Pcdhs. The phylogenetic tree shows that α - and γ -Pcdhs have a slower evolutionary rate than δ -Pcdhs, suggesting potentially more conserved functions over the course of vertebrate evolution.

Zebrafish and *X. tropicalis* γ cluster isoforms do not form monophyletic groups, which suggests that the duplication events that led to multiple γ clusters in these two species were independent.

Phylogenetic relationships of mouse and *X. tropicalis* cPcdhs

The analysis of the ORFs encoded by the different *X. tropicalis* Pcdh clusters (utilizing genomic as well as long-read cDNA sequences), revealed that the overall protein domain architecture of *X. tropicalis* is identical to the domain architecture of mammalian α - and γ -Pcdh proteins. All cPcdhs of *X. tropicalis* (as mapped and indicated in Figure 1) contain six extracellular cadherin-repeat domains (EC1–EC6). Assessing the conservation pattern of individual domains based on mouse and *X. tropicalis* amino acid sequences reveals, on average, a higher conservation of domains EC1 and EC5 compared to domains EC2–EC4. This conservation pattern is similar between mouse and *X. tropicalis* (Table 1, Figure 3, Figure S4, and Figure S5). In addition, a direct *trans*-species comparison of a few representative mouse and *X. tropicalis* isoforms (Figure S3, Figure S4, Figure S5, and File S1 quantification in Table 1) indicate good conservation of domains overall, and supports the notion that the *X. tropicalis* cPcdh proteins most likely retain their canonical Pcdh properties (*e.g.*, homophilic binding properties).

On average, the similarity of EC domains between mouse and *X. tropicalis* is moderately lower for the γ 2-Pcdh cluster as compared to the α -Pcdh and γ 1-Pcdh clusters, suggesting a higher degree of divergence for γ 2-Pcdh (Table 1). Importantly, the high similarity of the constant cytoplasmic domains (68–75%; Table 1) suggests a high degree of conservation in cytoplasmic effector interactions and biological functions of mouse and *X. tropicalis* cPcdh.

In order to characterize the individual paralogous genes/isoforms of the *X. tropicalis* Pcdh clusters, we generated a phylogenetic tree using the multiple sequence alignments of the amino acid sequences of the second and third extracellular Cadherin repeat region (*i.e.*, EC2 plus EC3 domains “EC2-3”) of each of the mouse and *X. tropicalis* cPcdh

isoforms. We focused our comparison on EC2-3 as these two domains were shown to account for the specificity of the homophilic *trans*-interactions of cPcdh (Schreiner and Weiner 2010; Thu *et al.* 2014; Rubinstein *et al.* 2015; Nicoludis *et al.* 2015), and are, therefore, thought to contain key sequences responsible for the functional specificity of a given isoform. Moreover, EC2-3 appears to be prone to a lower frequency of gene conversion events compared to the C-terminal ectodomains EC4-6, and hence carry most of the phylogenetic signal (Noonan *et al.* 2004b).

The phylogenetic tree suggests that no clear orthology relation between the individual genes of mouse and *X. tropicalis* Pcdh clusters is present, and that mouse and *X. tropicalis* Pcdh genes are orthologous only as paralog subgroups. One exception to that is the mouse Pcdhac2 and *X. tropicalis* pcdh- α 14 pair, for which a clear one-to-one orthology can be inferred from the phylogenetic tree. Both isoforms are encoded by the most distal variable exons of their respective α clusters. Mouse Pcdhac2 was reported to be a C-type cPcdh isoform, which exhibits significant differences in expression compared to A/B-type cPcdhs. Therefore, the relatively shorter evolutionary distance between mouse Pcdhac2 and *X. tropicalis* pcdh- α 14 further supports the notion of important and specialized functions of these C-type isoforms. In addition, *X. tropicalis* pcdh- α 13 is evolutionarily closer to the C-type Pcdh isoforms than to the pcdh- α 1–12 isoforms of its parent α cluster. This analysis suggests that *X. tropicalis* pcdh- α 13 and pcdh- α 14 might belong to the distinct C-type cPcdhs, analogous to their mammalian counterparts.

