

Multiple Sclerosis Journal

<http://msj.sagepub.com/>

Retinal ganglion cell and inner plexiform layer thinning in clinically isolated syndrome

Timm Oberwahrenbrock, Marius Ringelstein, Simon Jentschke, Katrin Deuschle, Katharina Klumbies, Judith Bellmann-Strobl, Jens Harmel, Klemens Ruprecht, Sven Schippling, Hans-Peter Hartung, Orhan Aktas, Alexander U Brandt and Friedemann Paul

Mult Scler 2013 19: 1887 originally published online 23 May 2013

DOI: 10.1177/1352458513489757

The online version of this article can be found at:

<http://msj.sagepub.com/content/19/14/1887>

Published by:



<http://www.sagepublications.com>

Additional services and information for *Multiple Sclerosis Journal* can be found at:

Open Access: Immediate free access via SAGE Choice

Email Alerts: <http://msj.sagepub.com/cgi/alerts>

Subscriptions: <http://msj.sagepub.com/subscriptions>

Reprints: <http://www.sagepub.com/journalsReprints.nav>

Permissions: <http://www.sagepub.com/journalsPermissions.nav>

>> [Version of Record](#) - Nov 25, 2013

[OnlineFirst Version of Record](#) - Nov 8, 2013

[OnlineFirst Version of Record](#) - May 23, 2013

[What is This?](#)

Retinal ganglion cell and inner plexiform layer thinning in clinically isolated syndrome

Multiple Sclerosis Journal
19(14) 1887–1895
© The Author(s) 2013
Reprints and permissions:
sagepub.co.uk/journalsPermissions.nav
DOI: 10.1177/1352458513489757
msj.sagepub.com



Timm Oberwahrenbrock^{1,*}, Marius Ringelstein^{2,*},
Simon Jentschke¹, Katrin Deuschle^{1,3}, Katharina Klumbies¹,
Judith Bellmann-Strobl^{1,3}, Jens Harmel², Klemens Ruprecht³,
Sven Schippling⁴, Hans-Peter Hartung², Orhan Aktas²,
Alexander U Brandt^{1,§} and Friedemann Paul^{1,3,§}

Abstract

Background: Axonal and neuronal damage are widely accepted as key events in the disease course of multiple sclerosis. However, it has been unclear to date at which stage in disease evolution neurodegeneration begins and whether neuronal damage can occur even in the absence of acute inflammatory attacks.

Objective: To characterize inner retinal layer changes in patients with clinically isolated syndrome (CIS).

Method: 45 patients with CIS and age- and sex-matched healthy controls were investigated using spectral domain optical coherence tomography. Patients' eyes were stratified into the following categories according to history of optic neuritis (ON): eyes with clinically-diagnosed ON (CIS-ON), eyes with suspected subclinical ON (CIS-SON) as indicated by a visual evoked potential latency of >115ms and eyes unaffected by ON (CIS-NON).

Results: CIS-NON eyes showed significant reduction of ganglion cell- and inner plexiform layer and a topography similar to that of CIS-ON eyes. Seven eyes were characterized as CIS-SON and likewise showed significant retinal layer thinning. The most pronounced thinning was present in CIS-ON eyes.

Conclusion: Our findings indicate that retinal pathology does occur already in CIS. Intraretinal layer segmentation may be an easily applicable, non-invasive method for early detection of retinal pathology in patients unaffected by ON.

Keywords

Clinically isolated syndrome, optical coherence tomography, retinal nerve fiber layer, retinal ganglion cell layer

Date received: 27th November 2012; revised: 8th April 2013; accepted: 11th April 2013

Introduction

Multiple sclerosis is an autoimmune disorder of the central nervous system that often manifests with optic neuritis (ON) as well as motor, sensory or cerebellar deficits in its earliest stage.¹ Current diagnostic criteria for MS require proof of dissemination of lesions or attacks in time and space.² In everyday clinical practice, patients presenting with a first clinical event that is highly indicative of MS are often instead diagnosed with a clinically isolated syndrome (CIS) or 'possible' MS.³ A confirmed diagnosis of MS is possible once additional attacks or lesions present, as is the case for a significant proportion of such patients.²

In light of this, pinpointing the aspects of CIS that are most predictive for subsequent diagnosis with MS has high

¹NeuroCure Clinical Research Center and Experimental and Clinical Research Center, Charité University Medicine Berlin and Max Delbrück Center for Molecular Medicine, Berlin, Germany

²Department of Neurology, Medical Faculty, Heinrich Heine University Düsseldorf, Düsseldorf, Germany

³Department of Neurology, Charité University Medicine Berlin, Berlin, Germany

⁴Department of Neuroimmunology and Clinical Multiple Sclerosis Research, Neurology Clinic, University Medical Center Zurich, Zurich, Switzerland

*Equally-contributing first authors in alphabetical order

§Equally-contributing senior authors in alphabetical order

Corresponding author:

Dr. Friedemann Paul, NeuroCure Clinical Research Center, Charité University Medicine Berlin, Charitéplatz 1, 10117 Berlin, Germany.

Email: friedemann.paul@charite.de

priority³ so that patients at risk can be identified. Diagnosing MS as early as possible and thus allowing for the widest range of therapeutic options, is therefore highly in the patients' interest, in particular as irreversible axonal and neuronal injury is a key aspect and correlate of disability in MS in early disease stages.³⁻⁵

One easily-accessible means of assessing neuroaxonal damage in MS is the investigation of the retina. Optical coherence tomography (OCT) has shown specific retinal alterations in MS patients:⁶ the retinal nerve fiber layer (RNFL) is reduced in MS,⁷ not only in eyes with a history of ON⁸ but also in eyes without any previous clinical event of ON.^{9,10} Additionally, microcystic macular edema (MME) in the inner nuclear layer (INL) has been reported in a subset of MS patients.¹¹ Although MME might not be specific to MS, but instead ON-dependent,¹² the INL has become a key focus of clinical investigation of MS pathology after a postmortem histopathology study reported neuronal loss in the INL.^{13,14}

Additionally, retinal changes in MS do not merely reflect the visual system, but potentially also overall disease pathology. RNFL thinning correlates closely with brain atrophy,¹⁵⁻¹⁷ and with reduction of N-acetyl-aspartate as marker of neuroaxonal integrity in the visual cortex.¹⁸

These findings suggest that the retina and, in particular, intraretinal layers may be an effective means of detecting subtle neuronal and axonal damage already present in CIS. To investigate this theory, we performed a cross-sectional study analysing intraretinal changes in CIS patients. We were especially interested in retinal pathology in eyes that had not suffered from previous ON and therefore applied a rigorous classification of eyes not only on clinical assessments but also visual evoked potentials (VEP).

