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Swelling and Mechanical Properties of Alginate Hydrogels with Respect to Promotion of Neural Growth

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Soft alginate hydrogels support robust neurite outgrowth, but their rapid disintegration in solutions of high ionic strength restricts them from long-term in vivo applications. Aiming to enhance the mechanical stability of soft alginate hydrogels, we investigated how changes in pH and ionic strength during gelation influence the swelling, stiffness, and disintegration of a three-dimensional (3D) alginate matrix and its ability to support neurite outgrowth. Hydrogels were generated from dry alginate layers through ionic crosslinks with Ca2+ (≤10 mM) in solutions of low or high ionic strength and at pH 5.5 or 7.4. High- and low-viscosity alginates with different molecular compositions demonstrated pH and ionic strength-independent increases in hydrogel volume with decreases in Ca2+ concentrations from 10 to 2 mM. Only soft hydrogels that were synthesized in the presence of 150 mM of NaCl (Ca-alginateNaCl) displayed long-term volume stability in buffered physiological saline, whereas analogous hydrogels generated in NaCl-free conditions (Ca-alginate) collapsed. The stiffnesses of Ca-alginateNaCl hydrogels elevated from 0.01 to 19 kPa as the Ca2+ concentration was raised from 2 to 10 mM; however, only Ca-alginateNaCl hydrogels with an elastic modulus ≤1.5 kPa that were generated with ≤4 mM of Ca2+ supported robust neurite outgrowth in primary neuronal cultures. In conclusion, soft Ca-alginateNaCl hydrogels combine mechanical stability in solutions of high ionic strength with the ability to support neural growth and could be useful as 3D implants for neural regeneration in vivo.

Introduction

Biocompatible three-dimensional (3D) scaffolds for neural tissue engineering must provide an adhesive matrix for regenerating cells, be mechanically compatible with elastic neural tissues, and display dimensional stability in the presence of physiological fluids because extensive swelling or shrinking will affect scaffold-tissue unity or compress the surrounding healthy tissues.1,2 Hydrogels are 3D networks of interconnected polymer fibers wherein entrapped liquid occupies most of the volume. Hydrogels are permeable to nutrients, support the viability of encapsulated cells, and can incorporate growth factors or other polymers. A variety of natural and synthetic biomaterials form hydrogels.3,4 Alginate is a biocompatible, natural, linear, binary copolymer composed of D-mannuronic acid (M) and its C5 epimer L-guluronic acid (G) monosaccharide units, which are covalently linked by β-1-4 glycosidic bonds. M and G are distributed within the polymer chain in varying proportions to produce heterogeneous alternating (MG) and homogeneous (MM or GG) sequences in the primary structure.5–9 In aqueous solutions (sols), alginate exists as a negatively charged polyanion that forms a hydrogel by the physical association of polysaccharide chains through ionic crosslinking of G residues by multivalent cations (e.g., Ca2+, Sr2+, Ba2+, Zn2+, Cu2+, Ni2+, or Fe3+). This process has been described by the egg-box model in which divalent ions interact with two adjacent G residues and with two G residues of an opposing chain.10,11 The density of the alginate fiber network within a hydrogel and its gel strength are regulated by the number of connected gelling sites; as a result, these properties are elevated with an increasing level of crosslinking ion saturation in the alginate and reduced with an increasing number of free G blocks.6,12–14 The behavior of charged polyelectrolytes, which includes alginates, is strongly regulated by the pH and ionic strength of the surrounding solution; pH controls the degree of dissociation of the guluronic and mannuronic acid groups, while ionic strength triggers the interaction of the resulting negative charges along the polymer chains.15,16 Most polyelectrolyte-based gels disintegrate upon immersion in solutions of high ionic strength. Ionically crosslinked alginate hydrogels undergo disintegration in the presence of calcium chelators (e.g., phosphates), monovalent ions (e.g., Na+), and