None of the *X. tropicalis* γ -Pcdh showed a clear one-to-one orthology with mouse γ -Pcdh (Figure 3). However, 23 of the 36 isoforms of the pcdh- γ 2 cluster (pcdh- γ 2 14–36) are evolutionarily closer to the C-type α - and γ -Pcdh of mouse, and to *X. tropicalis* pcdh- α 13, pcdh- α 14 than their sister isoforms of the pcdh- γ 2 cluster or the γ -Pcdhs of the *X. tropicalis* pcdh- γ 1 cluster. This pcdh- γ 2 14–36 subgroup constitutes the distal part of the pcdh- γ 2 cluster and might be involved in distinct functions comparable to that of the mouse C-type cPcdh isoforms. This also suggests the possibility that there is a significant expansion of the C-type cPcdhs in *X. tropicalis*. In addition to the phylogeny inferred from EC2-3 domain comparisons, we generated a phylogenetic tree with full length sequences of mouse and *X. tropicalis* cPcdh (Figure S2). The topologies of both phylogenetic trees are similar. This observation is consistent with the relatively lower divergence of *X. tropicalis* EC2-3 regions (Table 1), as well as with the lower levels of gene conversion events in *X. tropicalis*.

X. tropicalis cPcdhs are prone to low level gene conversion events

It was shown that cPcdhs of mouse, human, rat, zebrafish, fugu, coelacanth, elephant shark, and lizard are subject to gene conversion although to different extents (Jiang *et al.* 2009; Noonan *et al.* 2004a,