Methods

Study participants

Patients were prospectively recruited from outpatient clinics at two university medical centers (Berlin and Düsseldorf). Inclusion criteria were clinical and paraclinical (MRI, CSF, EP) diagnosis of CIS suggestive of MS after relevant differential diagnoses had been ruled out, and an age between 18 and 65 years.² Patients received MRI to exclude the possibility that the disease had developed into MS since first diagnosis of CIS. Neurological disability was assessed according to the Expanded Disability Status Scale (EDSS).¹⁹ A history of ON was diagnosed by a treating physician and was cross-checked using medical records. Patients with a refractive error of more than ± 5.0 dioptres or with any history of eye disease that could impact OCT measurements (i.e. glaucoma) were excluded. A second exclusion criterion was steroid therapy within 30 days prior to examination. A group of healthy controls matched by age (± 3 years) and gender was recruited from patients' family members, medical staff or volunteers. Both centres assessed the matched controls to their patients. To exclude potential

centre effects, we additionally performed centre-specific analysis or included centre as covariate. In these analyses, centre did not have a significant effect (data not shown). Local ethics committees approved the study and all participants gave written informed consent.

Visual evoked potentials

VEP were either performed during the clinical work-up or as part of the study protocol prior to or on the same day as the OCT assessment. We used the P100 latency values as a parameter to prove optic nerve conduction slowing potentially related to a history of ON. VEP amplitude was not analysed because the two centres involved in the study performed VEP using different devices in a non-standardized manner.

Optical coherence tomography

Experienced operators performed OCT on un-dilated eyes using Heidelberg Spectralis SD-OCT (Heidelberg Engineering, Germany). All scans were checked for appropriate image quality. All participants were examined using the peripapillary ring scan, which measures RNFL thickness (pRNFL) around the optic nerve head in a circle with an angle of 12° , resulting in a diameter of 3.4 mm (example shown in Figure 1(a)). Macular volume was assessed by a custom scan comprising 61 vertical B-scans (each with 768 A-Scans, Automatic Real-Time (ART) = 13 frames) with a scanning angle of $30^\circ \times 25^\circ$ focusing on the fovea. Using this scan, TMV and intra-retinal layers thicknesses were determined within a cylinder of 6 mm diameter (Figure 1(b)).

Intraretinal layer segmentation

Heidelberg Engineering provided beta software that employed a multilayer segmentation algorithm for macular volume scans. To analyse the inner retinal layers, a subset of B-scans were segmented and manually corrected by an experienced assessor in a blinded fashion. The multilayer analysis was performed on the central B-scan through the fovea and on six B-scans each in nasal and temporal direction. Manual correction of automatically segmented B-scans is a time-consuming step. As a compromise, we manually corrected every fourth B-scan, thus analysing an area largely covering the 6 mm diameter ETDRS grid with a distance between adjacent B-scans of approximately 500 μm . For the combined analysis of both eyes, thickness maps of the left eye were mirrored vertically to match the topology of the right eye. The mean thickness maps within each of the study groups were calculated for the four innermost retinal layers: macular RNFL (mRNFL), ganglion cell layer (GCL), inner plexiform layer (IPL) and INL (Figure 1(c)). Because differentiating between GCL and IPL proved to be a hurdle, we used the combined thickness of GCL and IPL (GCIPL). Please see the supplementary data for individual analyses of GCL and IPL. By subtracting the group-specific mean