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non-cross-linking divalent ions (e.g., Mg\(^{2+}\)), which are present in the tissue culture medium and tissue fluids.\textsuperscript{17–19} Disintegration of polyelectrolyte-based hydrogels strongly limit their usefulness for \textit{in vivo} applications, which require long-term 3D stability of the implant. Traditionally, alginate gels were fabricated in NaCl-free solutions; recent studies on gelation speed, swelling, and degradation and on the critical steps in Ca\(^{2+}\) binding to alginates were also performed on hydrogels fabricated in NaCl-free solutions.\textsuperscript{20–23} Conventional methods to stabilize alginate hydrogels include covalent crosslinking and application of high concentrations of crosslinking cations to ensure tight connections between the polymer chains.\textsuperscript{24–26}

Physical cues such as substrate stiffness are important factors for cell differentiation and behavior.\textsuperscript{27,28} The successful use of hydrogels greatly relies on the refined control of the mechanical properties, including stiffness. Neurons can sense the stiffness of a substrate and will behave accordingly. For example, soft but not stiff alginate hydrogels support robust neurite outgrowth \textit{in vitro}.\textsuperscript{29} Dorsal root ganglion neurons display maximal growth on soft substrates with stiffnesses \(<\) 1 kPa.\textsuperscript{30,31} Neurons plated on softer substrates showed decreased branching relative to neurons grown on stiffer substrates.\textsuperscript{32,33} Mesenchymal stem cells and primary neural stem cells differentiate into neurons on soft hydrogels with stiffnesses \(<\) 1 kPa.\textsuperscript{34–37} Stiff alginate hydrogels were demonstrated to be nonadhesive for neurons; to provide an adhesive surface, these hydrogels required functionalizations with signaling molecules such as laminin and fibronectin\textsuperscript{38–40} and integration of polyglycolic acid, heparin, or basic fibroblast growth factor (bFGF).\textsuperscript{41–44} Anisotropic, stiff alginate scaffolds, which were covered with collagen or polylysine or incorporated gelatin, were shown to support neurite elongation along capillary channels, but prohibited neuronal ingrowth into the hydrogel core.\textsuperscript{45,46}

Soft alginate hydrogels support neural growth, but nothing is yet known about how to enhance their mechanical stability in solutions of high ionic strength without elevation of their stiffness and affecting their interaction with neural cells. This study emphasizes that gelation of alginate layers with substoichiometric concentrations of calcium ions in the presence of NaCl generates soft Ca-alginates\textsubscript{NaCl} hydrogels, which combine mechanical stability with high adhesion to neural cells. This translational research opens a new perspective on the design of stable 3D polyelectrolyte-based hydrogels for long-term \textit{in vivo} applications.

Materials and Methods

Materials and reagents

Ultrapure sodium alginates with trade name PRONOVA were purchased from Novamatrix, now part of FMC Bio-Polymer, as guluronic acid rich alginate (LVG) and mannuronic acid rich alginate (LVM), and endogen-free S-alginate were purchased from Sigma (71238; Taufkirchen, Germany). The molecular compositions of LVG and LVM were provided by the supplier, but the composition of S-alginate was quantified for this study by a proton \(^1\text{H}-\text{NMR}\) spectroscopy method.\textsuperscript{47} LVM, LVG, and S-alginates contained 43%, 68%, and 68% guluronic acid residues, respectively. Reagents, including the neurobasal medium, DMEM/F12 (1:1), B-27 supplement, Dulbecco’s Phosphate-Buffered Saline without Ca\(^{2+}\) and Mg\(^{2+}\), and trypsin/EDTA solution were all purchased from Invitrogen (Darmstadt, Germany). CaCl\(_2\) dihydrate, HEPES, TRIS, NaCl, and DNase I originated from Sigma.

Preparation of alginate hydrogels at ambient conditions

Alginate gels were dissolved in deionized water under permanent stirring at room temperature (RT) for 12 h. The resulting aqueous 0.5%, 1.0%, or 1.5% sols were sterilized by pressurizing through a 0.45-\(\mu\)m sterile filter. Dry alginate layers were formed on glass coverslips (24 x 24 mm) when each sol was evenly distributed onto a glass surface and dried at RT for 24 h. Dry alginate layers were stored under humidity-free conditions and used within 10 days after preparation. To form a hydrogel, a glass coverslip with a dry alginate layer was placed into a Petri dish (Ø 5 cm) and overlayed with a 15 mL crosslinking solution; the gelation reaction was carried out at RT for 24 h, and a single interconnected gel slice was formed on the glass surface. The excess crosslinking solution was removed before gravimetric and rheological analyses. Representative images of hydrogels were taken with a digital camera. For the cell culture experiments, hydrogels were prepared under sterile conditions in a laminar flow hood as described previously.\textsuperscript{26} For rheological characterization, hydrogels were prepared in plastic Petri dishes (Ø 5 cm) from dry alginate layers derived from 1% alginate sol, and the gelation reaction was carried out at RT for 24 h.