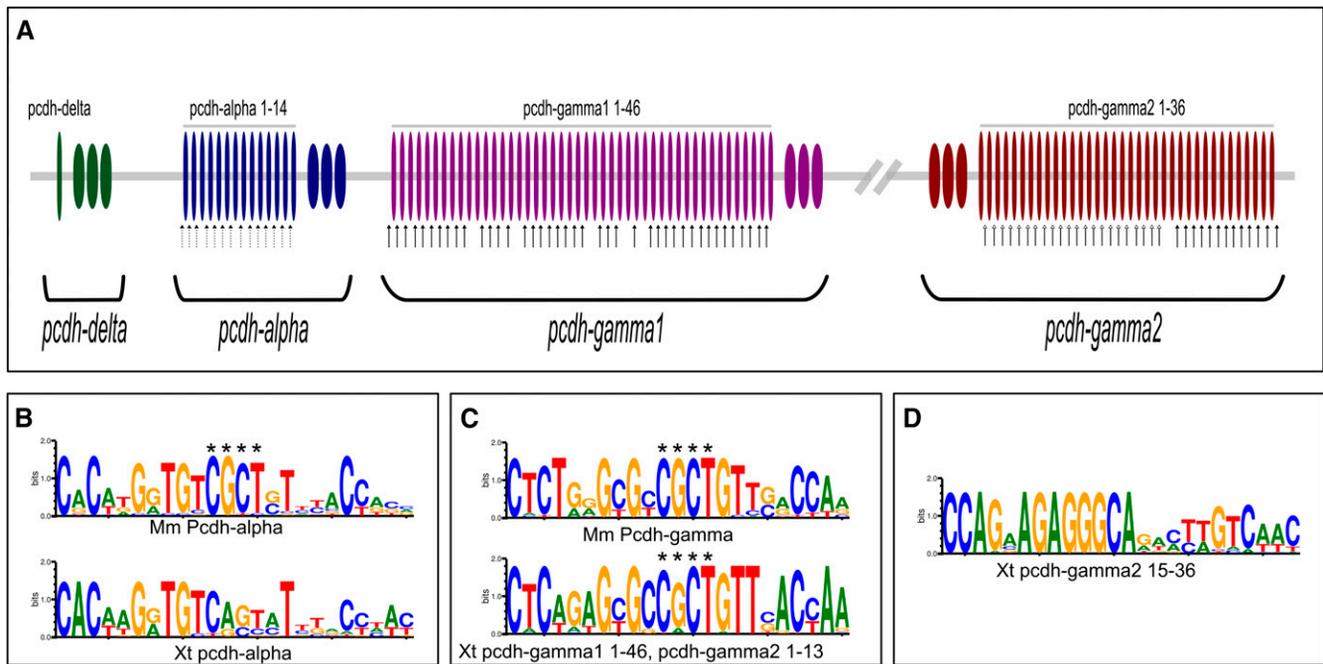


Figure 4 (A) Schematic representation of the conserved sequence motifs of *X. tropicalis* cPcdh promoters. Dotted arrows, filled arrows, and open arrows represent the genomic position of conserved motifs of (left-right) pcdh- α , pcdh- γ 1 and pcdh- γ 2. (B) CSE of mouse α -Pcdhs and *X. tropicalis* pcdh- α . (C) CSE of mouse γ -Pcdhs and *X. tropicalis* pcdh- γ 1 and pcdh- γ 2 1-13. (D) The newly identified sequence motif present in the promoters of pcdh- γ 2 15-36. The asterisks indicate the "CGCT box."

2004b; Yu *et al.* 2008, 2007). To determine the frequency of gene conversion events acting on the *X. tropicalis* cPcdhs, we calculated the average of "total number of synonymous substitutions per codon" (dS) for each of the major monophyletic groups observed in the above-mentioned phylogenetic analyses (Figure 3). dS is used as an indication of gene conversion since synonymous sites are not affected by functional constraints at the protein level; therefore, a higher number of synonymous substitutions reflects a lower rate of gene conversion events. Table 2 lists the average dS values for each of the six ectodomains of the *X. tropicalis* and mouse cPcdhs as well as the average dS values for a selected group of zebrafish cPcdhs (*Pcdh2ab1-12*), which were shown to be subject to a high frequency of gene conversion events (Noonan *et al.* 2004b). The last column of Table 2 lists the ratio of the highest and the lowest dS values, and is reflective of overall gene conversion events exercising over the entire length of EC1-6. In general, the highest dS values listed in Table 2 were observed in EC2 or EC3 domains, while the lowest dS values were observed in EC5 or EC6 domains (Table 2), which is concordant with previous findings. The ratios of the highest dS and lowest dS for each of the monophyletic groups are comparable for *X. tropicalis* and mouse, but significantly lower than zebrafish. These findings show that *X. tropicalis* cPcdhs experienced similar levels of gene conversion events as mouse cPcdhs, but significantly lower levels of gene conversion events compared to zebrafish cPcdhs.

Core promoter elements of the *X. tropicalis* cPcdhs

Promoters of cPcdhs of various species including human, mouse, and zebrafish carry a core element referred to as the CSE. In mouse, the CSE is positioned ~200 bp upstream of the translational start codon (Wu *et al.* 2001; Noonan *et al.* 2004b), and is composed of a "CGCT box," which is surrounded by additional conserved cluster-specific sequences. However, zebrafish α -Pcdhs have divergent CSEs that lack the CGCT box (Noonan *et al.* 2004b).

Our analysis revealed that many of the *X. tropicalis* cPcdh transcription units also contain the CSE signature sequences found in mammals in their promoters (Figure 4). In addition, we identified a novel distinct conserved promoter sequence motif, which may constitute an additional CSE-like element; however, this remains to be validated experimentally.

We scanned the promoters (2 kb upstream of the translational start codon) of *X. tropicalis* cPcdhs, and showed that the CSE is present in most *X. tropicalis* cPcdh promoters, and, similar to the mammalian CSE elements, they are located some 200 bp upstream of the translational start codon (Figure 4A).

All of the 14 α -Pcdhs of *X. tropicalis* carry the CSE motif; however, the CGCT box, which is conserved in mammalian α -Pcdh promoters, is not present in *X. tropicalis* α -Pcdh promoters (Figure 4B). In contrast, the *X. tropicalis* pcdh- γ 1 1-46 and pcdh- γ 2 1-13 (except for pcdh- γ 1 isoforms 11, 16, 25, and 29) carry the CSE motif, in which the CGCT box is conserved, although to a lesser extent than in mammals (Figure 4C).