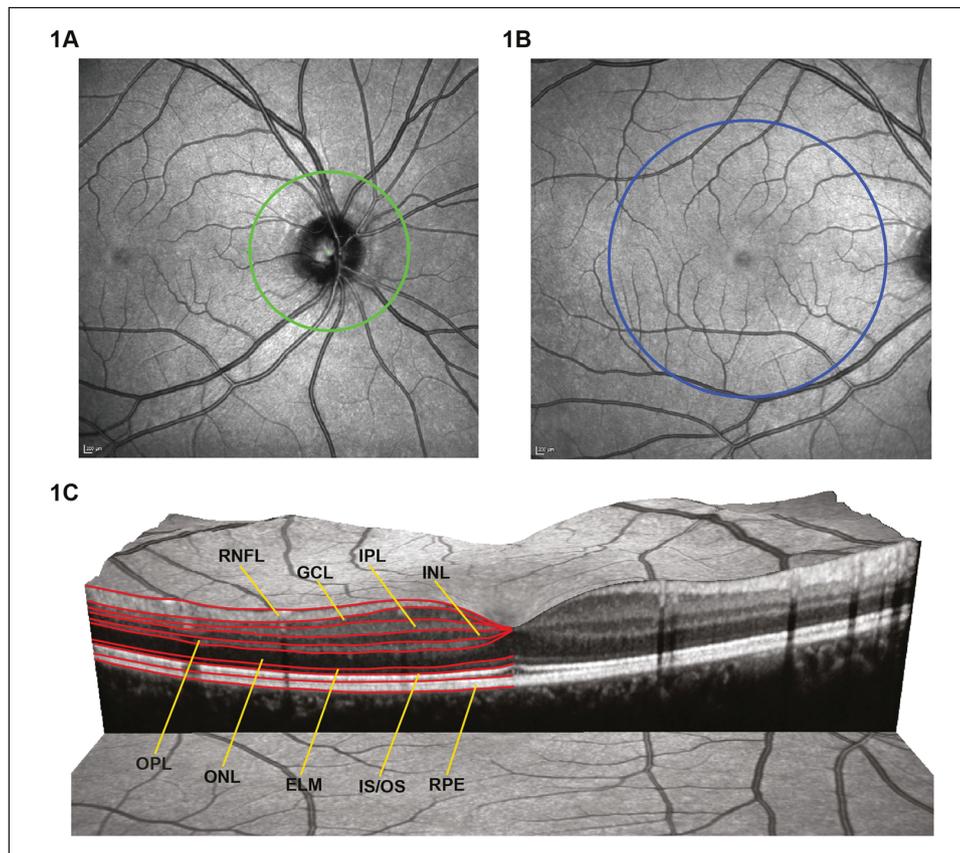


Figure 1. Examples of regions analysed in OCT.

A) Scanning laser ophthalmoscopy image showing the region of the peripapillary ring scan (green); B) Scanning laser ophthalmoscopy image of the macular scan with the blue circle indicating the area for total macular volume and intraretinal layer thickness determination; C) 3D reconstruction of a macular volume scan, depicting the identified intraretinal layers.

Abbreviations: RNFL = retinal nerve fibre layer; GCL = ganglion cell layer; IPL = inner plexiform layer; INL = inner nuclear layer; OPL = outer plexiform layer; ONL = outer nuclear layer; ELM = external limiting membrane; IS/OS = inner segments / outer segments; RPE = retinal pigment epithelium.

thickness maps we produced spatial difference maps (Figure 3), in which negative values indicate a thinning of the patients' group compared to matched healthy controls, whereas positive values indicated thickening.

Statistical analysis

Generalized estimation equation models (GEE) accounting for within-subject inter-eye effects were used to compare OCT results between the study cohorts. For the subgroup analysis, only controls that were matched to the respective CIS patients' eyes (NON, SON, ON) were used. Correlations between VEP and OCT results were performed by linear regression. All statistical analyses were performed and all figures were created using R version 2.15.0. Statistical significance was established at $p < 0.05$.

Results

Study participants

In total, 45 patients (Berlin 29, Düsseldorf 16) were enrolled and compared to matched healthy controls (Berlin 29,

Düsseldorf 16). All patients were diagnosed with CIS at the time of OCT examination and diagnosis and non-progression towards MS was confirmed by means of MRI. Of the patients, 16 had unilateral optic neuritis (seven on the right, 10 on the left) and 14 patients presented with spinal cord symptoms. Six patients experienced relapses with findings suggestive of infratentorial brain lesions, in seven patients supratentorial signs were found, and one patient exhibited both supratentorial and spinal clinical signs. Examination of one patient's eye did not pass the quality criteria due to image artefacts and was excluded. Demographic and clinical data are summarized in Table 1.

ON classification according to VEP latency and correlation to standard OCT results

As a clinical diagnosis of ON may have been missed by patients or physicians, we created another category of subclinical (or suspected) ON in eyes without a clinical ON history, as assessed by VEP. In addition to the group of confirmed ON eyes (CIS-ON), we defined a group of suspected ON eyes (CIS-SON), defined as eyes with