Composition of crosslinking solutions

Alginate hydrogels were prepared through ionic crosslinking, and an aqueous calcium chloride solution was used as a source of Ca\(^{2+}\) cations. The concentration of CaCl\(_2\) in the crosslinking solutions ranged from 0 to 10 mM. The pH values of the crosslinking solutions were adjusted to either 5.5 or 7.4 with the HEPES buffer solution. Some of the crosslinking solutions contained NaCl (150 mM, final). Our abbreviation scheme describes the characteristics of the hydrogel’s ionic composition in the crosslinking solution; for example, Ca-alginate and Ca-alginates\textsubscript{NaCl} hydrogels were generated with CaCl\(_2\) dissolved either in water or in 150 mM NaCl, respectively.

Measurements of alginate hydrogel swelling in gravimetric assay

The weight of each hydrogel (g) was measured on an analytical balance (Sartorius, Göttingen, Germany; 0.1 mg precision). To preserve gel integrity, the cumulative mass of a glass–gel sandwich was measured and the average weight of the glass coverslips (0.22 g) was subtracted to obtain the hydrogel’s mass. At least three independent samples were prepared for every experimental condition. The Student’s \(t\)-test for unpaired observations was applied to make a statistical analysis of the data. The rate of swelling was estimated after comparison of the hydrogel mass with the mass of the 1% alginate sol (taken as 100%).

Rheological characterization of alginate sols and hydrogels

Rheological measurements were carried out using an Anton-Paar Physica MCR301 (Graz, Austria) instrument. The
viscosity values of aqueous alginate solutions were recorded under continuous logarithmically increasing shear in a double slit chamber DG26.7 at 20°C. A stress sweep at a constant frequency of 1 Hz was performed to obtain values for the elastic modulus (G’) of alginate hydrogels. Viscosity values were recorded with the help of a measurement cell with parallel plate geometry (PP25) under logarithmically increasing amplitude (0.001–100 Pa) at 37°C. Each hydrogel was transferred onto the measuring device and cut into the shape of a disk (Ø 25 mm). Samples were equilibrated at 37°C for 5 min before measurements.

Culturing of primary neuronal cells

Neuronal cell cultures were prepared from E20 Wistar rat fetuses. Cells were isolated from cortices as described.48 As we reported previously, cleavage of the cell surface molecules during acute brain tissue dissociation using serine protease trypsin alters initiation of neural cell adhesion and growth on soft alginate hydrogels.29 Therefore, in the present study, we used primary cell spheroids, which were completely recovered from treatment with trypsin. Primary cells were seeded at an initial cell density of 1 × 10⁶ cells/well in uncovered six-well plates with 2 mL of expansion medium consisting of DMEM/F-12 (1:1) supplemented with 2 mM L-glutamine, 15 mM HEPES, 20 ng/mL of rhEGF (Sigma), and 10 ng/mL of rh-bFGF (Sigma). Subsequently, multiple spheroids formed after reaggregation or/and proliferation of cells on day 3 in vitro. Spheroids were collected, replated onto hydrogel samples, and cultured in a differentiating medium to potentiate differentiation toward neural cells. The differentiating medium consisted of 2% B-27 supplemented neurobasal medium, 0.5 mM L-glutamine, and 1% ampicillin and 1% streptomycin solution. Cell incubation was carried out at 37°C in a humidified 95% air and 5% CO₂ atmosphere.

Immunocytochemistry

The primary monoclonal mouse antibodies were anti-β-tubulin III, anti-GFAP, and anti-Tau-1 (from Chemicon, Schwalbach, Germany) and anti-nestin (R&D Systems, Wiesbaden-Nordenstadt Germany), and secondary antibodies were Alexa Fluor-488 chicken anti-mouse (Molecular Probes/Invitrogen). Labeling with antibodies was performed according to a standard procedure; Hoechst 33342 (10 μg/mL in TBS) was used to label the nuclei. Samples were examined under a fluorescence microscope (Axiovert 40; Zeiss, Jena, Germany) and confocal microscope (LSM-710; Zeiss).