Isoforms that lack the CSE motif in their promoters (pcdh- γ 2 15-36), carry a 13 bp conserved sequence ~200 bp upstream of their translational start codon (Figure 4D). This motif has previously not been reported in any other the species, and is not listed in the databases of transcription factor binding sites. Interestingly, this motif is present in a contiguous group of isoforms of the γ 2 cluster (pcdh- γ 2 15-36), which form a monophyletic group with mouse C-type cPcdh isoforms *Pcdhgc4* and *Pcdhgc5* (Figure 3).

Expression patterns of *X. tropicalis* cPcdhs

In order to further characterize *X. tropicalis* cPcdhs, we investigated their temporal and spatial expression patterns by reanalyzing publicly available RNA-seq data and by RNA *in situ* hybridization.

Tan *et al.* (2013) studied the *X. tropicalis* transcriptome in 23 distinct developmental stages (between the two-cell stage and stage 45) by paired-end RNA sequencing (RNA-seq). For their analyses, they used

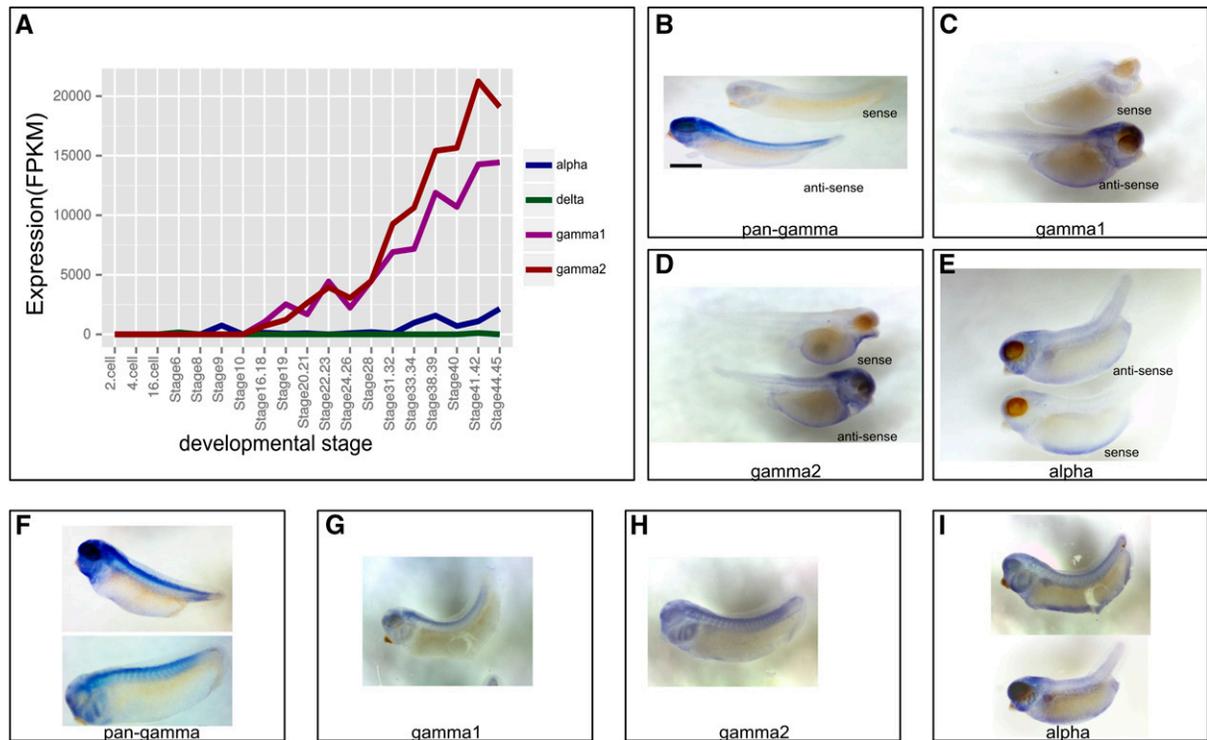


Figure 5 Expression of *X. tropicalis* cPcdhs. (A) *X. tropicalis* cPcdh expression over 19 different developmental stages in FPKM units (reanalyzed from Tan *et al.* 2013). (B–E) *X. tropicalis* cPcdh RNA *in situ* hybridization sense and antisense probes. (F–I) Spatial expression patterns of *X. tropicalis* cPcdhs as detected by RNA *in situ* hybridization. (B–I) mRNA *in situ* hybridization results of the *X. tropicalis* cPcdhs. The *pan-gamma* riboprobe targets the constant exons of *pcdh-γ1*, and most likely recognizes the *pcdh-γ2* cluster constant exons as well because of high sequence similarity of the constant exons of the two γ clusters (73.89% sequence identity). The *gamma1* and *gamma2* riboprobes recognize the 3'UTR regions of *pcdh-γ1* and *pcdh-γ2* clusters, respectively, and the *alpha* riboprobe recognizes the constant exons of the *pcdh-α* cluster. (B–E) Sense and antisense probes. (F–I) Additional images from the antisense probes.

Genome Build 4 (xenTro3), and gene models associated with this genome build.

This study provided important information about the *X. tropicalis* transcriptome in general; however, due to lack of complete gene models of cPcdhs in Genome Build 4, their expression could not be retrieved directly. Therefore we reanalyzed the raw sequencing data using Genome Build 7 (xenTro7), and our newly created cPcdh gene models. We determined “cluster-level” expression by using or read-pairs that map to the constant exons that are shared by all the genes of a given cluster. The expression levels were reported as “fragments per kilobase of exon per million fragments mapped” (FPKM), which is analogous to the “reads per kilobase of exon per million reads mapped” (RPKM) used in the original study (Tan *et al.