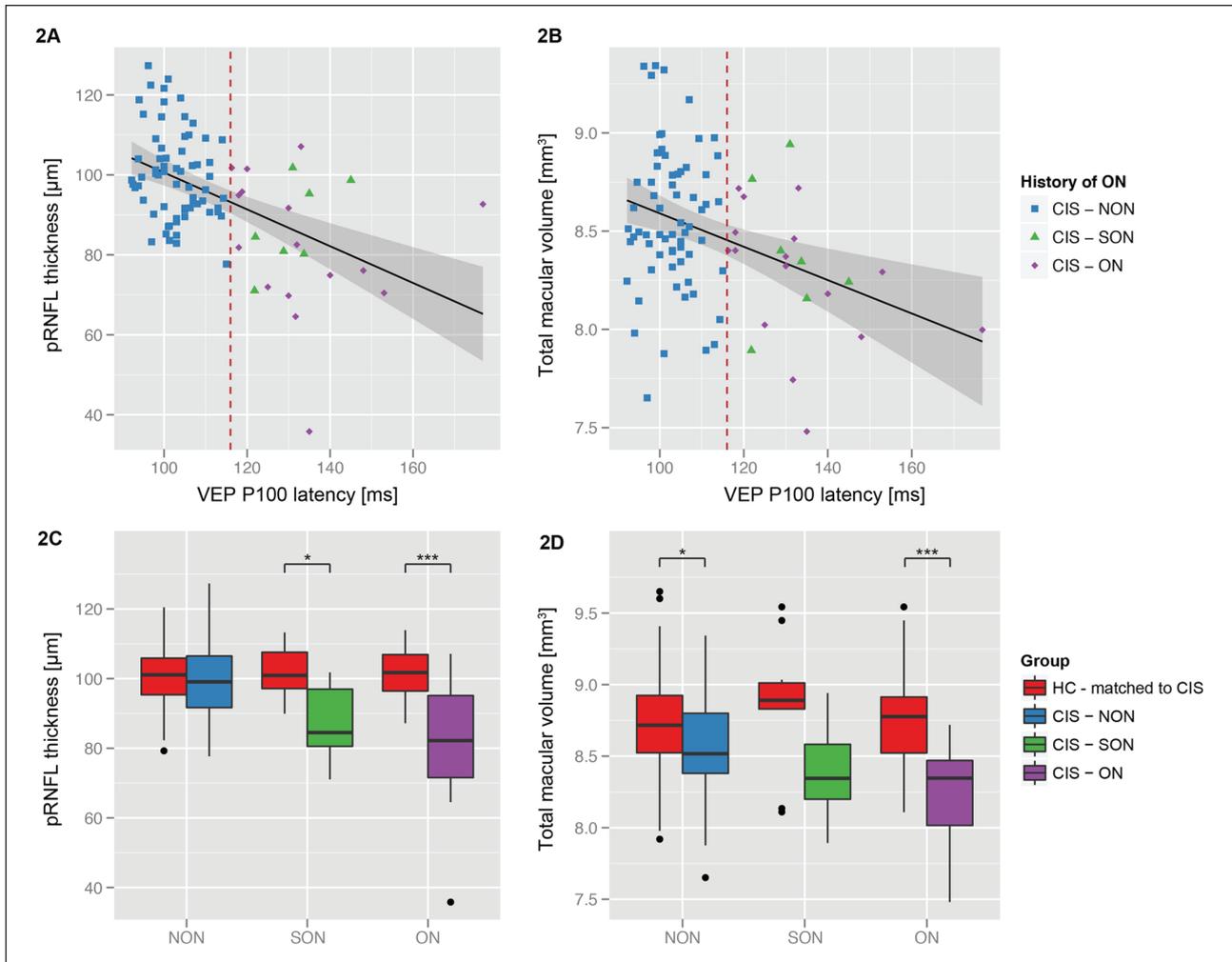


Figure 2. VEP and standard OCT results.

Scatterplots showing the relationship of the VEP P100 latencies with A) peripapillary RNFL (pRNFL) and B) total macular volume. The red dashed line at 115 ms indicates the threshold between CIS-NON and CIS-SON eyes. The black line is the result of the linear regression including all CIS eyes with the standard error given as gray shadow. Comparison of C) peripapillary RNFL thickness and D) total macular volume between the different CIS groups and the matching controls. Significant differences are marked with * ($p < 0.05$), ** ($p < 0.01$) and *** ($p < 0.001$). Abbreviations: HC = healthy control eyes; CIS-NON = patient eyes without history of optic neuritis and VEP P100 ≤ 115 ms; CIS-SON = eyes with VEP P100 latency > 115 ms but no ON diagnosis; CIS-ON = patient eyes with clinical ON diagnosis.

prolonged P100 latency of over 115 ms but, as stated above, without a clinical history of ON. The latter value of a 115 ms limit for normal eyes is in accordance with literature²⁰ and proved an effective means of distinguishing between eyes diagnosed with ON and unaffected eyes (Figure 2(a) and 2(b)). In total, seven eyes were classified as CIS-SON. Both eyes of two patients were classified as suspected ON and all other CIS-SON eyes were contralateral to CIS-ON eyes. Figure 2(a) shows the correlation between P100 latencies and pRNFL thickness, while Figure 2(b) is a graph of the relationship between the TMV and the VEP results. Linear regression showed significant correlation between pRNFL and P100 VEP latencies in all CIS eyes ($R^2 = 0.243$, $p < 0.001$) and in CIS-NON eyes ($R^2 = 0.065$, p

$= 0.039$) but not in CIS-SON and CIS-ON eyes. Similarly, TMV correlated significantly to P100 latencies for all CIS eyes ($R^2 = 0.124$, $p < 0.001$), but not for the other subgroups.