Neurite-bearing spheroids, neurite length, and statistical analysis

Spheroids that were formed from 1 × 10⁶ primary cortical cells were collected into a sterile tube containing a fresh portion (6 mL) of a differentiating medium, then mixed, and evenly distributed on top of three identical Ca-alginateNaCl hydrogels (e.g., Ca-alginateCaCl₂ hydrogels derived from S-alginate crosslinked with a solution containing 2 mM of CaCl₂, 150 mM of NaCl, and at pH 7.4) and cultured subsequently for 48 h. The diameter of spheroids ranged from 50 to 250 μm by day 5 in vitro. Spheroids that extended 10 or more neurite from at least a quarter of its surface contacting the hydrogel were defined as neurite-bearing spheroids; of note, the vast majority of neurite-bearing spheroids extend 50 or more neurite. Single cells as well as aggregates of cells (2–10 cells) were not considered during spheroid counting. The proportion of neurite-bearing spheroids was quantified after the examination of 50 random live spheroids cultured on top of three identical Ca-alginateNaCl hydrogels. Different types of hydrogels were derived either from S-alginate, LVG, or LVM alginites crosslinked at pH 5.5 or 7.4 with a solution containing different concentrations of aqueous CaCl₂ (i.e., 2, 4, 6, 8, or 10 mM) and 150 mM of NaCl as described above. In each independent cell culture experiment, 30 different hydrogel types were generated and tested for their ability to support neurite outgrowth. Every hydrogel type was generated in triplicate; subsequently, in each independent cell culture experiment, spheroids were examined on 90 individual hydrogels. The final proportion of neurite-bearing spheroids on every hydrogel type was estimated from two independent cell culture experiments after the examination of 100 (in total) individual spheroids on six identical hydrogels. The distance of neurite extension from spheroids was quantified in two independent cell culture experiments 48 h after plating of spheroids on top of different types of soft Ca-alginateNaCl hydrogels generated using S-alginate, LVG, and LVM with 2 mM of aqueous CaCl₂ at pH 7.4 as described above. Subsequently, 20 random spheroids were examined per one type of a hydrogel and the length of 20–50 random neurite per spheroid was quantified on live phase-contrast images (Zeiss Ph2 Plan-NEOFLUAR 20X/0.5 objective, Axiovert 40; Zeiss) using NeuronJ (www.imagescience.org/meijering/software/neuronj). The Student’s t-test for unpaired observations was applied for statistical analysis. Differences between samples with p-value<0.05 were accepted as statistically significant.

Results

Swelling of Ca-alginate hydrogels in aqueous CaCl₂ solution

Swelling is a result of liquid absorption by the polymer, and could be estimated by measuring the hydrogel mass. Dry alginate polysaccharide chains swell in water to produce a sol, but they can swell in the presence of crosslinking di- or multivalent cations, which lead to a hydrogel. Crosslinking of surface-attached dry alginate layers by a 2 mM aqueous CaCl₂ solution resulted in the interconnection of polysaccharide molecules and the formation of a single gel slice (soft hydrogel), which support robust neurite outgrowth in vitro; in contrast, gelation of the alginate sol in an aqueous solution under gentle stirring generated multiple gel fragments of irregular size. The appearance of the soft hydrogels (e.g., fragments or slices) did not alter their high adhesiveness to neurite. These results matched our previous data29 and demonstrated that dry alginate layers could efficiently form soft hydrogels with the ability to support robust neural growth.

While cultivating neurons on top of soft hydrogels, we often observed variability in the long-term hydrogel stability: some hydrogels disintegrated in the presence of a cell culture medium during the first 3 days, but others persisted for up to 10 weeks and longer. In this study, we generated hydrogels from surface-attached dry layers of different
alginate sol (1%) and (b) the concentration of an alginate in a sol at a constant sol volume of 600 μL. Hydrogels were generated from dry layers of S-alginate by crosslinking with 2 mM of CaCl₂ at pH 5.5 in the absence (Ca-alginate) or in the presence of 150 mM of NaCl (Ca-alginateNaCl). Data are represented as means±standard deviations.