* 2013). RPKM (and FPKM) allows transcript level comparisons within, as well as between, samples (Dillies *et al.* 2013).

The α - and δ -Pcdhs are expressed in low-levels up to stage 45, however α -Pcdh expression levels start to increase at stages 31–32 (Figure 5A). Conversely, the two γ -Pcdhs start to be expressed after stage 12, which marks the end of the gastrula stage and the beginning of the neurula stage. RNA *in situ* hybridization experiments (Figure 5, B–I) show that *X. tropicalis* cPcdh genes from all three clusters are expressed throughout the nervous system. α - as well as γ -Pcdh mRNA is already present in the early neuroectoderm region of embryos (data not shown), and their expression increases through subsequent stages of neural development (Figure 5) and persists into metamorphic stages (data not shown). This includes expression in the eye anlagen, retina, otic vesicle, olfactory bulb, all brain regions, spinal cord, and, at least in

part, in neural crest lineages (e.g., expression in pharyngeal arches) (Figure 5). Based on whole-mount *in situ* staining, it is clear that α - and γ -Pcdh are broadly expressed in the nervous system and exhibit substantial overlap in expression. However, whether α - and γ -Pcdh are indeed also coexpressed at the single cell level, and in most or all neurons, awaits further analysis.

DISCUSSION

In our efforts to characterize the *X. tropicalis* cPcdh receptors, we identified a single chromosomal locus containing three Pcdh clusters that we designated *pcdh-α*, *pcdh-γ1* and *pcdh-γ2*. The *pcdh-α* cluster contains 14, the *pcdh-γ1* cluster contains 46, and the *pcdh-γ2* cluster 36 variable exons encoding for a total of 96 Pcdh isoforms. The organization of three tandem arrays of Pcdh isoforms at a single chromosomal location is analogous to the gene organization in mammals and coelacanth (Wu *et al.* 2001; Noonan *et al.* 2004a). While *X. laevis* and zebrafish have multiple cPcdh loci due to whole-genome duplications, we found no evidence of additional Pcdh loci in *X. tropicalis*, which should facilitate future genetic analysis of cPcdh function in this vertebrate model organism.