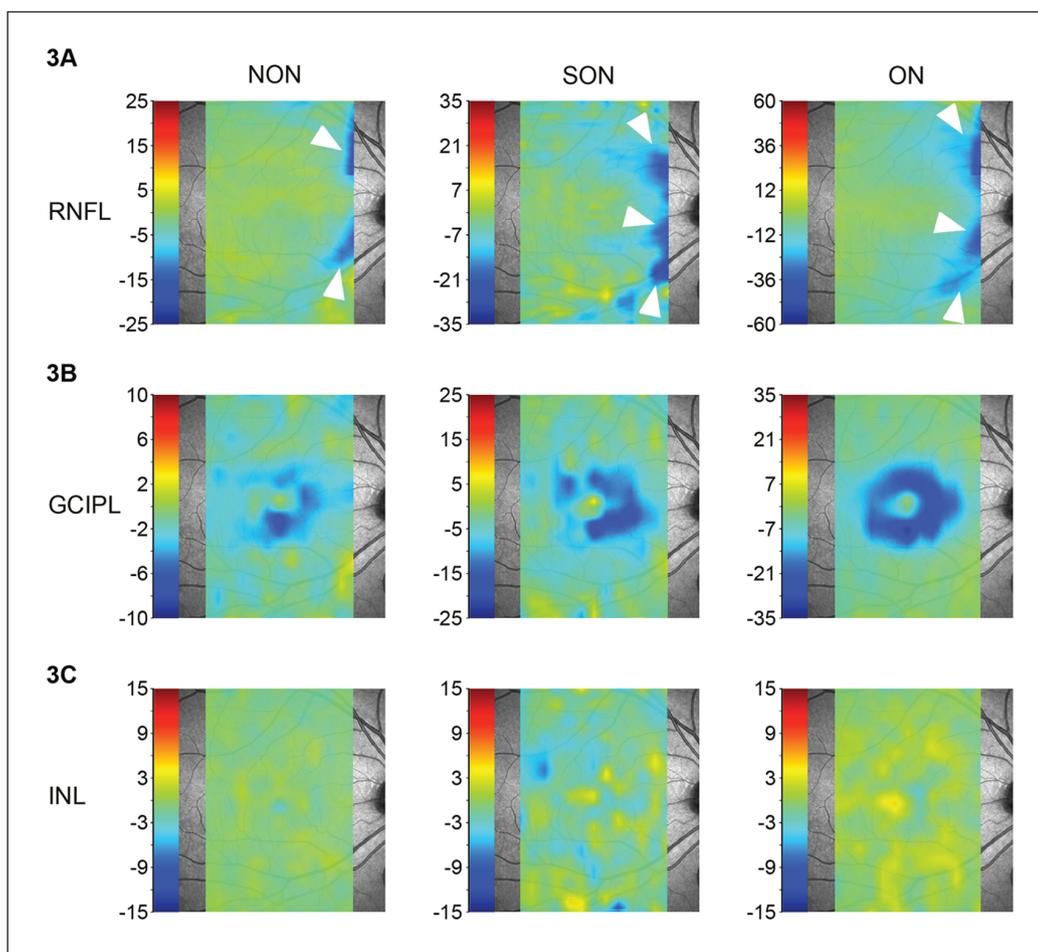
pRNFL and TMV comparison

When compared to the corresponding age- and sex-matched controls, pRNFL thickness was reduced in CIS-ON ($p < 0.001$) and CIS-SON ($p = 0.014$) but not in CIS-NON eyes ($p = 0.636$) (Figure 2(c)). Analysis of macular scans revealed significant TMV reduction in CIS-ON eyes ($p < 0.001$) and, importantly, also in CIS-NON eyes ($p = 0.031$) versus controls (Figure 2(d)). TMV reduction in the 7 CIS-SON eyes was not significant.

Table 1. Demographical and clinical parameters.

		CIS	HC
Subjects	N	45	45
Eyes	N total	89	90
	N with diagnosed ON	16	NA
	N with suspicion ON	7	NA
Age (years)	Mean (SD)	31.92 (7.95)	31.67 (7.80)
	Min–Max	19.13–56.25	18.64–54.20
Gender	N female (%)	31 (68.89)	31 (68.89)
First symptom (months)	Mean time between first symptom and OCT (SD)	8.60 (12.17)	NA
	Min–Max	1.37–59.70	
	Median	1	NA
EDSS	Median	1	NA
	Min–Max	0–4	NA

HC: healthy control; CIS: clinically isolated syndrome; ON: optic neuritis; SD: standard deviation; OCT: optical coherence tomography; Min: minimum; Max: maximum, NA: not applicable.

**Figure 3.** Spatial analysis of changes in CIS eyes versus healthy control eyes.

A) Changes in RNFL thickness between CIS patients and the corresponding group of age- and sex-matched healthy controls. Patients were stratified by history of ON: no history of ON (NON), suspected ON (SON) or clinically-diagnosed ON. Reduction in RNFL thickness was evident near the optic nerve head (white arrows) in all groups but was more pronounced in SON and ON eyes. B) Thickness changes in the GCIPL were identified in the perimacular region and were most evident in CIS-ON eyes. Significant thinning of the GCIPL in CIS-NON eyes compared to the matching controls were found in the perimacular area ($p = 0.027$). C) No group showed significant changes in the INL.

Abbreviations: CIS = clinically isolated syndrome; RNFL = retinal nerve fibre layer; GCIPL = combined ganglion cell and inner plexiform layer; INL = inner nuclear layer.

Table 2. Mean (SD) retinal layer thickness and total macular volume results.

Retinal layer	HC (matched to CIS-NON)	CIS-NON	Regression coefficient ^a	Standard error ^a	P value ^a
pRNFL (μm)	100.69 (8.01)	99.94 (11.28)	-1.01	2.13	0.636
TMV (mm ³)	8.724 (0.321)	8.570 (0.362)	-0.16	0.07	0.031
mRNFL (μm)	39.73 (4.45)	38.76 (4.32)	-1.30	0.96	0.173
GCIPL (μm)	71.27 (4.52)	68.88 (5.52)	-2.48	1.12	0.027
INL (μm)	33.76 (2.19)	33.39 (2.01)	-0.22	0.46	0.626
Retinal layer	HC (matched to CIS-SON)	CIS-SON	Regression coefficient ^a	Standard error ^a	P value ^a
pRNFL (μm)	101.75 (8.25)	87.49 (11.29)	-14.68	5.97	0.014
TMV (mm ³)	8.866 (0.465)	8.392 (0.358)	-0.44	0.26	0.091
mRNFL (μm)	42.01 (4.52)	37.31 (4.56)	-4.70	2.62	0.073
GCIPL (μm)	71.45 (4.87)	63.15 (7.43)	-7.62	3.85	0.048
INL (μm)	34.49 (2.42)	32.99 (0.96)	-1.31	1.05	0.212
Retinal layer	HC (matched to CIS-ON)	CIS-ON	Regression coefficient ^a	Standard error ^a	P value ^a
pRNFL (μm)	101.36 (7.44)	82.08 (18.02)	-20.15	4.62	<0.001
TMV (mm ³)	8.746 (0.335)	8.265 (0.350)	-0.48	0.11	<0.001
mRNFL (μm)	39.89 (4.57)	32.14 (5.64)	-8.05	1.68	<0.001
GCIPL (μm)	71.57 (4.62)	58.69 (9.77)	-3.68	2.64	<0.001
INL (μm)	34.34 (2.38)	34.86 (2.17)	0.64	0.75	0.397

HC: healthy control eyes; CIS-NON: patient eyes without history of optic neuritis and VEP P100 ≤ 115 ms; CIS-SON: eyes with VEP P100 latency > 115 ms but no ON diagnosis; CIS-ON: patient eyes with clinical ON diagnosis; SD: standard deviation; pRNFL: peripapillary retinal nerve fiber layer; TMV: total macular volume; mRNFL: macular retinal nerve fiber layer; GCIPL: combined ganglion cell and inner plexiform layer; INL: inner nuclear layer.

^aStatistical parameters of the comparison of CIS patients to the corresponding matching controls using generalized estimation equation models.

Intraretinal multilayer segmentation

The mean macular thickness values for inner retinal layers (mRNFL, GCIPL, INL) of the different groups are summarized in Table 2. A graphical representation of the spatial changes of CIS patients compared to the matching controls is given in Figure 3.