**FIG. 1.** Hydrogel volume increases with an increase in (a) the volume of alginate sol at constant alginate concentrations in a sol (1%) and (b) the concentration of an alginate in a sol at a constant sol volume of 600 μL. Hydrogels were generated from dry layers of S-alginate by crosslinking with 2 mM of CaCl₂ at pH 5.5 in the absence (Ca-alginate) or in the presence of 150 mM of NaCl (Ca-alginateNaCl). Data are represented as means±standard deviations.

**Swelling of Ca-alginateNaCl hydrogels in aqueous CaCl₂ solution**

Like Ca-alginate hydrogels, hydrogels generated in the presence of NaCl (150 mM, Ca-alginateNaCl) displayed nonlinear swelling profiles in solutions with pH values of 5.5 (not shown) and 7.4, and a decrease in the CaCl₂ concentration to below 4 mM was accompanied by a rapid increase in hydrogel mass (Fig. 3). At a constant concentration of CaCl₂, the Ca-alginateNaCl hydrogel’s mass was proportional to the sol volume and concentration of alginate in the sol (Fig. 1a, b). Na⁺ ions do not have any specific binding sites within the alginate, but interact electrostatically with the negatively charged carboxylic groups of the polysaccharide. We observed that at a low degree of crosslinking (≤4 mM of CaCl₂), exogenous Na⁺ ensures the generation of less swollen hydrogels (Fig. 3) and shifts the minimum gel-forming concentration of Ca²⁺ ions toward higher Ca²⁺ concentrations for G-enriched alginates (1.4 and 1.6 mM of CaCl₂ in S-alginate and LVG, respectively). Thus, when a high Na⁺/Ca²⁺ ratio exists, Na⁺ could inhibit some G-enriched sequences in the primary alginate structure from being crosslinked by Ca²⁺, but does not alter the fundamental principal of hydrogel formation with Ca²⁺ (CaCl₂≤10 mM). Moreover, as we demonstrated below, only strongly swollen Ca-alginateNaCl hydrogels with the degree of swelling above 66% supported robust neurite outgrowth in vitro.

**Mechanical stability of Ca-alginateNaCl and instability of Ca-alginate hydrogels in buffered physiological saline**

Sodium is a constituting ion of every extracellular medium in the body, and it contributes to the high ionic strength of extracellular fluids, including blood plasma and cerebrospinal fluid. Moreover, the pH value in normal blood and cerebrospinal fluid is tightly regulated between 7.35 and 7.45. To predict how long different in situ prepared hydrogels can maintain their dimensions in the presence of these physiological fluids in vivo (e.g., as implants into injured spinal cord) and subsequently support neural growth and regeneration, we quantified hydrogel disintegration in a buffered physiological saline. Ca-alginate and Ca-alginateNaCl hydrogels generated in the presence of 2 mM of CaCl₂ were immersed in buffered physiological saline (150 mM of NaCl, pH 7.4) supplemented with 2 mM of CaCl₂.
and the hydrogel mass was regularly quantified during the subsequent 30 days. Ca-alginate hydrogels derived from LVM completely collapsed, while those derived from LVG and S-alginate lost at least 50% of their mass within the first 30 days (Fig. 4a). Surprisingly, analogous Ca-alginate$_{\text{NaCl}}$ hydrogels demonstrated long-term mass stability during the entire observation period (Fig. 4b).

**Macroscopic morphology of hydrogels**

Rapid binding of calcium ions to alginate produces macroscopically inhomogeneous structures; conversely, slow
gelation kinetics, which commonly occur in the presence of components that compete with alginate for Ca\(^{2+}\) complexation such as oligoguluronates and EDTA, lead to more homogeneous gels.\(^{12,51}\) We noticed remarkable differences in the surface profiles between Ca-alginateNaCl and analogous Ca-alginate hydrogels. Many of the Ca-alginateNaCl hydrogels had a smooth and uniform surface, whereas Ca-alginate gels had ruffles and creases of irregular shape and pattern (Fig. 5). The differences in gel appearance were independent of alginate type and became more apparent in gels generated with solutions containing \(\geq 3\) mM of CaCl\(_2\) (Fig. 5f, g). Thus, at a high Na\(^+\)/Ca\(^{2+}\) ratio, Na\(^+\) could delay Ca\(^{2+}\)-mediated chain association, which optimizes the spatial organization of the alginate network.