In contrast to the mammalian cPcdh locus, the *X. tropicalis* genome lacks Pcdh- β receptors, yet contains a second distinct *pcdh-γ* cluster. It seems most likely that the two γ clusters are derived from lineage-specific gene duplications in *Xenopus*; however, a gene loss in other organisms can currently not be excluded based on available sequence information. Pcdh- β receptors are present in mammals (22 isoforms in mouse and 15 in humans), but also in coelacanth, lizard, and chicken

genomes. However, little is known about their potential functional specialization and significance, as genetic studies in mice have focused primarily on characterizing the importance of α or γ receptors. Nevertheless, the striking increase in *pcdh- γ* isoforms in *X. tropicalis* might suggest some degree of functional interchangeability between *pcdh- γ* and *pcdh- β* receptors.

It has recently been proposed that cPcdhs emerged in jawed vertebrates, in which a common ancestor comprised numerous different cPcdh types (α , β , γ , δ , ϵ , μ , ν , etc.) (Yu *et al.* 2008). Of those, the α -, β -, γ - and δ -types were retained in bony vertebrates. The fact that coelacanth (nonteleost fish) have a β -Pcdh cluster, while representatives of teleost fishes (zebrafish and fugu) were found to lack β -Pcdhs, suggests that β -Pcdhs were lost in the teleost fish lineage but not all lower vertebrates. In summary, the loss of a β -Pcdh cluster in the *Xenopus* lineage on top of an expanded γ -Pcdh diversification in *X. tropicalis* provides further evidence for an exceptionally dynamic evolution of Pcdh gene clusters.

In a broader context, the high degree of variability of the numbers of clusters, subtypes and isoforms of cPcdh receptors across species might support the notion that evolutionary selection primarily drives the generation of receptor diversity rather than the conservation of unique structural specificities of distinct cPcdh genes. This notion would be consistent with the hypothesis that vertebrate cPcdh receptors—analogue to the hypervariable Dscam1 receptor in flies—function primarily in providing a highly diverse pool of cell adhesion receptors with homophilic binding preferences. Studies in flies have shown that a significant amount of Dscam1 diversity is essential for providing unique surface labels for neurons to support a system of “neuronal self-recognition.” Furthermore, genetic manipulations have shown that, as long as the overall repertoire of receptor isoforms encoded by the Dscam1 gene is above a certain threshold (e.g., a few thousand in flies), such that it can ensure that interacting neurons express multiple nonidentical sets of isoforms, the number and binding specificities of isoforms can vary substantially (Hattori *et al.* 2009). Not surprisingly, the overall number of Dscam1 isoforms in different insect species can vary substantially (Brites and Pasquier 2015).

Our phylogenetic comparisons of cPcdh sequences indicate similar levels of gene conversion among cPcdh isoforms in *X. tropicalis* and mouse, yet significantly lower levels of gene conversions than zebrafish. More importantly, we found striking differences of gene conversion rates when different extracellular cPcdh domains were compared. Specifically, the EC2 and EC3 domains appear not to be subject of gene conversion, in contrast to other extracellular domains of cPcdhs. This is particularly interesting as recent *in vitro* and structural studies suggested that the EC2 and EC3 domains provide essential protein interfaces determining homophilic binding specificity in cPcdh dimers. These data, therefore, corroborate a positive selection of sequences in EC2/EC3 domains that are optimized for unique homophilic binding specificities.

In our comparative analysis, we also found a remarkably high degree of conservation of noncoding sequence motifs serving regulatory functions of cPcdh expression. In fact, many of the *X. tropicalis* cPcdhs variable exon units contain CSE signature sequences characteristic of related promoters in mammalian cPcdhs. This strongly suggests that transcriptional control factors and mechanisms (including chromatin regulation) are highly conserved between frog and mammalian cPcdhs. It is intriguing to note, however, that a distinct set of 22 tandemly arrayed isoforms (*pcdh- γ 2 15–36*) lacks a CSE motif in their promoters, and yet all contain a similarly positioned currently uncharacterized sequence motif. This suggests the presence of a novel regulatory factor, which may support or drive *X. tropicalis* specific specializations in cPcdh expression and function.

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