Analysis of the central macular area (6 mm in diameter around the fovea) showed significant reduction in mRNFL thickness in CIS-ON eyes, but not for CIS-SON and CIS-NON in comparison to matched controls (Table 2). Spatial difference maps showed that mRNFL thinning was most prominent in close proximity to the optic nerve head (Figure 3(a), white arrows). Here, even for CIS-NON eyes mRNFL thinning was visible very close to the optic nerve head. It should be noted that macular volume scans are not designed to investigate the papillary region and that this area is highly penetrated by blood vessels, potentially causing segmentation errors; thus, the mRNFL results have to be evaluated with caution.

All patient groups showed reduced GCIPL thickness compared to the matched healthy controls. Spatial differences of the GCIPL were found in the perimacular region (Figure 3(b)) and statistical analysis of the GCIPL confirmed that the thickness in this area was significantly reduced for all patient groups compared to controls (Table 2). The thinning in CIS-ON and CIS-SON eyes was more pronounced than in the CIS-NON group, while the spatial distribution of changes was similar. Please refer to the supplementary material for detailed data on the analysis of the GCL and IPL individually.

Analogous to pRNFL and TMV, we analysed a potential correlation between VEP latencies and intraretinal layer thicknesses: mRNFL ($R^2 = 0.203$, $p < 0.001$) and GCIPL ($R^2 = 0.315$, $p < 0.001$) were significantly correlated to VEP latencies (supplementary Figure 2). There was no correlation of intraretinal layer thicknesses or VEP latencies with symptom onset in the CIS-NON group (supplementary Figure 3).

Discussion

We analysed intraretinal changes in a cohort of CIS patients, which included both eyes with confirmed previous ON, eyes with suspected ON, and eyes without evidence of ON compared to age- and sex-matched healthy controls. Notably, we identified significant thinning of GCIPL in the eyes of CIS patients without any clinical history of ON or suspected previous subclinical ON as determined by VEP changes. A supplementary analysis using distinct GCL and IPL thicknesses localized this GCIPL thinning to the GCL in CIS-NON patients. Additionally, and as expected, eyes with a confirmed history of ON showed an even more pronounced thinning of retinal layers. In contrast, INL appeared unaltered. Our data indicate that retinal neuronal damage can accompany CIS independently of a prior history of ON.

Three previous studies have investigated retinal changes in CIS patients: The first study failed to detect pRNFL or TMV reduction in the eyes of CIS patients without prior ON.²¹ A second study reported no retinal damage in the eyes of patients with isolated unilateral ON.²² However,

these studies were conducted before the introduction of spectral-domain OCT (SD-OCT), the superior spatial resolution of which over time-domain OCT (TD-OCT)²³ allows for the investigation of intraretinal layers.²⁴ Previously and in particular, in the above studies, retinal alterations may have simply not been detectable by TD-OCT and, more importantly, GCIPL changes that can only be quantified using SD-OCT might be superior for detecting even subtle neurodegeneration in CIS over pRNFL. Peripapillary RNFL also failed to detect differences in our groups, suggesting that this parameter is in general less sensitive for detecting MS pathology than new intraretinal layer measurements like GCIPL. With this in mind, the failure to detect significant pRNFL alterations in our CIS-NON cohort may simply be a power issue. A third recent study comprising 45 CIS patients showed a reduction of pRNFL but not TMV using SD-OCT.²⁵

The present study is the first to investigate intraretinal layer changes or detect retinal neurodegeneration independent from ON in a larger cohort of CIS patients. A recent study that reported reduction of the GCIPL in MS patients with and without a history of ON included seven CIS patients while the remaining patients had long-standing diagnoses of MS, which precluded reliable assessment of retinal damage in early disease stages.²⁶ Other studies have shown INL impairment (i.e. microcystic macular oedema) in MS patients with longer disease duration.^{11,14} Such changes were not detected in our CIS patients, suggesting that INL impairment might be a symptom of later or more severe disease stages.

Our finding that damage to the GCIPL is detectable in CIS eyes without clinical history of ON and with normal VEP latency lends additional support to the increasingly widespread understanding of MS as both a demyelinating and neurodegenerative disease.²⁷ We show that neurodegeneration is not, in fact, limited to advanced disease stages, in which it is considered responsible for the continuous progression of neurological disability, even in the absence of relapses. Instead, neurodegeneration can begin very early in disease development. Our data corroborate MRI data showing neuroaxonal damage during the very earliest MS stages,^{4,28} as well as histopathology data from brain²⁹ and eye,¹³ and from experimental autoimmune encephalomyelitis.^{30,31} In line with previous investigations, our study provides evidence that inflammatory attacks to the optic nerve to the extent of a clinical or subclinical ON may not be a pre-requisite for damage to the retinal GCIPL.²⁶

Our finding that neuronal retinal damage begins during very early disease stages raises urgent questions, the answers to which may challenge our understanding of the underlying pathology and mechanisms of MS.³² Is the damage we found in the retina a consequence of the retrograde degeneration of retinal nerve fibres that occurs as a consequence of autoimmune brain inflammation in MS? If the answer is yes, it follows that retrograde RNFL damage

would subsequently initiate a degenerative process in the GCL via a *dying back* mechanism. Indeed, the hypothesis that retrograde retinal neuroaxonal damage takes place both after ON as well as brain inflammation without clinical ON is supported by experimental animal data from intracranial optic nerve sections.³³ Here, ocular pathology was shown to be limited to the inner retina. Evidence for inner retinal layer damage has been further provided by the first large scale pathological description of retinæ from autopsied MS patients showing – apart from the anticipated extensive axonal damage – neuronal loss in both the GCL and the INL.¹³ In contrast, a recent OCT study has suggested a primary retinal pathology as a novel distinct subtype of MS, which would implicate that a *dying back* pathomechanism does not apply to all patients:²⁴ the study identified MS patients exhibiting substantial reduction of TMV and significant thinning of the outer and inner nuclear layers despite normal RNFL values. The authors suggested that retinal pathology in this disease subtype (termed ‘macular thinning predominant’) occurs independently of optic nerve pathology and may be a harbinger of a more aggressive disease course. However, these findings have yet to be confirmed by other groups and with other OCT devices in larger cohorts.³⁴

Some important caveats of our study should be noted. Firstly, undetected subclinical ON episodes in our patient cohort may have skewed our results. However, we dealt with this potential cohort bias swiftly by conducting a thorough clinical assessment and examination of the individual patients. Additionally, each patient had to undergo VEP: Eyes with P100 latency suspicious for ON were classified as subclinical ON and not as unaffected eyes. Furthermore, all patients received MRI as proof that a confirmed diagnosis of MS could not yet be established. Although this approach cannot be guaranteed to prevent all errors in ON identification, it does ensure that the risk of misclassification as CIS-NON or MS is negligible and that the conclusions drawn from our data are valid.