Rheological characterization of Ca-alginate\(_{\text{NaCl}}\) hydrogels

We carried out oscillatory shear measurements in a rheometer and quantified the stiffness of the Ca-alginate\(_{\text{NaCl}}\) hydrogels. The stiffness of all tested hydrogels increased with higher levels of Ca\(^{2+}\) in the crosslinking solution. In addition, LVM-derived gels were softer than analogous S-alginate- and LVG-derived gels (Fig. 6). For example, the average stiffness of S-alginate- or LVM-derived Ca-alginate\(_{\text{NaCl}}\) hydrogels formed in the presence of 2, 4, 8, and 10 mM of CaCl\(_2\) were 0.011, 1.470, 3.665, and 19.230 kPa (S-alginate) and 0.025, 0.622, 3.085, and 7.356 kPa (LVM), respectively.

Neurons rapidly extend neurite on soft Ca-alginate\(_{\text{NaCl}}\) hydrogels

Neurons have an intrinsic capacity to extend neurite on surfaces they can adhere to. Previously, we described that neurons extend neurite on soft alginate hydrogels prepared with 2 mM of Ca\(^{2+}\), but gelation with 10 mM of Ca\(^{2+}\) resulted in a stiff and nonadhesive hydrogel; this change was attributed to increased tightness of the alginate network.\(^{29}\)

FIG. 4. Ca-alginate\(_{\text{NaCl}}\) hydrogels demonstrate prolonged volume stability in buffered physiological saline. Ca-alginate and Ca-alginate\(_{\text{NaCl}}\) hydrogels were generated at pH 5.5 from dry alginate layers in the presence of 2 mM of CaCl\(_2\) (a) in the absence or (b) in the presence of 150 mM of NaCl and were then incubated in buffered physiological saline (150 mM of NaCl at pH 7.4 and 2 mM of CaCl\(_2\)) for 1, 10, and 30 days. Alginate layers were derived from 600 \(\mu\)L of a 1% sol of S-alginate, LVG, and LVM. Data are represented as means \pm standard deviations.

FIG. 5. Phase-contrast images of Ca-alginate and Ca-alginate\(_{\text{NaCl}}\) hydrogels. Hydrogels were generated at pH 7.4 by crosslinking dry alginate layers of S-alginate with 2–10 mM of aqueous CaCl\(_2\) solutions (a–e) in the absence or (f–j) in the presence of 150 mM of NaCl. Alginate layers were derived from 600 \(\mu\)L of a 1% S-alginate sol. The scale bar is 1 cm.
We tested whether Ca-alginateNaCl hydrogels are still capable of promoting neural adhesion and growth under growth factor-free conditions and without serum. An array of hydrogels were generated from LVM, LVG, and S-alginate; crosslinking was performed in the presence of a constant concentration of NaCl (150 mM) and different concentrations of CaCl$_2$ (i.e., 2, 4, 6, 8, or 10 mM) at pH 5.5 or 7.4.

As we reported previously, the ability of a soft alginate hydrogel to promote neurite extension did not depend on the cell culture system because two-dimensional (2D) adherent monolayers of neurons and 3D neural spheroids produced identical results. Thus, 3D spheroids were chosen for the cell culture experiments in this study. Spheroids consisted of living cells, which neither accumulated trypan blue dye nor contained fragmented nuclei (data not shown). Three hours after being plated on soft Ca-alginateNaCl hydrogels (S-alginate, LVG, and LVM gelled with 2 mM of CaCl$_2$), multiple neurite extended in all directions and away from the center of the spheroids. The spheroid-derived neurite expressed the neural markers MAP2 and β-tubulin III (Fig. 7a, e, i and b, f, j) and developed clearly defined growth cones (Supplementary Fig. S1; Supplementary Data are available online at www.liebertpub.com/tec). Growth cone migration elongates neurite, while growth cone splitting creates a branch point resulting in two or more branches. Neurite elongation prevailed over neurite splitting on soft Ca-alginateNaCl hydrogels since vast majority of neurite (Supplementary Fig. S1a) including MAP2-labeled dendrites (Fig. 7a, e, i) did not branch. Neurite predominantly extended from the area of the spheroid that was in direct contact with the underlying hydrogel. Neurite growth was detected on the surface and in the entire hydrogel volume, gentle shaking could not detach the neurite or the neurite-bearing spheroids. Average distance of neurite extension from spheroids was measured 48 h after plating of spheroids on top of hydrogels and corresponded to 261 ± 120, 277 ± 126, and 312 ± 135 µm on S-alginate, LVG, and LVM Ca-alginateNaCl hydrogels generated in the presence of 2 mM of CaCl$_2$, respectively. The proportion of neurite-bearing spheroids on top of S-alginate, LVG, and LVM Ca-alginateNaCl hydrogels generated in the presence of 2 mM of CaCl$_2$ was 80%, 83%, and 93%, respectively, but only 18%, 21%, and 22% on analogous hydrogels prepared in the presence of 4 mM of CaCl$_2$. Gelation with ≥6 mM of CaCl$_2$ abolished the attachment of spheroids to the hydrogels and, consequently, eliminated neurite extension (Fig. 7c, d, g, h, k, l). Unlike the importance of calcium concentrations in a crosslinking solution, the pH values during alginate gelation (5.5 or 7.4) had no influence on spheroid attachment and neurite growth on Ca-alginateNaCl hydrogels during subsequent cultivation. Spheroid-derived neurite continued to elongate and formed a 3D meshwork by 2 weeks in vitro and express MAP2 and Tau-1 that label dendrites and axons, respectively (Supplementary Fig. S1b, c).

In agreement with data in the literature, we observed that multiple nestin- and GFAP-labeled cells migrated from the spheroids on poly-l-lysine-coated surfaces to form an adherent 2D cell monolayer (not shown). In contrast, spheroids preserved their 3D shape on alginate hydrogels, and only neurons extended neurite on the top of the soft hydrogel (Fig. 7a, b, e, f, i, j). Unlike neurons, GFAP-positive cells did not extend their processes outside spheroids neither on soft nor on stiff Ca-alginateNaCl hydrogels.

Since soft Ca-alginateNaCl hydrogels can be shaped to take any form or size and do not collapse during gentle mechanical treatment, they can be implanted in vivo by direct placing of a piece of hydrogel into a tissue lesion (e.g., lesion formed after traumatic spinal cord or brain injury). Biocompatible, soft Ca-alginateNaCl hydrogels promote neural growth and could be tested in the future as neural bridges in vivo.
Discussion

This study describes soft Ca-alginateNaCl hydrogels as stable 3D matrices with high adhesiveness to neural cells. Within a pH range that support dissociation of sodium alginate in aqueous solutions, fluctuations in the pH or ionic strength of the solution and drying of the alginates did not affect neural growth on/within the hydrogel, but gelation in the presence of NaCl enabled the generation of mechanically stable and soft 3D matrices with mechanical compliance close to brain tissue (≤1.5 kPa) that could be tested in the future as neural bridges in vivo.

Mechanical compliance of the substrate determines many aspects of cellular behavior, cell morphology, and gene expression.27,55 We previously demonstrated that soft, but not stiff alginate hydrogels were able to create an adhesive matrix for primary neurons. Now, we have shown that gelation in the presence of NaCl enabled the generation of mechanically stable and soft 3D matrices with mechanical compliance close to brain tissue (≤1.5 kPa) that could be tested in the future as neural bridges in vivo.52-54

Polymers behave differently on surfaces than in solutions.15,16 Nevertheless, our results obtained for hydrogels derived from surface-attached alginate layers are largely in agreement with the egg-box model, which was developed for alginate hydrogels in general. We observed that (1) the swelling profiles and stiffnesses of hydrogels fabricated in the presence or absence of NaCl were controlled by the concentration of Ca\(^{2+}\) and (2) high-viscosity G-enriched alginates required less Ca\(^{2+}\) to form a gel and eventually created larger networks than low-viscosity M-enriched alginates. Alginate concentrations directly influenced the hydrogel mass, but had no noticeable effects on the hydrogel swelling profile; therefore, manipulation of the amount of alginate in a layer could be used to fine tune the hydrogel's dimensions (i.e., to generate a thin matrix to culture neurons or a thick scaffold for implantation in vivo).