A further limitation of our study is that we could not correlate morphological data to functional visual measures such as low contrast letter acuity. However, we are currently addressing this aspect in an ongoing CIS study that includes Sloan charts as suggested by a previous study.³⁵ The high number of statistical analyses in comparison to the relatively low number of patients should also be noted. As we did not perform a previous power calculation and since OCT parameters are related and thus likely correlated, we did not correct for multiple comparisons, since doing so would have likely caused an overcorrection. We did carefully examine our cohorts for a possible influence of outliers and distribution effects, finding no such effect. However, it is important to reproduce our findings in an independent cohort.

Segmentation of intraretinal layers is a novel technique and no studies have been performed so far to better

understand how segmentation-derived results relate to in-vivo morphological changes that appear in MS (e.g. through histopathological studies). However, a number of recent studies have successfully applied intraretinal segmentation,^{9,14,17,26,36} and comparison of different segmentation techniques showed excellent reproducibility and reliability.³⁷ We have investigated reliability of the novel algorithm applied in this study in a cross-centre inter-rater reliability study on a defined set of OCT macular B-scans. Results support the excellent reliability of intraretinal segmentation reported by others,³⁷ with the exception that no histopathological correlation has been performed so far (publication in preparation). However, GCL and IPL are still difficult to differentiate in OCT scans and therefore we based our study results mostly on the combined layer of both (GCIPL) and present individual layer analyses as supplementary data only.

Of note is the large amount of eyes that were classified as suspected ON ($n = 7$) in comparison to the number of eyes with definite clinical ON ($n = 16$). Retinal layer-thinning in these eyes was in-between NON and ON eyes, further supporting the notion that optic nerve inflammation is not a *yes* or *no* event. Instead, substantial damage might be caused by optic nerve inflammation before clinical visibility in form of an apparent clinical ON might be established. As our cohort comprised only patients with CIS, failure to detect subclinical ON potentially might compromise the discrimination between CIS patients and patients who already have definite MS. Clearly, detection of subclinical alterations in visual and other functional systems urgently needs improvement. Our study did not investigate the discriminatory properties of OCT and VEP between CIS and MS patients, and consequently, this question must be addressed by a future study.

In summary, our study shows that retinal neurodegeneration is already detectable in CIS patients and is dependent but importantly also independent of clinical relapses (i.e. ON). Accordingly, irreversible neuronal damage in MS might be much more prevalent than previously thought. Long-term follow-up of our study patients, who exhibited very early substantial and presumably irreversible neuroaxonal damage, is vital to ascertain diagnosis in patients likely to develop MS as early as possible.

Conflict of interest statement

The authors declare that there is no conflict of interest.

Funding

This study was supported by grants from the German Research Foundation (DFG Exc 257) and the German Federal Ministry of Economics and Technology (BMW ZIM KF2286101FO9). The MS center at the Department of Neurology, Heinrich-Heine-Universität Düsseldorf, is supported in part by the Walter-and-Ilse-Rose-Stiftung (to O.A. and H.-P.H.), the Eugène Devic European Network (E-rare/EU-FP7; to O.A. and H.-P.H.), and the German Ministry for Education and Research (BMBF, 'German

Competence Network Multiple Sclerosis', KKNMS-BMBF; to H.-P.H.). The funding bodies neither influenced the study design, data collection and analysis, nor the decision to publish, and preparation of the manuscript.

References

1. Compston A and Coles A. Multiple sclerosis. *Lancet* 2008; 372: 1502–1517.
2. Polman CH, Reingold SC, Banwell B, et al. Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. *Ann Neurol* 2011; 69: 292–302.
3. Miller DH, Chard DT and Ciccarelli O. Clinically isolated syndromes. *Lancet Neurol* 2012; 11: 157–169.
4. Filippi M, Bozzali M, Rovaris M, et al. Evidence for widespread axonal damage at the earliest clinical stage of multiple sclerosis. *Brain*. 2003;126: 433–437.
5. Rovaris M, Gambini A, Gallo A, et al. Axonal injury in early multiple sclerosis is irreversible and independent of the short-term disease evolution. *Neurology* 2005; 65: 1626–1630.
6. Frohman E, Costello F, Zivadinov R, et al. Optical coherence tomography in multiple sclerosis. *Lancet Neurol* 2006; 5: 853–863.
7. Petzold A, De Boer JF, Schippling S, et al. Optical coherence tomography in multiple sclerosis: a systematic review and meta-analysis. *Lancet Neurol* 2010; 9: 921–932.
8. Costello F, Coupland S, Hodge W, et al. Quantifying axonal loss after optic neuritis with optical coherence tomography. *Ann Neurol* 2006; 59: 963–969.
9. Albrecht P, Ringelstein M, Müller AK, et al. Degeneration of retinal layers in multiple sclerosis subtypes quantified by optical coherence tomography. *Mult Scler* 2012; 18: 1422–1429.
10. Oberwahrenbrock T, Schippling S, Ringelstein M, et al. Retinal Damage in Multiple Sclerosis Disease Subtypes Measured by High-Resolution Optical Coherence Tomography. *Multiple Sclerosis International* 2012; 2012: 1–10.
11. Gelfand JM, Nolan R, Schwartz DM, et al. Microcystic macular oedema in multiple sclerosis is associated with disease severity. *Brain* 2012; 135: 1786–1793.
12. Balk LJ, Killestein J, Polman CH, et al. Microcystic macular oedema confirmed, but not specific for multiple sclerosis. *Brain* 2012; 135: e226; author reply: e227. doi:10.1093/brain/aw216
13. Green AJ, McQuaid S, Hauser SL, et al. Ocular pathology in multiple sclerosis: retinal atrophy and inflammation irrespective of disease duration. *Brain* 2010; 133: 1591–1601.
14. Saidha S, Sotirchos ES, Ibrahim MA, et al. Microcystic macular oedema, thickness of the inner nuclear layer of the retina, and disease characteristics in multiple sclerosis: a retrospective study. *Lancet Neurol* 2012; 11: 963–972.
15. Gordon-Lipkin E, Chodkowski B, Reich DS, et al. Retinal nerve fiber layer is associated with brain atrophy in multiple sclerosis. *Neurology* 2007; 69: 1603–1609.
16. Dörr J, Wernecke KD, Bock M, et al. Association of retinal and macular damage with brain atrophy in multiple sclerosis. *PLoS ONE* 2011; 6: e18132.
17. Zimmermann H, Freing A, Kaufhold F, et al. Optic neuritis interferes with optical coherence tomography and magnetic resonance imaging correlations. *Mult Scler* 2013;19(4): 443–50.