The acidic form of alginate is not water soluble, but its sodium salt is. In water, sodium alginate dissociates into negatively charged polymer chains, and mobile sodium ions that form a loosely bound counterion cloud around negative

![Discussion](https://example.com/fig7.jpg)
charges assure the polymer’s water solubility and cause a decrease in intrachain repulsion. Depending on the charge distribution along the chain, polymers could exist either in globular or persistent configurations. Subsequently, hydrogels formed by the association of globular polymer chains occupy more space, while hydrogels formed by persistent polymers are more compact. Additionally, slow gelation kinetics result in macroscopically homogeneous alginate gels. Thus, the macroscopic homogeneity and compactness of Ca-alginate hydrogels suggest that exogenous Na\(^+\) could enable slow gelation kinetics and optimize the spatial organization of an alginate network.

One Ca\(^{2+}\) ion binds up to four guluronic acid residues. In firmly connected, soft Ca-alginate hydrogels (e.g., in hydrogels derived from 1% LVG sol and 2 mM of CaCl\(_2\), where ~12% of the G residues have the potential to be crosslinked), the number of endogenous Na\(^+\) counterions are not sufficient to neutralize the negative charges on the polysaccharide chains, and repulsion between non-cross-linked sequences results in the formation of extremely swollen hydrogels. After immersion of such soft Ca-alginate hydrogels into physiological saline, exogenous Na\(^+\) ions penetrate into the hydrogel and, presumably, reduce intrachain repulsion to trigger global 3D changes in the entire network and leakage of Ca\(^{2+}\) ions from gelling sites, which lead to the eventual collapse of the hydrogel. In contrast, soft neurite-supportive Ca-alginate hydrogels did not collapse after infiltration of exogenous Na\(^+\) and maintained their mass and volume after immersion in buffered physiological saline, which makes them interesting for in vivo applications.

Polysaccharides (i.e., sugars) decorate proteins within the ECM and have been found to influence cell migration, axonal guidance, synapse development, and functioning. Most recently, wobble oligosaccharide motifs of chondroitin sulfate/dermatan sulfate proteoglycans had been demonstrated to induce signaling pathways that are essential for proliferation, self-renewal, and cell lineage commitment of neural stem cells. Although stiff alginate hydrogels were described as an inert matrix, alginate-derived oligosaccharides have been reported to costimulate growth factor-mediated proliferation, migration, and adhesion of endothelial cells, keratinocytes, and skin fibroblasts in vitro
cytokine production by RAW264.7 cells,
phagocytic activity of peritoneal monocytes.

The density of substrate-immobilized signaling molecules is crucial to initiate extra-cellular signaling events that lead to cell differentiation, adhesion, growth, and migration, and this must be considered during scaffold designing. In light of these reports, we hypothesize that firmly crosslinked alginate networks (and, therefore, soft hydrogels) could create a matrix with a high density of immobilized, adhesion-promoting carbohydrate sequences. Identification of the molecular mechanisms involved in neural growth on alginate matrices could provide valuable information for regenerative medicine and open new perspectives on using alginate polysaccharides for the functionalization of nonadhesive matrices.

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Disclosure Statement

No competing financial interests exist.

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SUPPLEMENTARY FIG. S1. Live images show a rat primary cortical neurite-bearing spheroid cultured for 48 h on (a) Ca-alginate$_{\text{NaCl}}$ hydrogel. Images in (a1, a2, a3) are enlarged frames of the corresponding images in (a). Neurite have visible growth cones (marked by *) on their tips. Spheroid-derived neurite continued to elongate and formed a (b, c) three-dimensional meshwork; soft hydrogel supports outgrowth of dendrites and axons. Fluorescence images show (b) MAP2 and (c) Tau-1 antibody-labeling of neurite-bearing spheroids and neurite meshwork on top of Ca-alginate$_{\text{NaCl}}$ hydrogels after 2 weeks in vitro. Ca-alginate$_{\text{NaCl}}$ hydrogels were generated at pH 7.4 by crosslinking of dry layers of LVM alginate with 2 mM of aqueous CaCl$_2$ in the presence of 150 mM of NaCl. Secondary antibodies were labeled with the fluorescence dye Alexa Fluor-488. The scale bar is 100µm.