18. Pfueller CF, Brandt AU, Schubert F, et al. Metabolic changes in the Visual cortex are linked to retinal nerve fiber layer thinning in multiple sclerosis. *PLoS ONE* 2011; 6: e18019.
19. Kurtzke JF. Rating neurologic impairment in multiple sclerosis: an expanded disability status scale (EDSS). *Neurology* 1983; 33: 1444–1452.
20. Sisto D, Trojano M, Vetrugno M, et al. Subclinical visual involvement in multiple sclerosis: a study by MRI, VEPs, Frequency-Doubling Perimetry, Standard Perimetry, and Contrast Sensitivity. *IOVS* 2005; 46: 1264–1268.
21. Outteryck O, Zephir H, Defoort S, et al. Optical coherence tomography in clinically isolated syndrome: no evidence of subclinical retinal axonal loss. *Arch Neurol* 2009; 66: 1373–1377.
22. Kallenbach K, Sander B, Tsakiri A, et al. Neither retinal nor brain atrophy can be shown in patients with isolated unilateral optic neuritis at the time of presentation. *Mult Scler* 2011; 17: 89–95.
23. Bock M, Brandt AU, Dorr J, et al. Time domain and spectral domain optical coherence tomography in multiple sclerosis: a comparative cross-sectional study. *Mult Scler* 2010; 16: 893–896.
24. Saidha S, Syc SB, Ibrahim MA, et al. Primary retinal pathology in multiple sclerosis as detected by optical coherence tomography. *Brain* 2011; 134: 518–533.
25. Gelfand JM, Goodin DS, Boscardin WJ, et al. Retinal axonal loss begins early in the course of multiple sclerosis and is similar between progressive phenotypes. *PLoS ONE* 2012; 7: e36847.
26. Syc SB, Saidha S, Newsome SD, et al. Optical coherence tomography segmentation reveals ganglion cell layer pathology after optic neuritis. *Brain* 2012; 135: 521–533.
27. Zipp F and Aktas O. The brain as a target of inflammation: common pathways link inflammatory and neurodegenerative diseases. *Trends Neurosci* 2006; 29: 518–527.
28. Sbardella E, Tomassini V, Stromillo ML, et al. Pronounced focal and diffuse brain damage predicts short-term disease evolution in patients with clinically isolated syndrome suggestive of multiple sclerosis. *Mult Scler* 2011; 17: 1432–1440.
29. Lucchinetti CF, Popescu BFG, Bunyan RF, et al. Inflammatory cortical demyelination in early multiple sclerosis. *N Engl J Med* 2011; 365: 2188–2197.
30. Vogt J, Paul F, Aktas O, et al. Lower motor neuron loss in multiple sclerosis and experimental autoimmune encephalomyelitis. *Ann Neurol* 2009; 66: 310–322.
31. Fairless R, Williams SK, Hoffmann DB, et al. Preclinical retinal neurodegeneration in a model of multiple sclerosis. *J Neurosci* 2012; 32: 5585–5597.
32. Trapp BD, Peterson J, Ransohoff RM, et al. Axonal transection in the lesions of multiple sclerosis. *N Engl J Med* 1998; 338: 278–285.
33. Holländer H, Bisti S, Maffei L, et al. Electroretinographic responses and retrograde changes of retinal morphology after intracranial optic nerve section. A quantitative analysis in the cat. *Exp Brain Res* 1984; 55: 483–493.
34. Brandt AU, Oberwahrenbrock T, Ringelstein M, et al. Primary retinal pathology in multiple sclerosis as detected by optical coherence tomography. *Brain* 2011; 134: e193; author reply: e194.
35. Balcer LJ, Baier ML, Cohen JA, et al. Contrast letter acuity as a visual component for the Multiple Sclerosis Functional Composite. *Neurology* 2003; 61: 1367–1373.
36. Saidha S, Syc SB, Durbin MK, et al. Visual dysfunction in multiple sclerosis correlates better with optical coherence tomography derived estimates of macular ganglion cell layer thickness than peripapillary retinal nerve fiber layer thickness. *Mult Scler* 2011; 17: 1449–1463.
37. Seigo M, Sotirchos E, Newsome S, et al. In vivo assessment of retinal neuronal layers in multiple sclerosis with manual and automated optical coherence tomography segmentation techniques. *J Neurol* 2012; 259: 2119–2130.